

Cruise Report for USCG Cutter Healy Cruise 0803 (HLY0803)

July 3 - July 31, 2008, Dutch Harbor to Dutch Harbor, AK

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Healy Captain: Captain Frederick Sommer USCG



Acknowledgements

USGC Healy cruise 0803 was a complex and ambitious undertaking. In addition to the large number of scientists and educators involved (55), the scientific goals were interdisciplinary and therefore logistically complex. The cruise included the deployment of moorings and sediment traps, over two hundred CTD casts, many net hauls and extensive on-deck work that took place around the clock. In the midst of all of this sampling work, there also were several educational outreach activities, both from the ship and in the form of visits by scientists and educators to St. George Island. The success of these many aspects of the cruise reflects the high level of expertise and cooperation of the officers and crew of the Healy. Although the science party gratefully acknowledges the help of all of the Coast Guard personnel on-board, the cooperation and expertise of Captain Fred Sommer and his officers was outstanding. The scientists were uniformly appreciative of the work of Executive Officer Dale Bateman, Operations Officer Jeff Stewart and Science Officer Silas Ayers. MSTC Mark Reig and his crew of marine science technicians (Chuck Bartlett, Rich Layman, Tom Kruger and Tiffany Wright) worked tirelessly during the cruise and made the scientific work possible. All science results from this cruise owe a large debt to this valued cooperation.

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Cruise Summary

This report summarizes the cruise of the USCG cutter Healy from July 5 through July 31, 2008 (HLY0803). This was the third NSF funded, dedicated cruise for the Bering Ecosystem Study (BEST) project that focuses on the impact of sea ice on the marine ecology of the region. In particular it focused on pathways of nutrients and organic matter that lead to the abundant upper trophic levels and valuable fisheries on this extensive, high latitude continental shelf. The first two BEST cruises took place in April-May of 2007 and 2008 and focused on the conditions directly associated with the retreating ice edge. The HLY0803 cruise was the first BEST cruise to characterize summer conditions on the eastern shelf, including the seasonal evolution of the nutrient and phytoplankton fields, as well as the distribution and abundance of zooplankton and ichthyoplankton. The cruise covered most of the eastern Bering Sea shelf from the Aleutian Islands to St. Lawrence Island. A complex, multidisciplinary sampling plan was carried out that included continuous measurements of meteorological and surface water characteristics with autonomous sensors on the ship as well as bird and marine mammal observations along the over 4000 nautical miles of cruise track. In addition, 177 unique stations were occupied during the cruise. Sampling activities at these stations included 210 CTD casts, 52 bio-optics casts, 22 productivity casts, 59 MOCNESS tows, 83 CalVet tows, 25 bongo tows, 37 multicore drops, 3 sediment trap deployments and recoveries as well as the deployment of 11 long-term moorings. Some of these samples were processed on-board by the 17 research groups that participated in the cruise, although thousands more will be analyzed on-shore. Initial results include the characterization of the shelf cold pool, a feature that reflects the ice extent from earlier in the year and its relationship with the nutrient distributions on the shelf. The northern part of the cold pool was associated with an intense subsurface Chl *a* layer. Mesozooplankton distributions were patchy and few Euphausiids were found inshore of the 100 m isobath. Pollock larvae were found in greatest abundance along the Aleutian Peninsula and seaward of the Pribilof Islands. Education and outreach activities also were part of the cruise led by two teachers who reported to student and teacher groups on shore through webinars from the ship. They also organized community outreach at St. George Island during the cruise.

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1 Introduction

The Bering Ecosystem Study Project (BEST) focuses on the impact of seasonal sea ice on the environment of the eastern Bering Sea. More specifically, BEST seeks to clarify how sea ice influences the ecological pathways of nutrients and organic matter that lead to the abundant upper trophic levels and valuable fisheries on this extensive, high latitude continental shelf. More detailed background information can be found on the BEST home page (<http://www.fish.washington.edu/research/best/>). BEST also is part of a larger interagency effort to model the response of upper trophic levels to variations in climate forcing and more information on these collaborative efforts can be found on the web site of the Bering Sea Integrated Ecosystem Response Project (BSIERP; <http://bsierp.nprb.org/index.htm>). Science parties supported by both BEST and BSIERP participated in HLY0803.

The cruise described in this document is the third dedicated cruise for the BEST project. The first two took place in April-May of 2007 and 2008 and focused on the conditions directly associated with the retreating ice edge. HLY0803 examined the summer conditions on the eastern Bering Sea shelf. Although this region is ice-free in summer, the presence of ice earlier in the year influences the subsequent development of physical and biological conditions. The overall goal was to improve the understanding of these influences significantly during HLY0803. The cruise covered the entire shelf from the Aleutian Islands to St. Lawrence Island. A multidisciplinary sampling plan was carried out that included the deployment of moorings and physical oceanography, a hydrographic survey that collected discrete samples for a variety of chemical and biological analyses, zooplankton and ichthyoplankton net hauls, sediment sampling with a coring device, a variety of biological rate measurements that were done in on-board incubators on the bow, the deployment and retrieval of sediment traps that required small boat operations, as well as a variety of underway observations both from autonomous instruments sampling the sea chest water and visual observation of birds and marine mammals from the bridge.

There were several unique features of this initial summer cruise of the BEST/BSIERP program. Although no sea ice was present in the study region during the cruise, the residual effects of sea ice were still obvious. These effects included the cold bottom water that roughly reflected the extent of the earlier ice, and low salinity surface water that still retained the signature of the ice melt from April and May. HLY0803 provided an opportunity to build on the spring observations already collected in the first two cruises. Importantly, in addition to the physical development of the shelf waters, HLY0803 focused on the biological populations during their early summer conditions. For example, extensive net hauls were done to determine how far on the shelf populations of large oceanic grazers, that are important prey items for juvenile fish, had moved since the spring sampling. An important addition in this respect from earlier cruises was the inclusion of a science component on the ichthyoplankton to determine where and to what extent some of

the key fish populations such as pollock were spawning. Although the core hydrographic and primary productivity programs on HLY0803 were very similar in nature to those done on the spring cruises, the sampling approach differed in that the lack of ice favored a more standard survey approach in comparison to the featured based sampling associated with the productive ice edged of spring.

This report summarizes the sampling program carried out during HLY0803 along with a brief summary of the hydrographic conditions recorded. Many of the details on the science that will eventually result from this work are contained in the individual group reports that are included later in this report. Also, for more detailed information on the Healy data acquisition systems that continuously logged data from the many meteorological, sea water and other autonomous instruments used during the cruise, please refer to the Data Synopsis for HLY0803 prepared by: Tom Bolmer, David Forcucci, David Hassilev and Steve Roberts. One of the main goals of bringing together the many lines of information in the current report is to serve as a guide for researchers in identifying information relevant to their work that was collected during the HLY0803.

Final preparations for HLY0803 commenced on morning of July 2 when the Healy arrived at the Coast Guard dock at Dutch Harbor. Most of the science party arrived in town by the previously day and moved onto the ship during the mobilization day in Dutch Harbor (July 2). Cruise preparations proceeded quickly due to the fact that most of the science components had stored gear in Dutch Harbor after HLY0802. Cruise preparations also were aided by the development of a remarkably complete consumer environment in Dutch Harbor (rental trucks, hardware, and supplies of all kinds). Among the first priorities was a meeting among the senior scientists and officers of the Healy that would be interacting to implement cruise activities. Tom Weingartner from UAF agreed to act as co-chief scientist with R. Sambrotto on this ambitious and logistically complex cruise.

2 Science Groups & Participants

The major research components on HLY0803 and the associated participants were as follows:

Long-term moorings – Tom Weingartner, Jim Johnson, Dave Leech, Kevin Taylor.
This group deployed 9 moorings during the cruise.

Hydrography – Cal Mordy, Bill Floering, Dean Stockwell, Sigrid Salo and Dave Kachel (Dylan Righi and Ned Cokelet replaced the later 2 for the second leg).
This group analyzed salts, nutrients, oxygen and chlorophyll from the Niskin casts at each station as well as helped to manage cruise information.

Carbon Productivity - John Casey and Matt Tiahlo. This group carried out standard ^{14}C productivity experiments as well as size fractionated and cell sorted productivity.

Nitrogen uptake and cycling - Ray Sambrotto, Didier Burdloff and Kali McKee. This group carried out ^{15}N productivity experiments as well as urea and growth response experiments.

Particle flux – Roger Kelly, URI. Roger deployed a floating sediment trap on 2 occasions that collected particles for 24 hr. periods as well as measured Radium and Radon concentrations.

Iron analyses (leg 2 only) – Rob Rember, and Ana M Aguilar-Islas. This group collected samples for iron analyses on special casts from trace metal clean samplers.

Euphausiid and macrozooplankton collections - Alexei Pinchuk, UAF, and Tracy Shaw, OSU. Alexei collected macrozooplankton with a MOCNESS and CalVET net for quantitative distributions. Tracy collected live euphausiids with a Bongo net for rate measurements and organic tracer assays.

Euphausiid rate measurements - Megan Bernhardt, Tracy Shaw, Virginia Engel, UW (Lessard group) performed grazing, growth and reproduction experiments with euphausiids collected with Bongo nets using water collected on CTD casts.

Organic tracers of trophic transfer/euphausiid population age structure - Rachel Pleuthner, Karen Taylor, Charles Morgan, UMD (Harvey Group) extracted organic pools from zooplankton and their prey from net tows and water from CTD casts.

Ichthyoplankton – Nicola Hillgruber, Elizabeth Siddon and Ron Heintz. This group collected larval fish in collaboration with A. Pinchuk's net hauls.

Microzooplankton grazing – Diane Stoecker and Kristin Blattner, UMD. This group performed grazing experiments on water from the daily productivity cast.

Benthic biological characterization and fluxes - David Shull, and Greg Brusseau WWU. This group collected benthic samples with the multicorer and measured nutrient fluxes and Radon.

Benthic biogeochemical fluxes - Heather Whitney and Amy Cash. This group measured benthic fluxes of oxygen and nutrients on cores retrieved from the multicorer.

Bird distribution and abundance – Paul Suchanek and David Porter (leg 1) and Tom Van Pelt and Mark Rauzon (leg 2). FWS. They made observations from the bridge during the day.

Marine mammal distribution and abundance – Gary Friedrechsén, NOAA (guest investigator). Gary made observations from the bridge during the day.

Bio-optical and nitrate variations - Lisa Eisner, NMFS Auk Bay (guest investigator). Lisa added two instruments to the suite of instruments that analyze the science seawater – a WetLabs acs hyperspectral absorption instrument and a nitrate sensor. She also calibrated these instruments using samples from the Niskin rosette.

Water column bio-optics - Joaquim Goes, Eurico J. D'Sa, Puneeta Naik and Maria Fatima Helga do R. Gomes (guest investigators). This group measured profiles in the upper 100 m with their own bio-optical package that includes an FRRF. They also used an Advanced Laser Fluorescence System (ALFS) in both underway mode with the science seawater system as well as in discrete mode from samples from the Niskin rosette.

CTD operations and support - Scott Hiller and Sue Reynolds.

Science support activities - Steve Roberts and Tom Bolmer.

Data support - Janet Scannell (leg 1) and Scott Loehrer (leg 2)

Educational component - Teachers Jillian Worssam (PolarTrek) and John Karavias (Armada) took part in the cruise. A trainee from the MATE program (Alex Nanez) also participated.

3 Sampling Operations

3.1 Bridge Observations

During daylight hours, quantitative observations for birds and marine mammals were made from the bridge by the bird and marine mammal group. Both groups recorded GPS coordinated observations on laptops on the bridge.

3.2 Autonomous measurements from the ship

Continuous and autonomous sensing was done from the ship's seawater system as well as from meteorological instruments and depth recorders. A complete listing of these measurements along with their data format and calibration is available in the data synopsis by Tom Bolmer et al., cited above. Briefly, surface seawater was pumped from 8 m to the science laboratory where several measurements were made. These include temperature and salinity from a thermosalinograph, oxygen from a membrane electrode and chlorophyll *a* from a fluorometer. In addition to the ship's instruments, Lisa Eisner installed a nitrate sensor as well as a WetLabs ac-s that measured hyperspectral adsorption. Joaquim Goes installed an Advanced Laser Fluorescence System (ALFS) for phytoplankton characterization for part of the cruise.

3.3 On Station, Over-the-side Operations

Most stations began with a CTD cast from the starboard A-frame that included water sampling from a rosette of 12 - 30 L Niskin bottles to within 5 m of the bottom. This provided water samples for nutrients and other hydrographic measurements such as chlorophyll *a*. A variety of additional discrete samples were taken that varied from station to station. The stations can most easily be grouped by length into short, intermediate and long stations as described below.

3.3.1 Short Stations

In addition to the CTD cast, a short station may include sampling activities that add not more than 30 minutes onto the length of the station. These additional sampling activities may include for example, a productivity cast, a CalVET Net tow or ring net tow from the 3/8" wire off of the stern and an optics cast from an ancillary winch.

For these operations, the ship was stationary. The optics casts were done only during daylight hours with the optimum time period being within several hours of local noon (~14:30 ship time that remained on Alaska Time). In addition, the side of the ship from which the optics package was deployed faced the sun. After the CTD, the order of operations for short stations was the optics cast and then the net hauls.

3.3.2 Intermediate Stations

These stations may include almost any of the activities that were performed on HLY0803 with the exception of the sediment trap deployments that have a 24 hr. requirement. The difference between the intermediate and long stations is based on the number of activities carried out and the total water depth. Some of the activities that can be done during an intermediate station in shallow water (multicorer or iron for example) required a long station over slope and basin regions. Intermediate stations provide time for a variety of process oriented and experimental work including - CalVET net tow from the stern A-frame, 3/8" wire (Pinchuk); Bongo Net tows (2-3) at night (Lessard group); MOCNESS net tow (1) from stern A-frame, 0.68" conducting wire (Pinchuk and Hillgruber). A number of MOCNESS only stations were done in the first phase of the cruise to provide adequate sampling for the ichthyoplankton.

The MOCNESS tow was conducted at a speed of 1-2 knots. The multicore casts were conducted with the ship stationary. Benthic sampling typically was done at the end of the station or at a location slightly offset from the station location in order to avoid washing sediment into the water column during sample sieving and processing and deck cleanup.

3.3.3 Long Stations

Several CTD casts were conducted at each long station to assess diurnal variation. For multi-day stations, CTD were done on the morning of each day with succeeding casts interspersed with activities occurring on the stern in order to maximize efficiency and minimize down time while the CTD bottles are being emptied. Diurnal sampling was also done for the zooplankton collections. The long stations also provided an opportunity for deep iron casts. Long stations also provided enough time for sediment trap deployments consisting of a trap line (5/8" dia polydac rope) that is 110m long with samples collected at 25 m, 40 m, 50 m, 60 m and 100 m. Stations were limited to shelf-slope locations with water depths greater than 300 m, and deployments lasted approximately 24 hours. The traps were deployed from the ship but were retrieved using a small boat. The ship typically continued to sample spatially during the trap deployments.

3.3.4 Mooring Deployments

The moorings deployed during the cruise held CTDs at multiple depths as well as ADCPs to assess water movement. These are described in more detail in the individual group report section on moorings.

4 Additional Logistical and Operational Considerations

4.1 Educational outreach visit to St. George on July 11

A visit to St. George Island by the Healy was made on July 11 to interact with a Ocean Science Camp that was directed by Michele Ridgeway. Twenty students from the Pribilof Islands between the 5th and 12th grades participated in the camp that ran from July 10 through 18. In addition to the teachers, several science personnel visited the camp. These visits coincided with sampling in the waters around the Pribilof Islands and the collection of ship supplies from the airport at St. Paul.

4.2 Personnel exchange at St. Paul on July 17

A mid-cruise exchange of personnel occurred at St. Paul Island, Alaska on July 17. These exchanges were facilitated by the NOAA facility on the island (206-526-6453).

4.3 Existing Moorings to Avoid

We worked near four NOAA moorings. The positions are listed below.

Bering Sea 2 (M2) 56.877°N, 164.057°W, 73m water depth. (There also were some other moorings within 1 nm).

Bering Sea 4 (M4) 57.853°N, 168.870, 71m water depth (Note – This mooring was inoperable was located during the cruise).

Bering Sea 5 (M5) 59.898°N, 171.711°W, 72m water depth

Bering Sea 8 (M8) 62.194°N, 174.668°W, 73m water depth

5 Cruise Track and Narrative on 4 Phases

The overall cruise track consisted of several cross shelf sections together with a long shelf (70m) section that re-occupied regions that were sampled in prior BEST cruises. An inshore line northwest of Nunivak Island and 9 additional stations were added on HLY0803 to accommodate the mooring deployments (Figure 1). In addition to the division of HLY0803 into 2 legs by the personnel exchange at St. Paul on July 17, the sampling activities were further organized into 4 phases of operation (2 phases before the St. Paul stop and 2 afterwards). The 4 phases and their respective major foci were as follows:

Phase 1 – July 3 – 8: *Sampling of the non-ice impacted region of southeastern Bering Sea and ichthyoplankton collection.* The southern region was the only one not impacted significantly by ice this year and provides a contrast to the heavily ice-impacted northern shelf. In addition, it is the main spawning region for pollock and other important commercial species and collections of ichthyoplankton were a top priority during this phase. Collection of larval fish by Nicola Hillgruber and Alexei Pinchuk reflected a variety of species including Pacific cod, arrowtooth flounder and walleye pollock. Hydrographic sampling began in Unimak Pass in the southern part of the eastern Bering shelf (line UP). Four stations were occupied across this pass to characterize the water entering the Bering Sea from the Gulf of Alaska, particularly the fresh Alaska Coastal Current (ACC) that continues along the inner

shelf of the eastern Bering. We next sampled along a series of cross-shelf sections in the southeastern Bering Sea (lines UAP, CN and CNN). This phase included stations 1 to approximately 29.

Phase 2 - July 8-16: *Deployment of moorings in inner and middle shelf regions.* The initial deployment of moorings by Tom Weingartner's group began during this phase. The mooring group deployed three moorings along each of the NP, W and SL lines at 25m, 40m and 55m. Tom decided not to deploy a tenth mooring near the SL-55m mooring site. The mooring deployments are logistically complex when combined with the many simultaneous process studies on HLY0803. The requirement for daytime deployments for the moorings, and the avoidance of long work periods for the mooring crew meant that other work needed to be scheduled around the mooring efforts to accommodate its schedule. We received cooperation from the other research groups however, and were able to characterize summer physical and biological conditions in the inner shelf during summer in addition to deploying the moorings. We also completed most of the NP line. Unfortunately, a bio-optical sensor package brought on-board by Eurico D'Sa and Joaquim Goes was lost on July 8. A replacement system was obtained for the second leg of the cruise however, that contained a spectral radiometer, back-scatter meter, fluorometer and CTD and this new package restored most of the lost measurement capabilities. Pat Kelly did two drifting sediment trap deployments from Brad Moran's group in the slope region. Microplankton productivity was the concern of several groups on board including Ray Sambrotto's nitrogen productivity group, Mike Lomas's carbon productivity group and Diane Stoecker's microzooplankton group. These three groups coordinated their sampling activities on the daily productivity cast. Euphausiid rate measurements were done when enough animals were obtained by Evelyn Lessard's group and these conditions were found at several stations in the outer shelf. Roger Harvey's group collected samples for later organic analyses. David Shull has been able to collect a number of sediment cores with the multicore, and together with Al Devol's group worked on the sediment profiles and fluxes. Lisa Eisner calibrated her underway nitrate measurements against the on-board autoanalyzer system and the results from the continuous nitrate sensor appear useful. The CTD operations continued without major problems and the data support group has been able to utilize the existing resources to update and expand the BEST field catalog as we progress. The bird observers manned the bridge each day, although fog restricted observations. The two teachers on board have contributed daily journals of their activities and observations on the cruise. The educational outreach on St. George was done during this phase and it ended with the participant exchange at St. Paul.

Phase 3 - July 18-26: *Sampling of the northern, outer shelf region.* This region has been the most consistently impacted by the ice edge during the last decade. The region supported intense primary production in spring 2007 and scattered production in spring 2008. Sampling characterized the seasonal development of production in this region and will help to determine how the earlier ice cover influenced it. Of particular interest is the shelf break frontal region (at locations

between approximately 100-200 m water depth). This region supports consistent summer production along the southern shelf and is associated with the physical mixing of deeper shelf waters to the surface. This phase included the occupation of section MN and the outer part of section SL. Together with the more inshore stations on SL occupied in phase 2, this completed the SL line. Additional moorings were also deployed in the inner shelf. This phase included stations 83 to approximately 121.

Phase 4 - July 26-31: *Mid-shelf sampling along 70 m isobath and characterization of the shelf cold pool.* This provided a mid-summer sampling of the middle shelf and contributed towards a better understanding of the seasonal evolution of the physics, chemistry and biological distributions on the shelf. In particular, it helped to define the extent of the shelf cold pool that is formed in winter in association with ice production. This current occupation of the 70 m line, and the prior occupations done on previous BEST and NOAA cruises, has provided several years of seasonal observations on the changes in the middle shelf.

6 Meteorological Conditions

Sea state did not stop sampling activities at any time during the cruise even though winds exceeded 25 knots. several times during the cruise (Figure 2A). Winds were generally out of the north and west, with some southerly winds during phase 2. The northwesterly winds generally were associated with greater wind speeds. Wind was

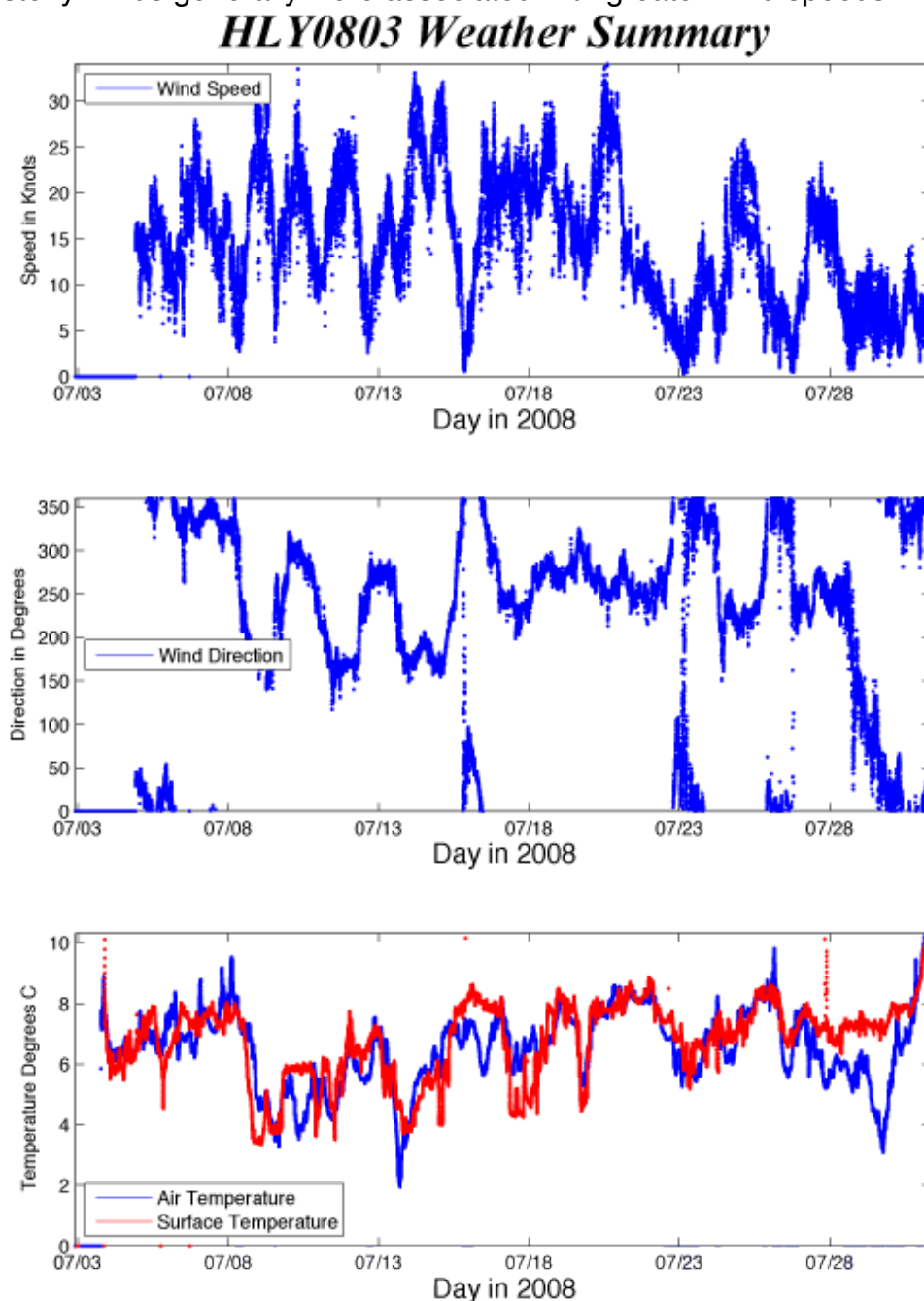


Figure 2. Meteorological conditions during HLY0803. Top – wind speed; middle – wind direction; 3 – surface water and air temperatures.

an issue with sea-keeping on station due to Healy's large freeboard area, and particular care was taken to keep the bow into the wind during casts. The major meteorological impediment to sampling was fog that was common over the cold waters of the inner and middle shelf. This required slower speeds and delayed progress along the planned station lines. Due to the lack of weather days caused by sea state and the faster than anticipated speeds made under clear conditions, however, the delays due to fog had no appreciable impact on overall sampling.

7 Underway and Remotely Sensed Data

The continuously measured properties from the ship's underway system provide a natural compliment to the remotely sensed features of surface waters. Some of the major parameters measured along the ship's track are shown in Figure 3. The ship recorded the coldest waters around Nunivak Island and these cold temperatures extended well offshore north of the Pribilof Islands. Water over the outer shelf and slope regions was generally the warmest, although surface waters of the southern shelf warmed appreciably between the beginning and end of the cruise. The pattern of surface salinity was different that that of temperature (Figure 3 B). Surface salinities were much lower in the middle shelf waters of the northern region

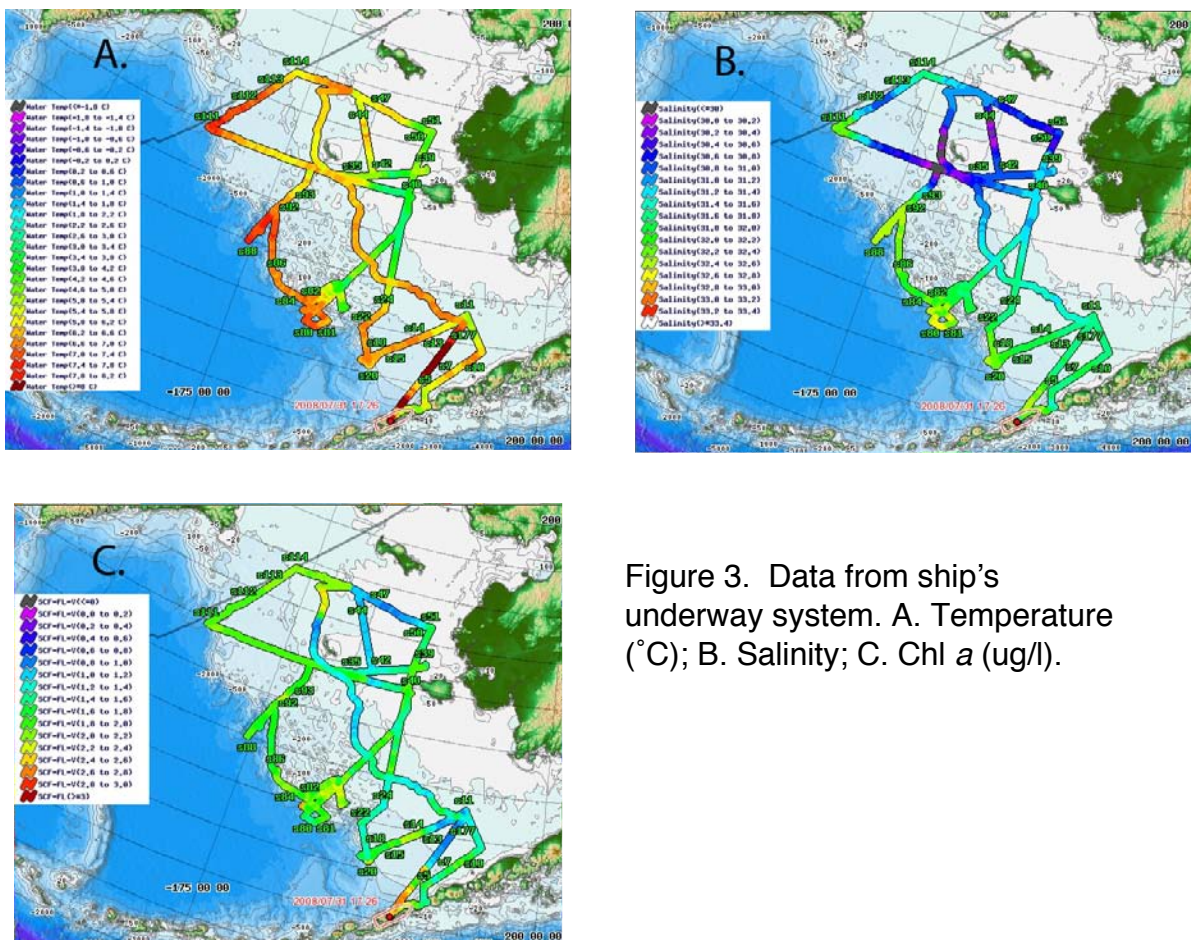


Figure 3. Data from ship's underway system. A. Temperature (°C); B. Salinity; C. Chl a (ug/l).

than in the south. This pattern also was reflected in the surface Chl *a* levels (Figure 3 C) that were lower in the low salinity regions than in the south.

This is a good example of the link we hope to make between the earlier ice extent and the subsequent patterns of biological production. In this case, it appears that the regions of ice melt that caused the low salinity surface waters that persisted to July, also resulted in less surface Chl *a* in summer. This may be due to greater vertical stratification in the northern regions and the resulting lower supply of nutrients that reach the upper euphotic zone.

The remotely-sensed, sea surface temperature (SST) at the end of July 2008 was similar to the pattern shown by the ship measurements (Figure 4 A). Most obvious, was the cold surface water in the near shore region, with the lowest temperatures in the Nunivak Island region. In the MODIS Chl *a* distribution from early July (Figure 4 B), the region of low Chl *a* on the northern shelf as suggested by the later ship sampling is also apparent. The regions of elevated Chl *a* recorded in the southern shelf and near Unimak pass in the MODIS scene, also appear in the ship data a week or 2 later. Unfortunately, later weeks in the cruise did not provide an opportunity for another clear scene.

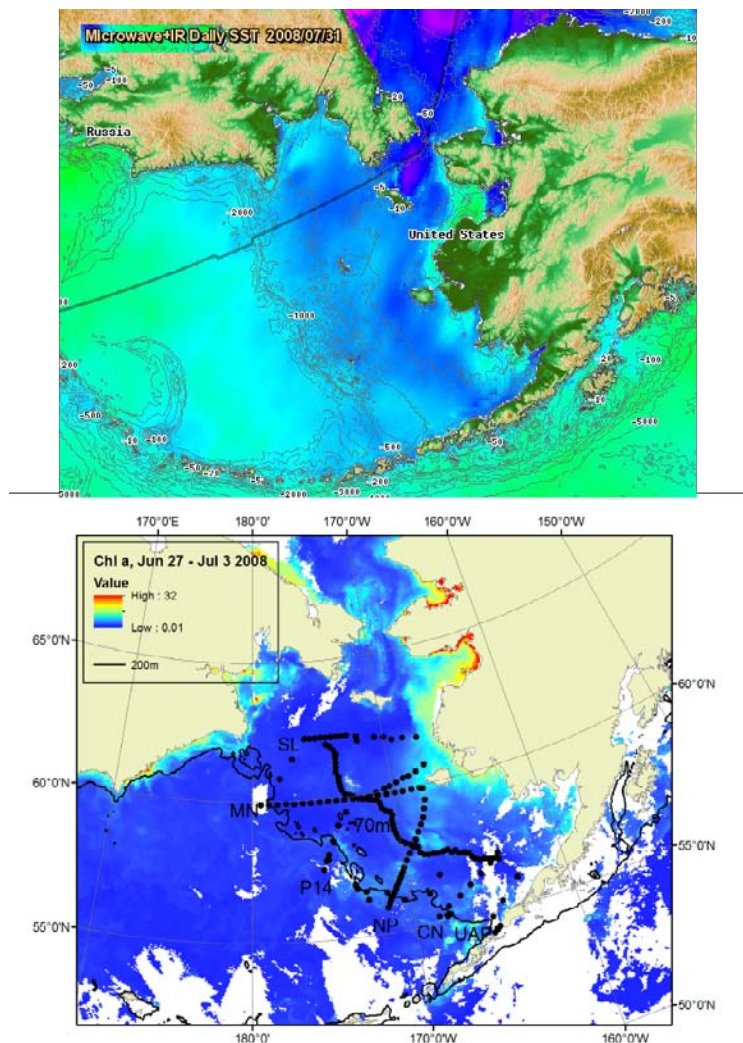
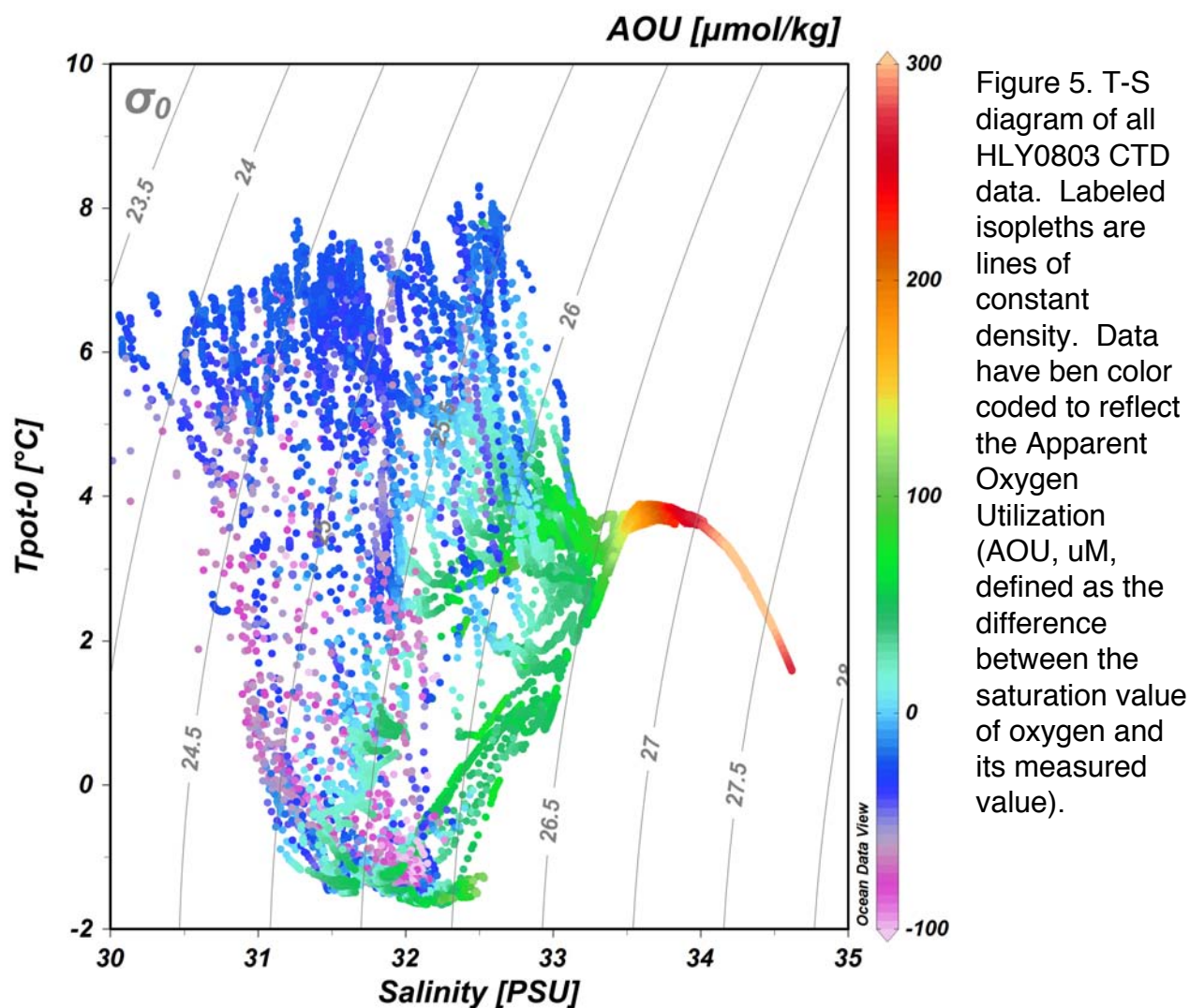


Figure 4. Remotely sensed data of the Bering Sea in and around the time of HLY0803. Top – Daily SST image from 7/31/08. Bottom – MODIS Terra Chl *a* image for the 8-day period from June 27-July 3, 2008.

8 Preliminary Hydrographic results

The salinity and temperature data were processed on the ship to remove spurious measurements and to calibrate the sensor values against the salinometer measurements that were made. The T-S plot for all of the CTD casts made on HLY0803 is shown in Figure 5. The plot clearly shows the high salinity 'tail' of deep basin and slope water mixing into the lower salinity shelf waters as denoted by the decrease in temperature as the water mixes below a salinity of 33.5. The plot indicates the mixing between these warmer waters and the cold pool waters. This mixing is largely at depth as reflected by the elevated AOU (~ 100 μM) along the mixing line. The warming of this mixture is associated with increases in oxygen as AOU falls to negative values. This is compatible with the mixing of this water into the surface where it ventilates and gains oxygen from photosynthesis.



The nutrient data measured under the direction of Cal Mordy has not been finalized, but is suitable for developing sections for preliminary review of data from the cruise. For this report, 9 properties were plotted for each of the 5 major sections (CN, NP, MN, SL and 70m lines). The properties include salinity, temperature, the 5 nutrients (nitrate, nitrite, ammonium, phosphate and silicate), Chl *a* fluorescence and density. Even at this stage of processing, the property distributions are very interpretable and show little if any spurious data. It appears that the hydrographic data from HLY0803 will soon be ready to use and yield extensive information on the summer conditions on the eastern shelf.

Although it is not the purpose of this report to analyze the hydrography from the cruise in detail, a brief overview will serve to illustrate the robust nature of the data and some of the ways it can be used. For each of the sections, the 9 properties have been grouped into 5 sections, each with one property distribution colorized and another overlain as labeled isopleths. The five combinations are: temperature with salinity overlain; nitrate with density overlain; fluorescence with nitrate overlain; ammonium with nitrite overlain; phosphate with density overlain; and finally silicate with density overlain. Starting from the southern cross-shelf section CN (Figures 6 & 7), the salinity distributions shows the higher salinity slope water encroaching on the middle shelf at depth (Figure 6A). Colder ($<1^{\circ}\text{C}$) water persists at depth in the deeper middle shelf region. This is a pattern common to all cross shelf sections on the eastern shelf. It also manifests itself in the distribution of nutrients. The deep-water nutrients nitrate, phosphate and silicate, all show the same spatial distribution as salinity (Figures 6B & 7 top & bottom). Although Chl *a* levels are much lower than they were in spring (Figure 6C), there is still significant Chl *a* in the surface waters of the southern shelf. A major feature of the summer shelf is the intense regeneration signal associated with the middle shelf sediments, here illustrated by the elevated ammonium levels (Figure 6D). The ammonium distribution also displays another feature that is consistent on this shelf. This is the subsurface ammonium maximum that connects the high ammonium bottom waters on the middle shelf to the surface waters of the shelf break front.

The distribution along the NP line is a useful contrast with the others because they show the large impact of the Pribilof Islands and the vertical mixing in the nearby water (Figures 8 & 9). This mixing is reflected by the elevated nutrient concentrations that have made their way well onto the shelf as the slope water mixes around the islands (Figures 8B & 9). The elevated ammonium levels at depth and their extension offshore between the 25.5 and 26 isopycnals are still obvious however (Figure 8D). The relationship between the deep water nutrients and the salinity distributions remain clear along the MN line (Figure 10 & 11). The MN line passes through a much colder region of the shelf cold pool and had lower surface salinity values than the more southerly sections, however (Figure 10A). The MN line also exhibits a more intense sub-surface Chl *a* layer and sub-surface ammonium layer than was present in the south (Figure 10C&D). The middle shelf ammonium maximum at depth is also present, but in the colder bottom water has a lower concentration than in the south.

The SL line terminates at the international boundary and does not include the slope region that the more southerly sections do (Figures 12 & 13). The relationship between the slope water and deep-water nutrients is still clear however. The subsurface Chl a levels are more intense still than the more southerly lines (Figure 12C). The mid-shelf ammonium levels at depth along the SL line are as large as those along the CN line (Figure 12D).

The distributions along the 70 m line tie together many of the features seen in the cross shelf sections (Figure 14 & 15). For example, the 70 M line shows the disappearance of the sub-zero cold pool temperatures between 400 and 600 km (just off Nunivak Island). It also shows the long shelf extent of the low salinity surface waters that were first seen in the MN section. These low salinity surface values show a distinct pool, probably reflecting the residual melt water from the spring. The 70 M line also confirms that the subsurface Chl a layers are more intense in the north and that the bottom water ammonium more concentrated, in the south. Interestingly, the 70 m sections also show a distinct difference between the silicate and phosphate concentrations of shelf bottom waters (Figure 15 top & bottom). While the silicate levels drop over 50% in the region between Nunivak and Pribilof Islands, the phosphate levels decrease less than 30%. The smooth variations and interpretable nature of these sections is encouraging.

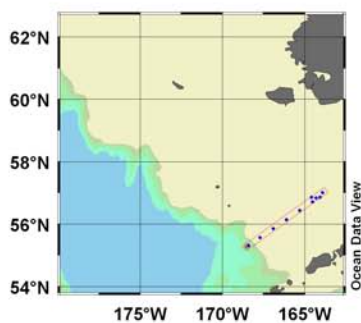
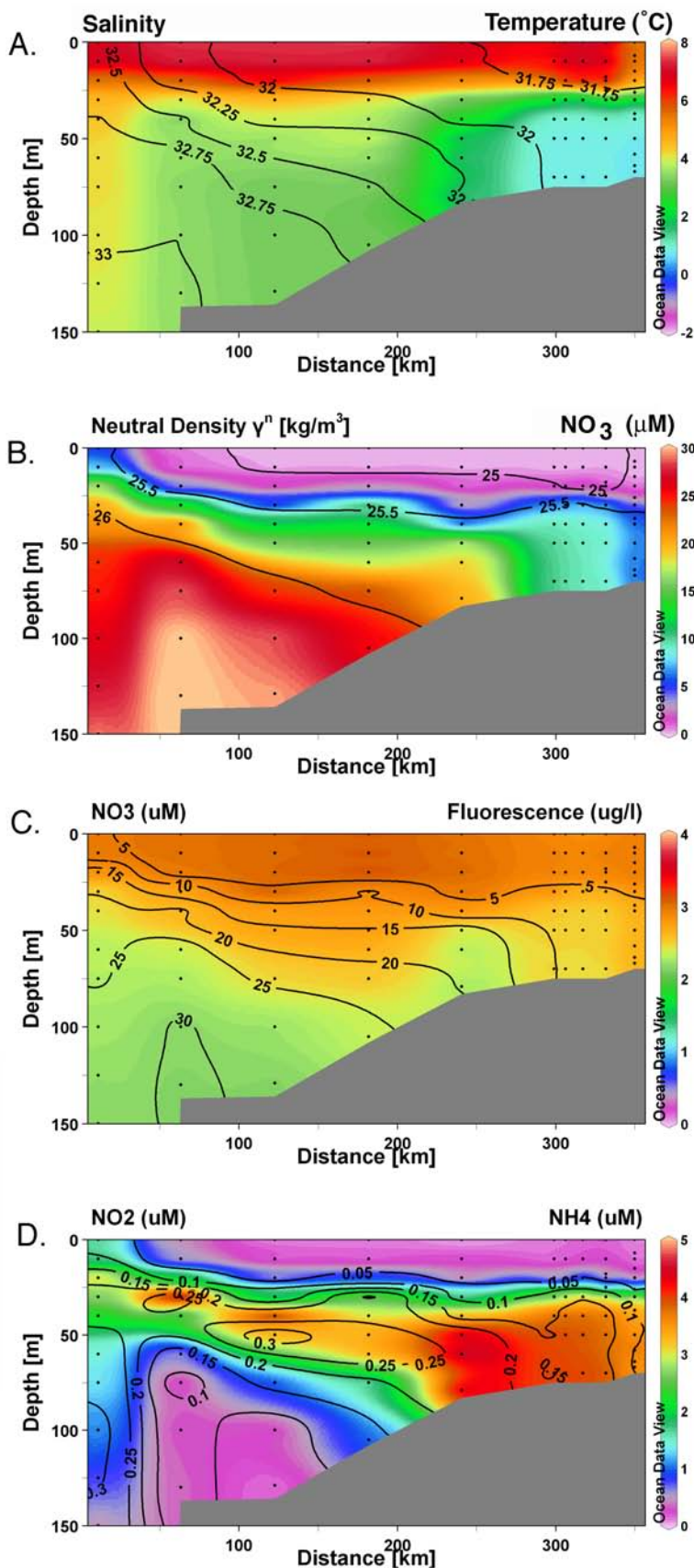


Figure 6. Sections from the CN line (Fig. 1 & inset above; distances are from northern end of line). A. Temperature ($^{\circ}\text{C}$; color bar) with salinity (labeled isopleths). B. Nitrate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). C. Chl *a* fluorescence ($\mu\text{g l}^{-1}$; color bar) with nitrate (μM ; labeled isopleths). D. Ammonium (μM ; color bar) with nitrite (μM ; labeled isopleths).



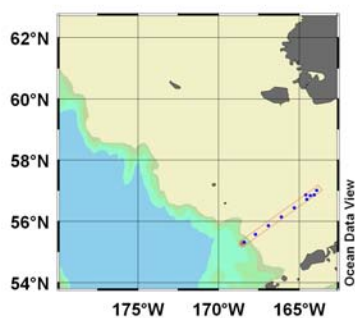
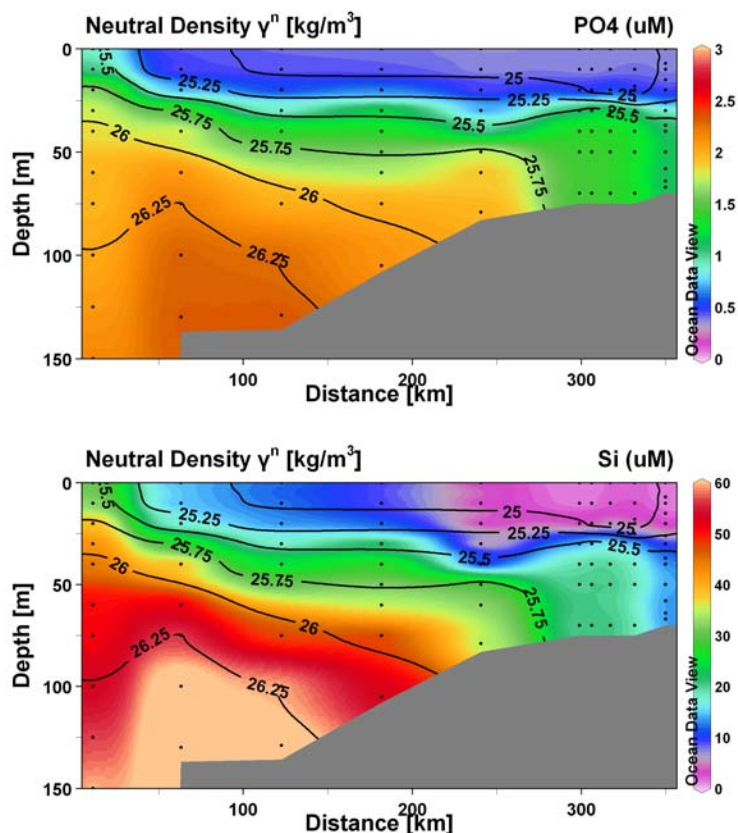


Figure 7. Sections from CN line, continued. Top. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). Bottom. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths).



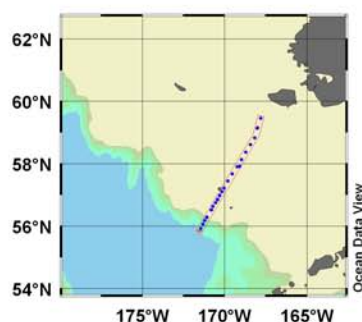
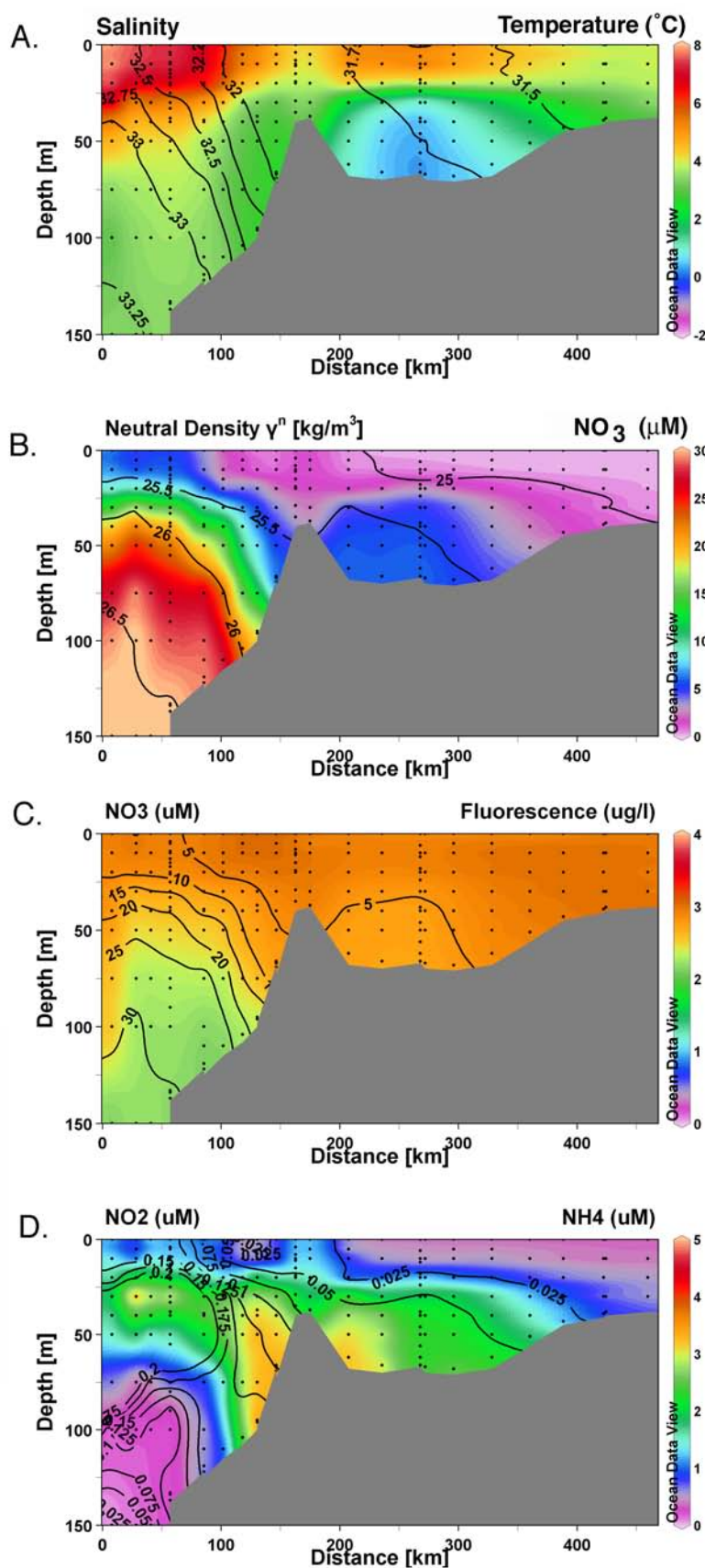


Figure 8. Sections from the NP line (Fig. 1 & inset above; distances are from northern end of line). A. Temperature ($^{\circ}\text{C}$; color bar) with salinity (labeled isopleths). B. Nitrate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). C. Chl *a* fluorescence ($\mu\text{g l}^{-1}$; color bar) with nitrate (μM ; labeled isopleths). D. Ammonium (μM ; color bar) with nitrite (μM ; labeled isopleths).



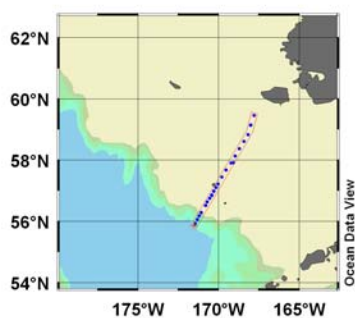
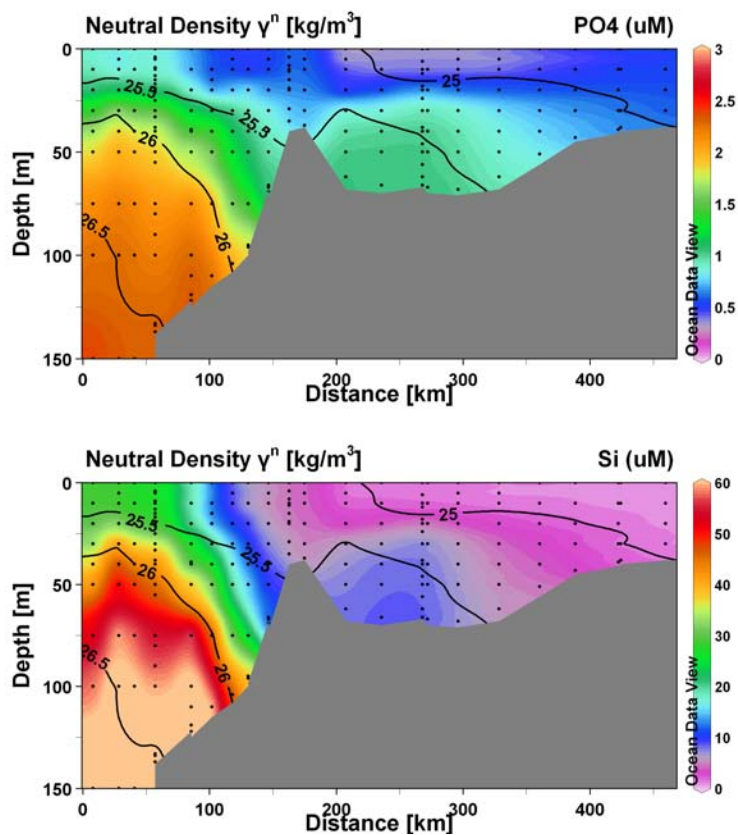


Figure 9. Sections from NP line, continued. Top. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). Bottom. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths).



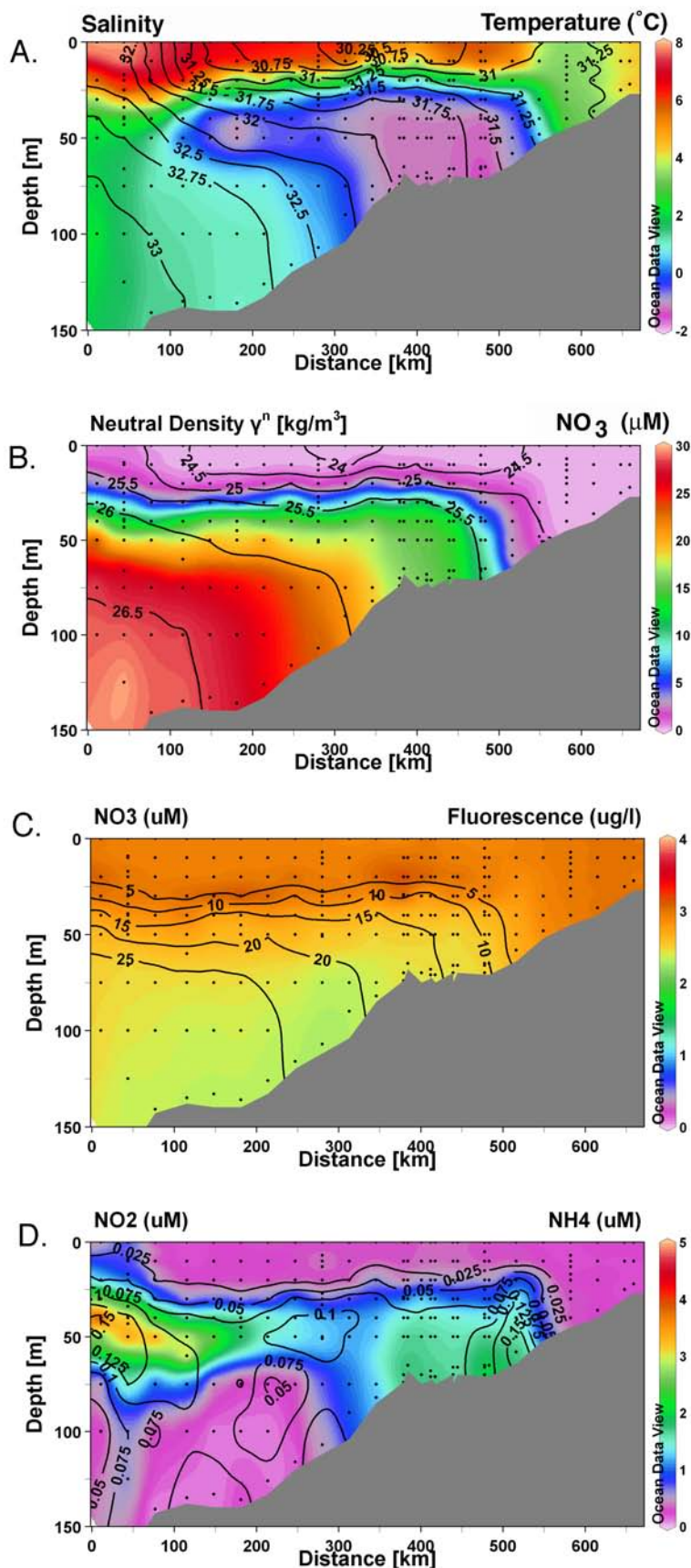
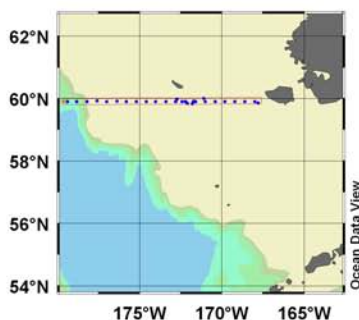


Figure 10. Sections from the MN line (Fig. 1 & inset above; distances are from northern end of line). A. Temperature (°C; color bar) with salinity (labeled isopleths). B. Nitrate (μM; color bar) with neutral density (kg m⁻³; labeled isopleths). C. Chl *a* fluorescence (μg l⁻¹; color bar) with nitrate (μM; labeled isopleths). D. Ammonium (μM; color bar) with nitrite (μM; labeled isopleths).

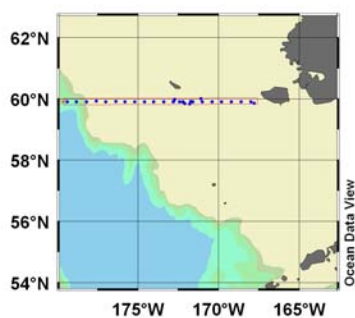
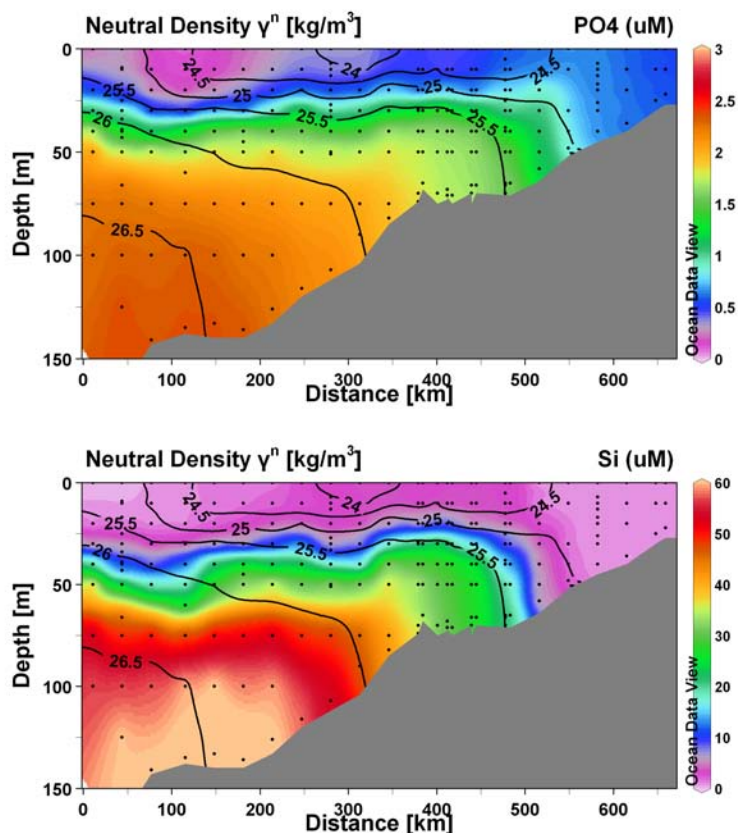


Figure 11. Sections from MN line, continued. Top. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). Bottom. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths).



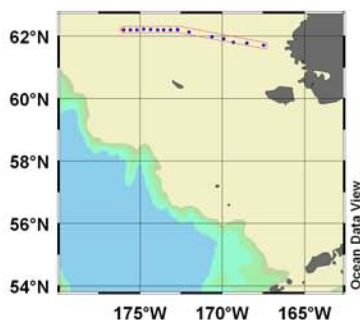
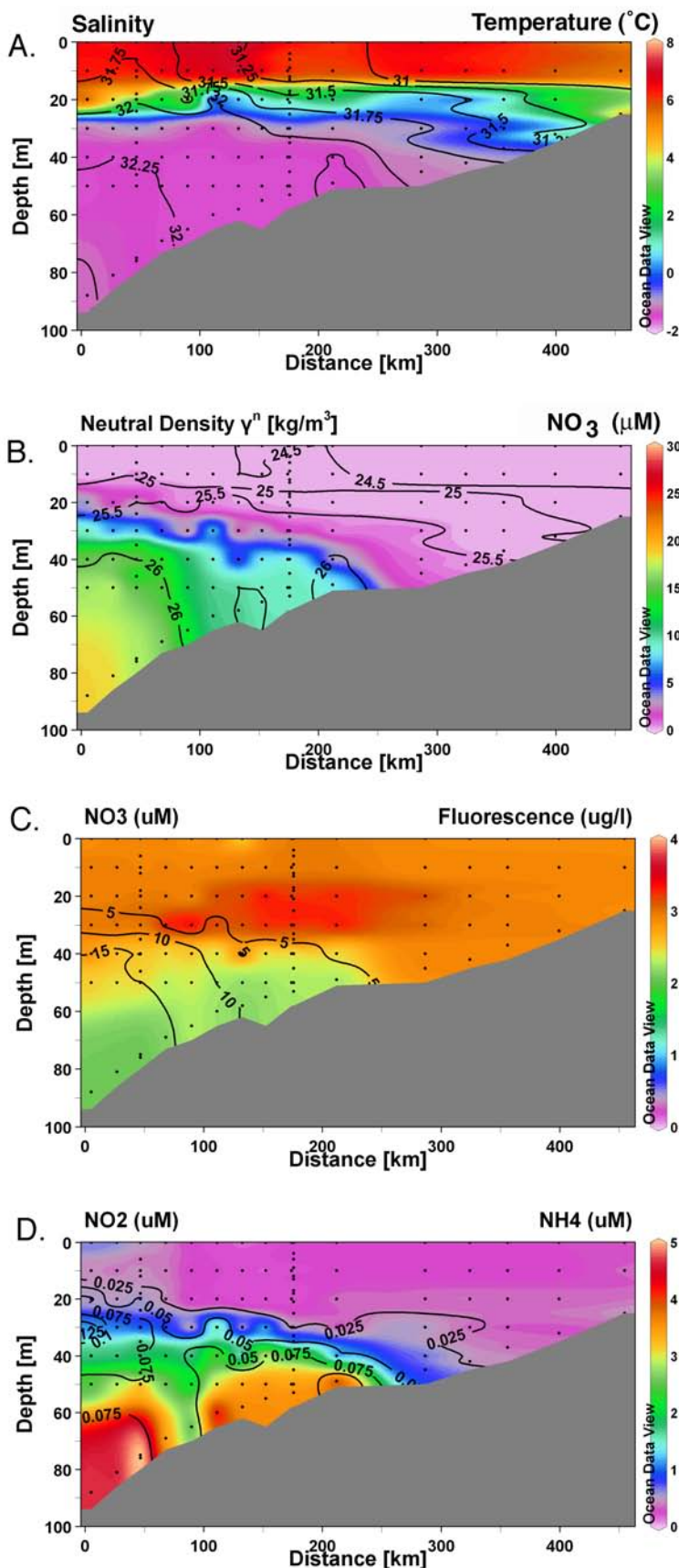


Figure 12. Sections from the SL line (Fig. 1 & inset above; distances are from northern end of line). A. Temperature ($^{\circ}\text{C}$; color bar) with salinity (labeled isopleths). B. Nitrate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). C. Chl *a* fluorescence ($\mu\text{g l}^{-1}$; color bar) with nitrate (μM ; labeled isopleths). D. Ammonium (μM ; color bar) with nitrite (μM ; labeled isopleths).



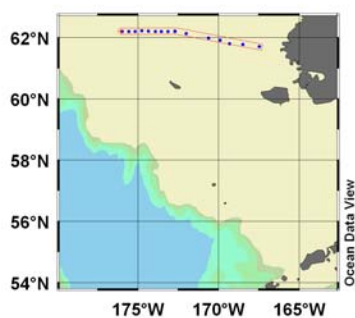
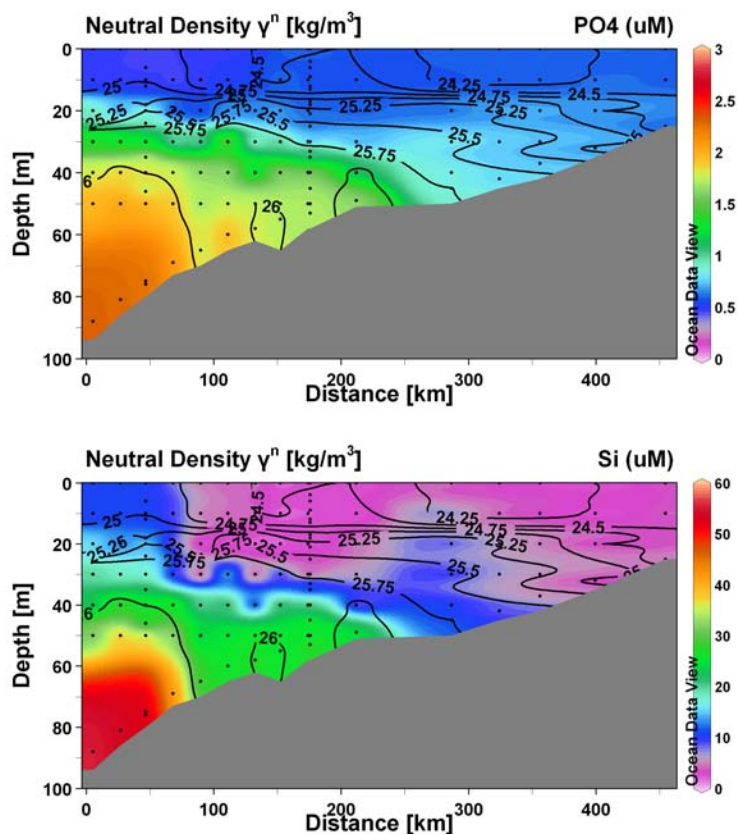


Figure 13. Sections from SL line, continued. Top. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). Bottom. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths).



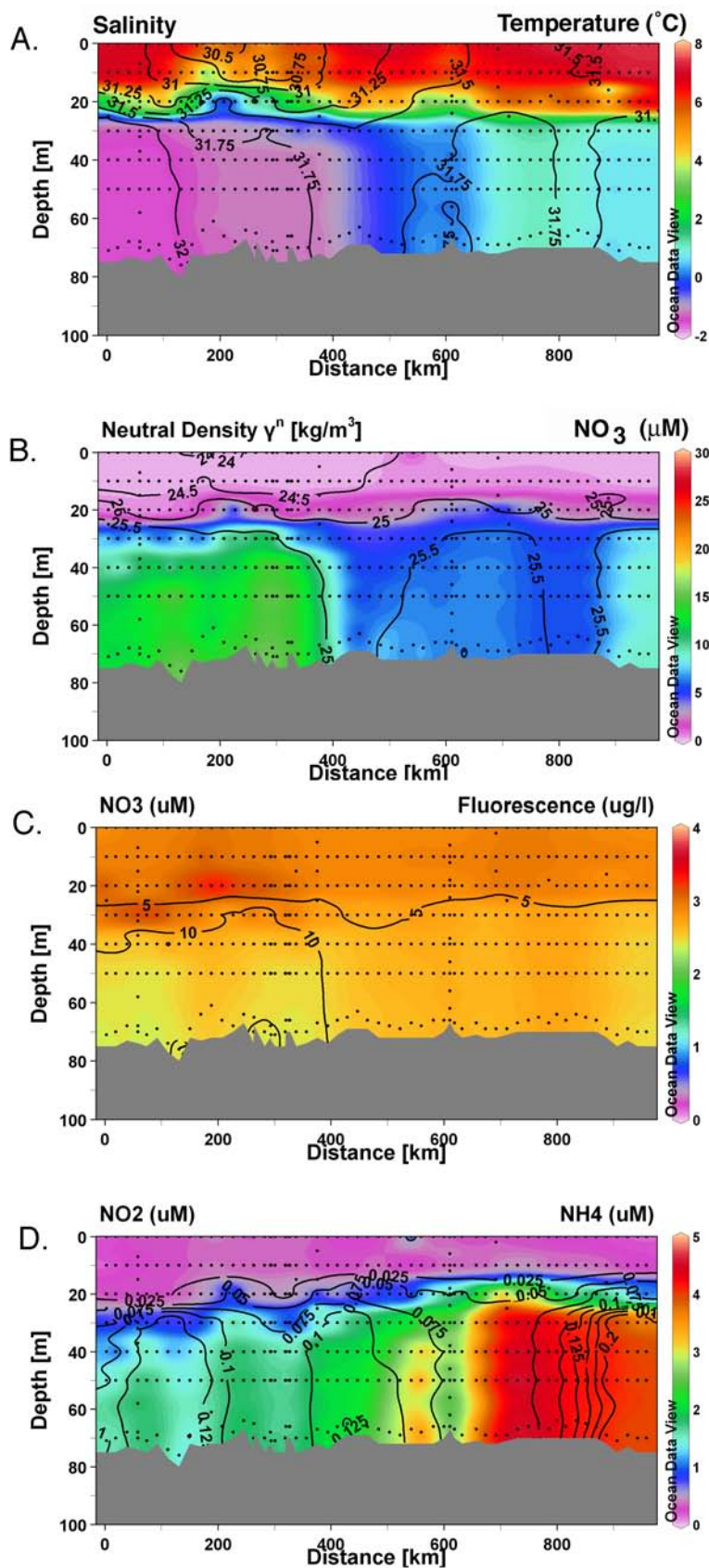
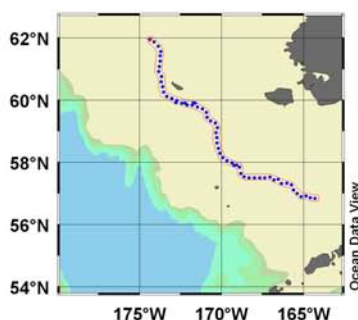


Figure 14. Sections from the 70 m line (Fig. 1 & inset above; distances are from northern end of line). A. Temperature (°C; color bar) with salinity (labeled isopleths). B. Nitrate (μM; color bar) with neutral density (kg m⁻³; labeled isopleths). C. Chl *a* fluorescence (ug l⁻¹; color bar) with nitrate (μM; labeled isopleths). D. Ammonium (μM; color bar) with nitrite (μM; labeled isopleths).

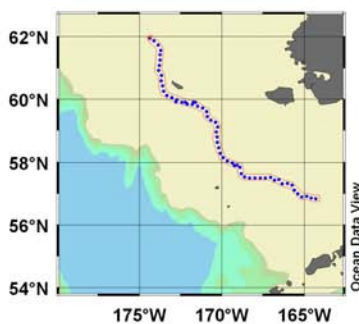
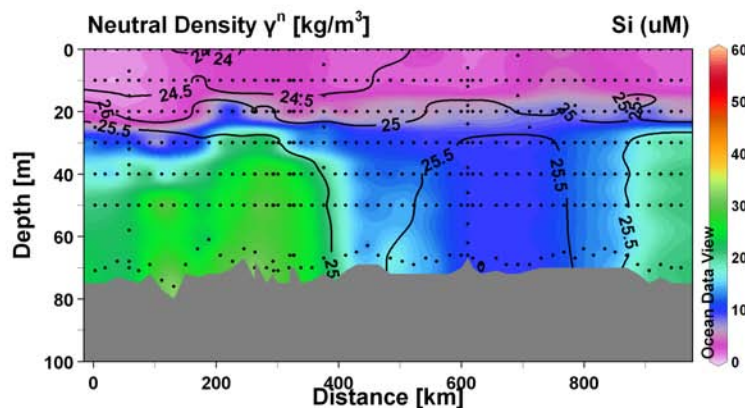
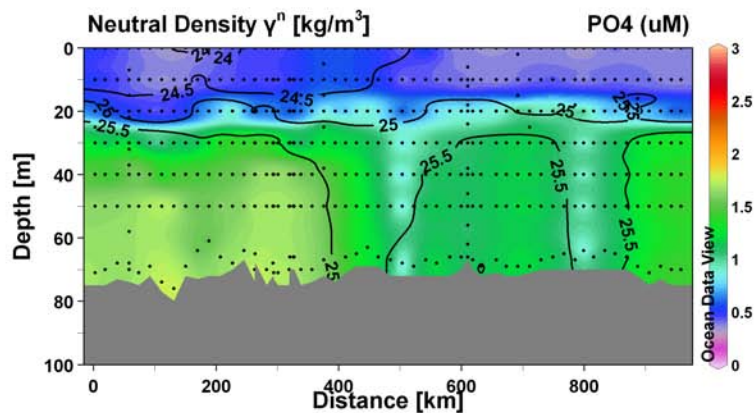


Figure 15. Sections from 70 m line, continued. Top. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths). Bottom. Phosphate (μM ; color bar) with neutral density (kg m^{-3} ; labeled isopleths).



9 Individual Group Reports

9.1 Mooring group - Tom Weingartner

Collaborative Research - Stratification on the Bering Shelf and its consequences for nutrients and the ecosystem: The effects of ice and coastal water advection (K. Aagaard and T. Weingartner)

Our observational program is focused on how freshwater, either re-distributed by the shelf circulation or introduced from sea ice, modifies stratification and nutrient distributions over key portions of the central Bering shelf (between Nunivak and St. Lawrence islands). To achieve this goal we will:

1. describe the cross-shelf and vertical structure of selected physical properties in the coastal and mid-shelf domains, and examining how these vary seasonally and interannually.
2. identify the physical mechanisms controlling cross-shelf and vertical distributions of freshwater and nutrients, including the exchange processes between the coastal and mid-shelf domains; and
3. assess the relative roles of ice melt and meteoric waters (including runoff) in stratifying the shelf, and how changes in these freshwater sources can be expected to modify the shelf ecosystem.

Two principal hypotheses underlie these objectives. First, we propose that the central Bering shelf is significantly freshened by offshore transport of low-salinity coastal waters through most of the year. This cross-shelf freshwater flux influences not only the stratification on the shelf, but also the nutrient supply, since the coastal waters are typically deficient in macro-nutrients, even though they appear to be enhanced in iron. Second, we hypothesize that the effect on stratification of the cross-shelf flux of coastal waters varies seasonally, depending on the wind forcing and the strength of the coastal flow, the latter in turn being dependent on runoff. For example, in summer, when river discharge is high, we expect that the interaction of a vigorous buoyant coastal flow and relatively weak, but upwelling-favorable winds, with the change in coastline orientation north of Nunivak Island will result in along-shelf convergence and the offshore expulsion of coastal waters. Unless the inner shelf front is unstable, however, that front will remain intact, so that essentially the coastal domain expands seaward with little cross-frontal transport. On the other hand, in late summer and fall, strong upwelling-favorable winds and reduced runoff weakens the inner front so that the coastal and mid-shelf domains merge as the coastal waters move offshore. Both of these hypotheses involve interannual variability. In part this comes about because of interannual changes in runoff and winds, but in part also because of an intricate interplay between stratification, winds, and tides.

Our observational program includes shipboard CTD/rosette sampling that includes $\delta^{18}\text{O}$ measurements. We have collected ~ 300 $\delta^{18}\text{O}$ samples with measurements typically at the surface, 10 m and bottom on the shelf. We also deployed a nine-mooring array over the central shelf between the 55 and 25 m isobaths, which links to the NOAA-PMEL moorings deployed along the 70 m isobath (Figure 1).

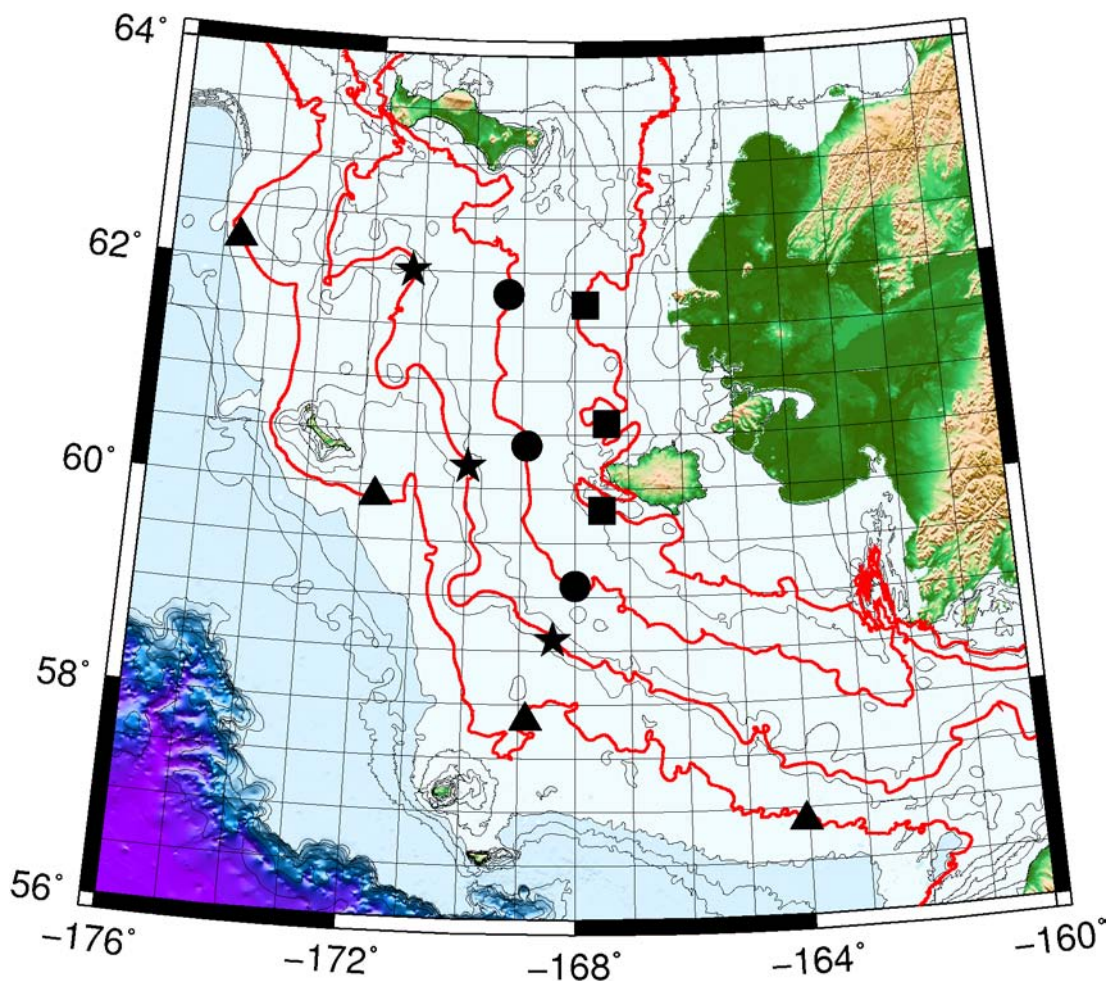


Figure 1. Mooring location map:

Triangles are the NOAA moorings M2, M4, M5 and M8 (on the 70 m isobath).

Stars are moorings S55, C55 and N55 (on the 55 m isobath)

Circles are moorings S40, C40 and N40 (on the 40 m isobath)

Squares are moorings S25, C25 and N25 (on the 25 m isobath)

Red contours: 25, 40, 55 and 70 m isobaths. Black contours: 10, 20, 30, 50, 60, 80, 90, 100, 150, 200, 250, 500, 1000, 2000 & 3000 m isobaths.

The mooring deployments are summarized in Table I. The instruments sample at half-hourly rates and store data internally. They will be recovered and replaced with fresh instruments in summer 2009, with the final recovery scheduled for summer 2010. Table II lists the instruments on each mooring.

Table I. Healy 0803 BEST Mooring deployments

Mooring	Date/time ¹ (ADT)	Longitude (W)	Latitude (N)	Depth ² (m)	Comments
N55	2131 July 11	171 58.488'	61 57.717	54.4	25 kts, rainy. Deployment smooth. 1454 m/s
N40	16:30 July 12	169 16.722'	61 48.328	40.3	12 kts, light seas, deployment smooth. 1450 m/s
N25	22:54 July 12	167 27.112	61 42.006	25.6	14 kts, light seas, smooth deploy. :1468 m/s
C55	1151 July 11	170 5.367'	60 10.316'	54.91	Deployment smooth, 15 kts overcast. 1440m/s
C40	1534 July 10	169 1.266'W	60 20.204'N	40.5	Deployment smooth calm and overcast
C25	2116 July 10	167 20.257'W	60 40.959'N	24.9	Deployment smooth; calm and overcast.
S55	22:40 July 13	168 23.646	58 35.378	55.3	1457 m/s. foggy, 32 kts, deployment smooth
S40	1423 July 8	167 58.94	59 8.175'	42	Deployment smooth; 20 -25 S and seas building. Rain
S25	2230 July 8	-167 46.885	59 51.088'	26.2	Deployment smooth 25 kts S and 7 – 8' seas. About 10 miles NW of nominal position due to shoaling bottom.

¹ GMT = ADT + 8 hours

²includes hull and sound speed correction based on CTD prior to deployment.

Table 2. BEST MOORING PLAN (Aagaard, Danielson and Weingartner)

Mooring	Instrumentation
N55	SBE 16/37 @ 20*, 50 m; Upward-looking 300Khz ADCP @ 49 m; T chain**; IceCat @ 10m
N40	SBE 16/37 @ 20* & 35 m, Upward-looking 300Khz ADCP @ 34 m
N25	1-m Bottom Tripod SBE 37 Upward-looking 300Khz ADCP
C55	SBE @ 20* & 50 m Upward-looking 300Khz ADCP @ 49 m; T chain**; IceCat @ 10 m
C40	SBE 16/37 @ 20* & 35 m Upward-looking 300Khz ADCP @ 34 m
C25	1-m Bottom Tripod SBE 37 Upward-looking 300Khz ADCP
S55	SBE 16/37 @ 20*, 50 m; Upward-looking 300Khz ADCP @ 49 m; T chain**; IceCat @ 10 m
S40	SBE 16/37 @ 20* & 35 m, Upward-looking 300Khz ADCP @ 34 m
S25	1-m Bottom Tripod SBE 37 Upward-looking 300Khz ADCP

* SBE @ 20 m on N40, C40, S40 and N55, C55, S55 include fluorometer

** T chain consists of 15 internally logging T recorders at 2.5 m intervals

9.2 Hydrography & PMEL group

Leg 1: Sigrid Salo¹, David Kachel¹, Calvin Mordy², William Floering¹

Leg 2: Edward D. (Ned) Cokelet¹, Dylan Righi³, Calvin Mordy², William Floering¹, Andrew Koehn³

¹NOAA/Pacific Marine Environmental Laboratory

²GenWest Systems, Edmonds, WA

³University of Washington Joint Institute for the Study of Atmospheres and Oceans

The BEST Hydrography Group conducted 210 CTD casts at 177 oceanographic stations. The CTD was a Sea Bird Electronics SBE 911 plus with dual temperature and conductivity sensors. It carried an auxiliary SBE 35 temperature sensor, a

WETLabs optical transmissiometer, a Chelsea Aqua Tracker fluorometer, a Biospherical QSP2300 PAR sensor and a Benthos 916 altimeter. The group conducted CTD casts which included nutrient samples from up to 12 thirty-liter Niskin bottles, two or more Winkler oxygen samples for calibration of the CTD oxygen sensor, three O^{18} samples for Tom Weingartner of the University of Alaska Fairbanks (UAF), and two to seven Dissolved Organic Carbon (DOC) and Dissolved Inorganic Carbon (DIC) samples for Jeremy Mathis of UAF. Table 1 summarizes the sampling. Scott Hiller and Sue Reynolds, Scripps Institution of Oceanography, operated the CTD console during the cruise and analyzed salinity samples for calibration.

Nutrient Measurements

Nutrient samples were collected from the Niskin bottles in acid-washed 35-ml polyethylene bottles after three complete seawater rinses and typically analyzed within 12 hours of sample collection. Nutrients were analyzed with a continuous flow analyzer (CFA) using the standard and analysis protocols for the WOCE hydrographic program as set forth in the manual by L.I. Gordon, et al (2000). Approximately 1900 samples were analyzed for phosphate (PO_4^{3-}), nitrate (NO_3^-), nitrite (NO_2^-), orthosilicic acid (H_4SiO_4), and ammonium (NH_4^+).

A mixed stock standard consisting of silicic acid, phosphate and nitrate was prepared at PMEL by dissolving high purity standard materials (KNO_3 , KH_2PO_4 and Na_2SiF_6) in deionized water using a two step dilution for phosphate and nitrate. This standard was stored at room temperature. Nitrite and ammonium stock standards were prepared about every 10 days by dissolving in distilled water, and these standards were stored in the refrigerator. Working standards were freshly made at each day by diluting the stock solutions in low nutrient seawater. The low nutrient seawater used for the preparation of working standards, determination of blank, and wash between samples was filtered seawater obtained from low-nutrient Pacific surface waters.

A typical analytical run consisted of distilled water blanks, standard blanks, working standards, a standard from the previous run, samples, replicates, and working standards, and standard and distilled water blanks. Four replicates were usually measured on each run, plus any samples with questionable peaks, and the overall precision of the analysis was within 1% of full range.

Table 1. Sampling by Hydrographic Group, 3-31 July 2008

Hydrographic Stations	177
CTD casts	210
Nutrient Samples Analyzed	1770
Winkler Oxygen Samples	263
DOC Samples	311
DIC Samples	311
O ¹⁸ Samples	296
Underway Samples	
Nutrient Samples	121
Winkler Oxygen Samples	96
Total Chlorophyll Samples	93

Oxygen Measurements

Winkler titrations were conducted according to WOCE protocols. On each cast, the number of samples and the depths sampled were dependent on the oxygen profile from the CTD. In deep water, samples were typically collected at every depth below 100m. On the shelf, samples were usually collected in the upper layer, and in the bottom layer. End point determinations of the Winkler titration were determined potentiometrically. Thiosulfate was standardized for each batch of sample titrations, and blanks were measured periodically during the cruise.

DIC and DOC Sampling

The sampling protocol for the DIC sampling was as follows: Samples were drawn into glass bottles immediately after oxygen sampling directly from the Niskin bottles using tubing to reduce the amount of bubbles entrained in the sample. The bottles were rinsed three times and then filled almost full. A little head space was allowed for gas expansion, but the level in the bottle was not below the white tape on the bottle neck. After the bottle was filled, it was injected with mercuric chloride. The lid was screwed on as tightly as possible, and the bottle shaken a little. Glass bottles were labeled with the station number, cast number and bottle number on each sample.

The sampling protocol for the DOC sampling was as follows: The plastic bottles were rinsed three times from the Niskin and then filled about 70% full. The caps were screwed on tight, labeled the same as the DIC samples and placed in a freezer for the duration of the cruise.

Ned Cokelet arranged for the underway seawater sampling system to be augmented for this cruise, and Scott Hiller set up the instruments. Additions included a WetLabs ac-9 optical absorption and attenuation meter and a Satlantic ISUS nitrate meter (both on loan from Lisa Eisner, NOAA Auke Bay Laboratory). Seawater samples were collected from the system and analyzed for dissolved oxygen, nitrate and chlorophyll concentration for calibration.

Typical Results

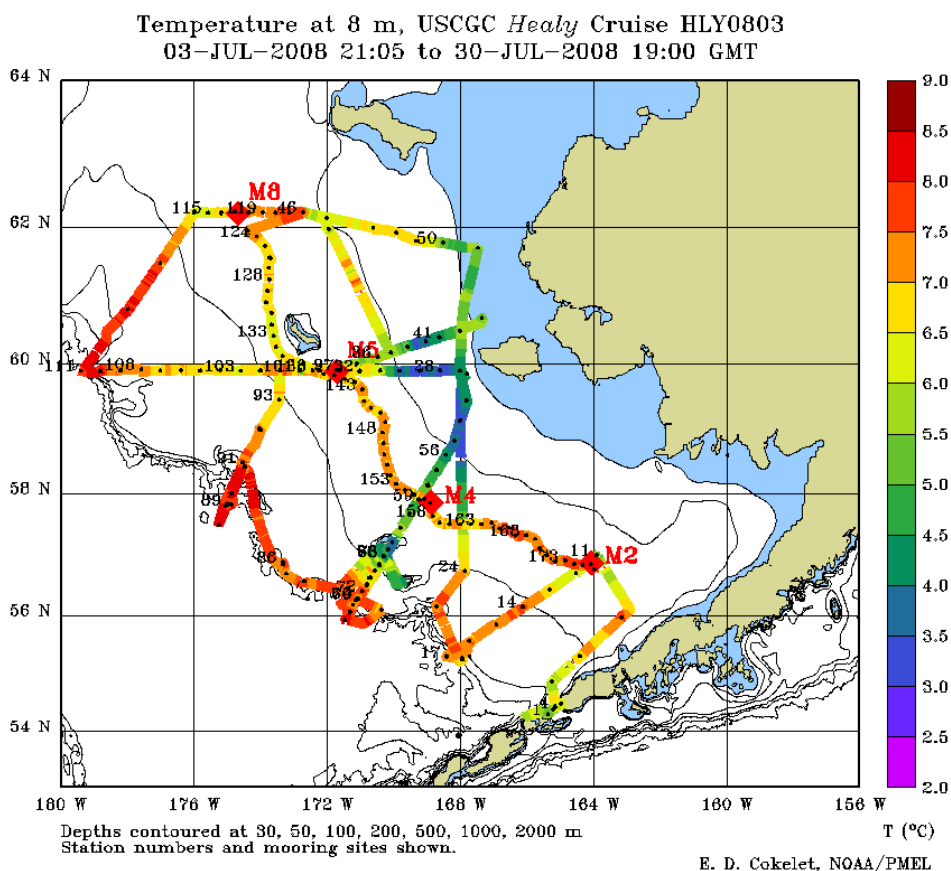
Figures 1-5 show the water temperature, salinity, nitrate, chlorophyll and oxygen concentrations as measured by the underway seawater sampling system. The

underway nitrate and oxygen values were calibrated against discrete samples taken and analyzed during the cruise. These measurements are preliminary and may change after further analysis.

Figure 6 shows the locations of various CTD transects taken during the cruise.

Figures 7-13 show the water temperature, salinity, density (sigma-t), photosynthetically available radiation (PAR), optical beam attenuation, chlorophyll and oxygen concentrations for the MN transect (Figure 6). These measurements are preliminary and may change after further analysis.

Figure Hydro-1. Water temperature at 8 m depth measured by the underway seawater sampling system.



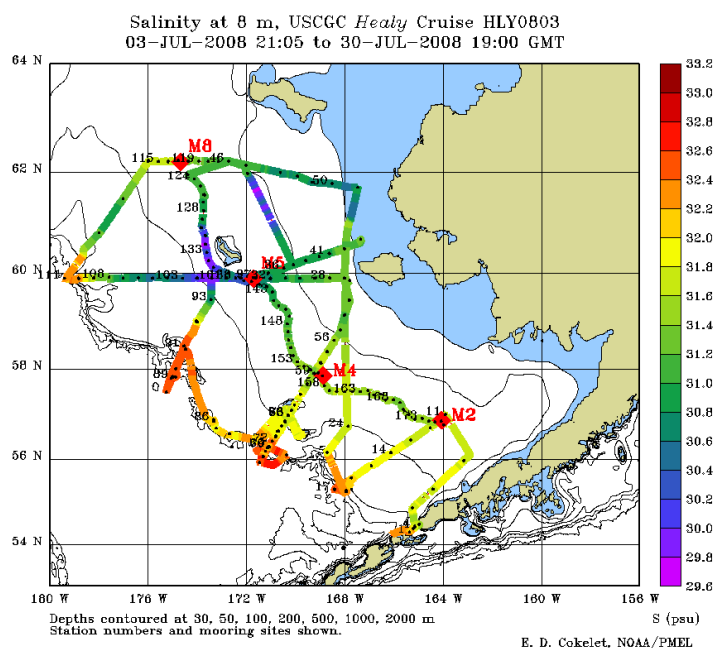


Figure 2. Salinity at 8 m depth measured by the underway seawater sampling system.

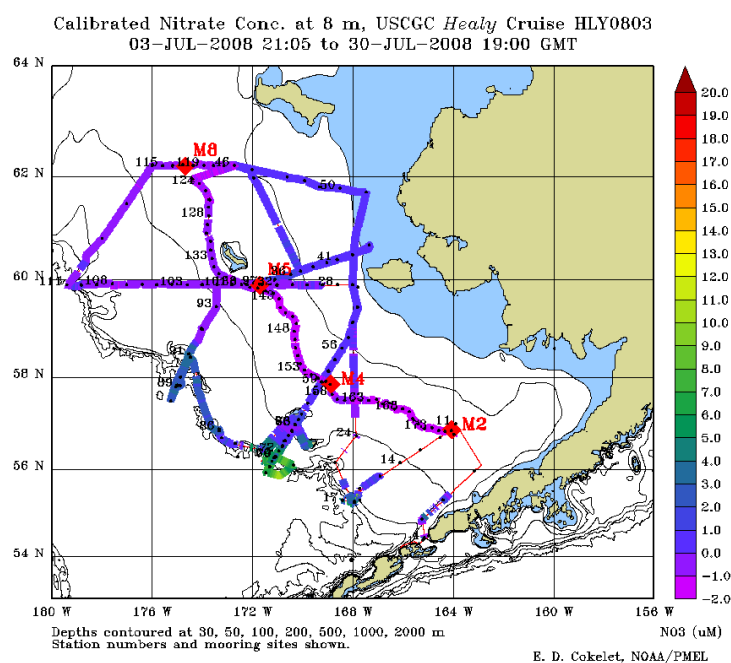


Figure Hydro-3. Calibrated dissolved nitrate concentration at 8 m depth measured by the underway seawater sampling system.

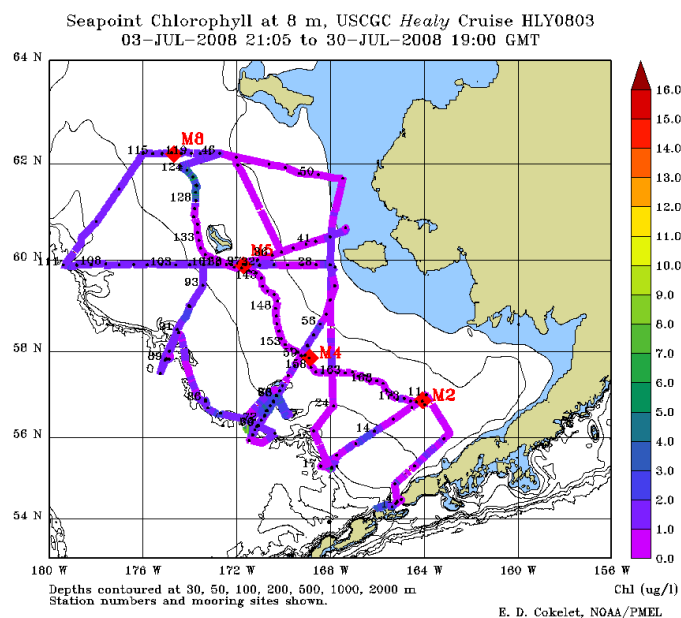
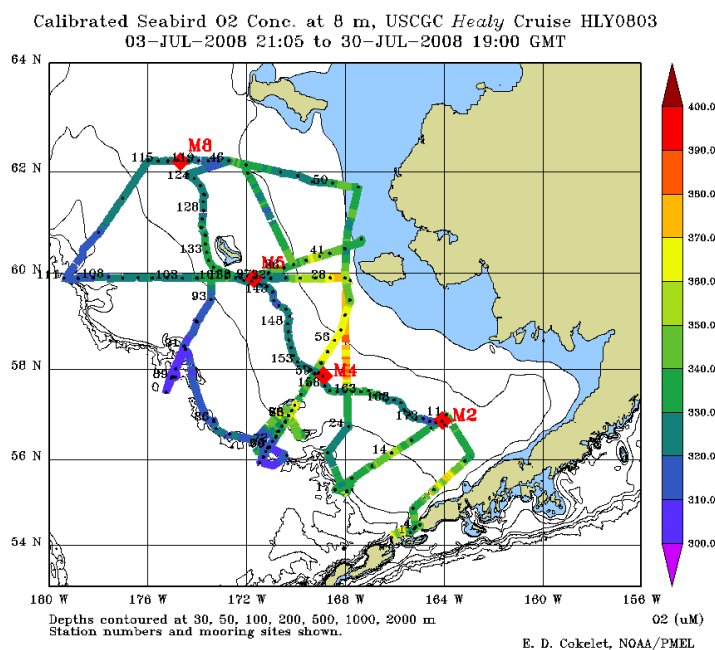


Figure Hydro-4. Chlorophyll concentration at 8 m depth measured by the underway seawater sampling system.

Figure Hydro-5. The calibrated dissolved oxygen concentration at 8 m depth measured by the underway seawater sampling system.



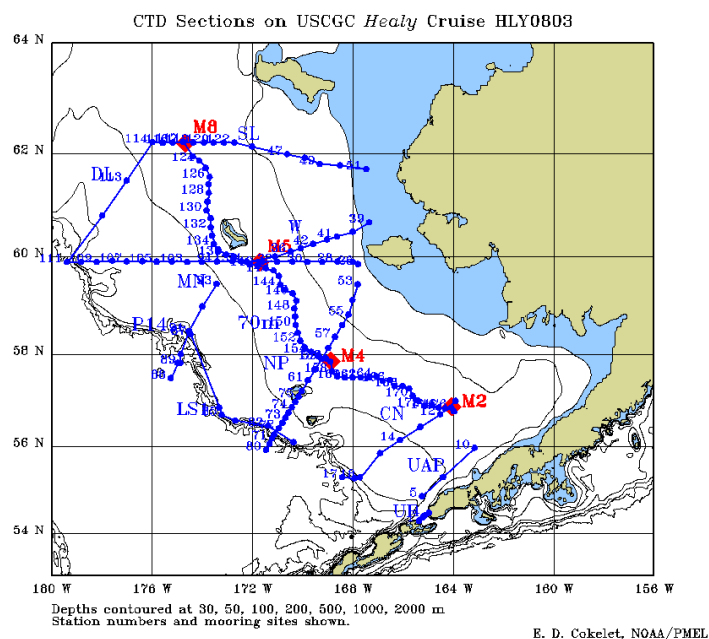


Figure Hydro-6. The locations of the SL, DL, 70-m, W, MN, P14, NP, LS1, NP, CN, UAP and UP CTD transects made during the cruise.

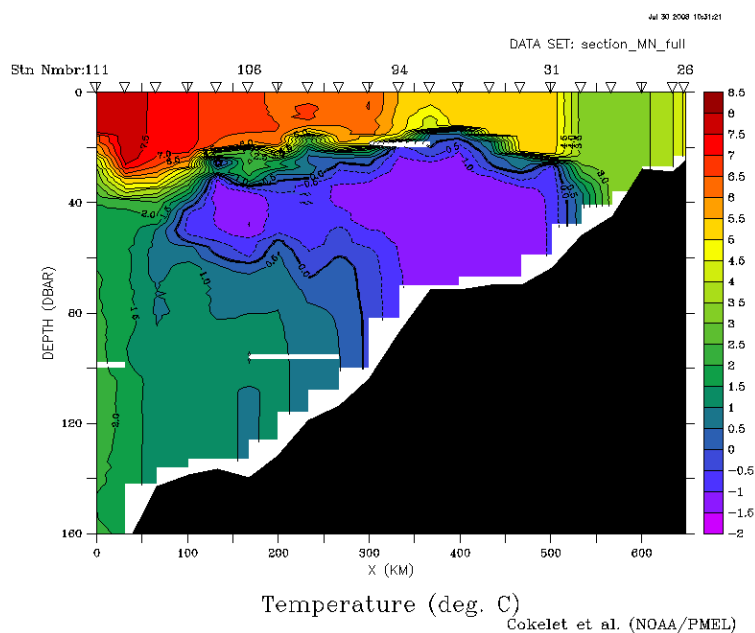


Figure Hydro-7. Water temperature along the MN transect.

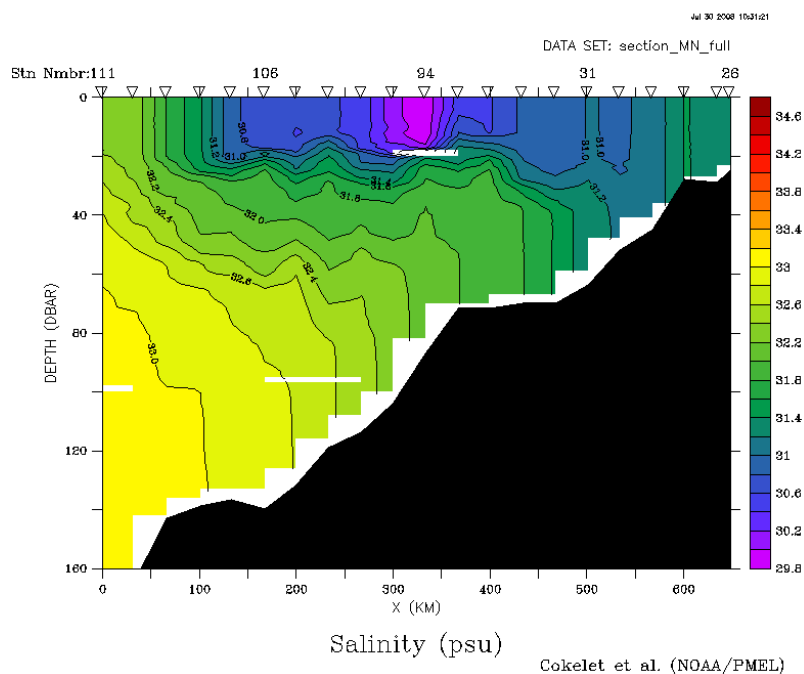


Figure Hydro-8. Salinity along the MN transect.

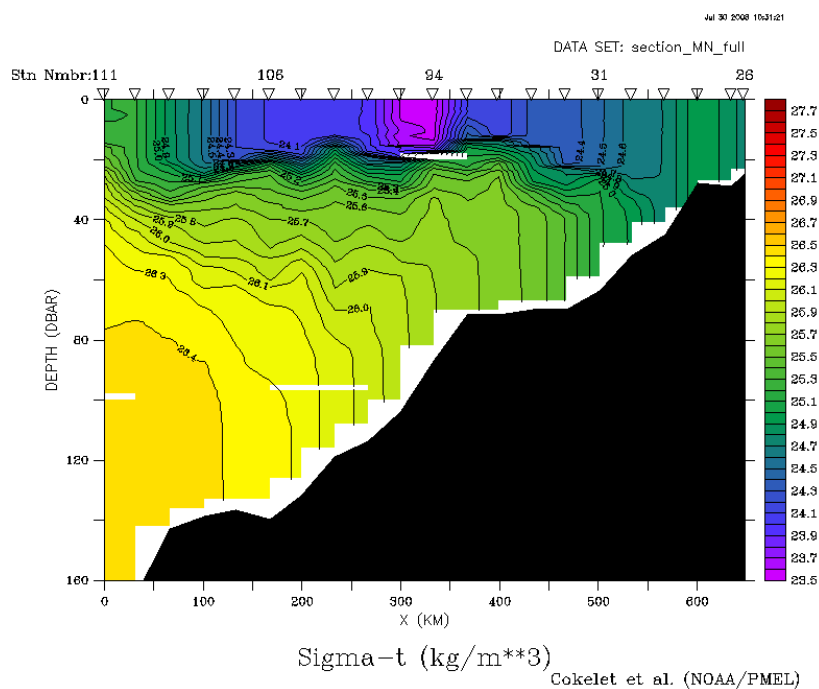


Figure Hydro-9. Sigma-t density along the MN transect.

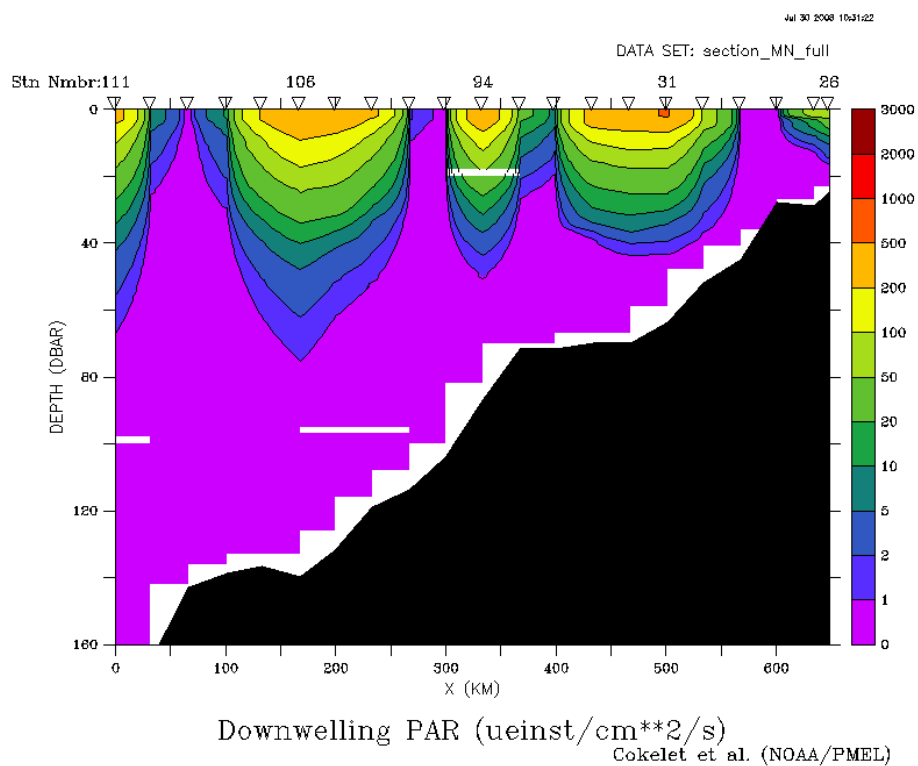


Figure Hydro-10. Photosynthetically Available Radiation (PAR) along the MN transect.

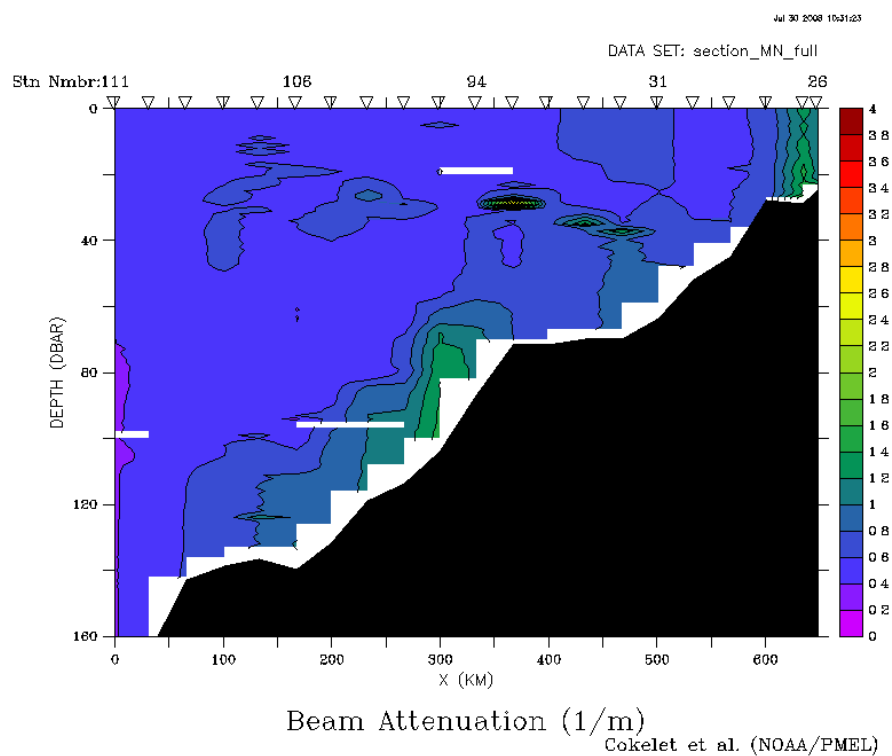


Figure Hydro-11. Optical beam attenuation along the MN transect.

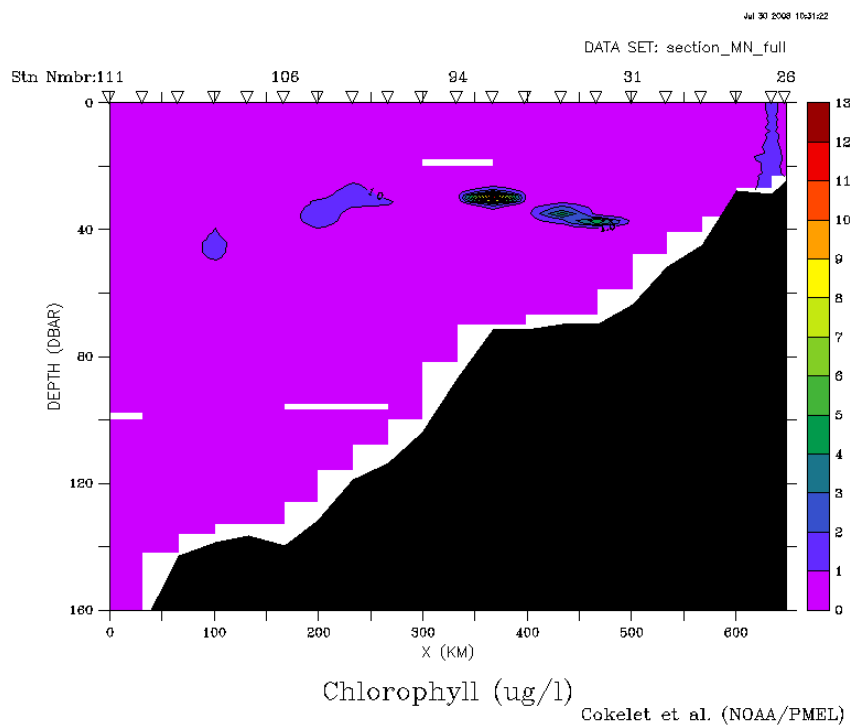


Figure Hydro-12. Chlorophyll concentration along the MN transect.

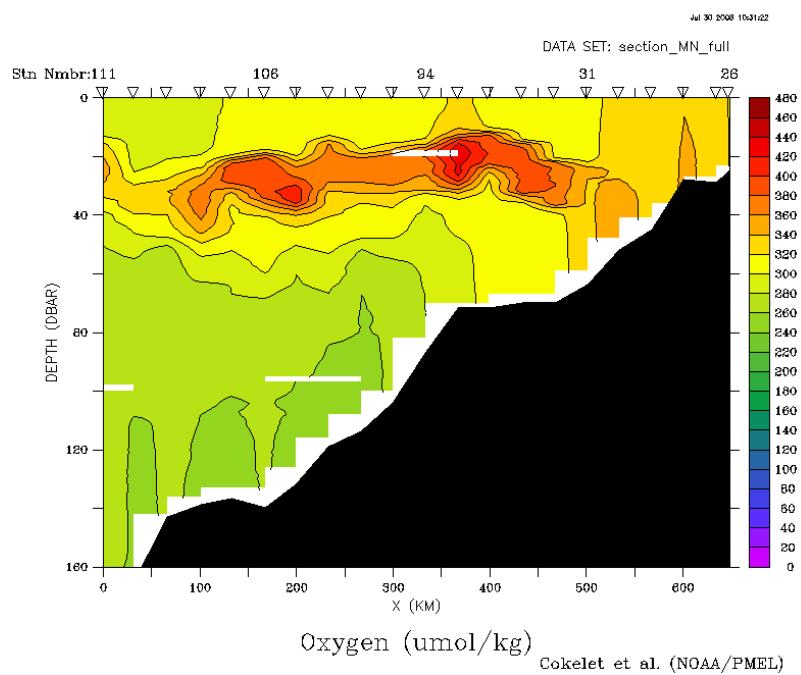


Figure Hydro-13. Dissolved oxygen concentration along the MN transect.

9.3 Carbon productivity – Lomas/ BIOS group

Component goals:

1. Quantify the magnitude and regional variability of gross primary production and net community production in MIZ and open-water blooms associated with seasonal and interannual changes in sea ice extent.
2. Quantify the main floristic patterns (using a diversity of chemotaxonomic methods) and autotrophic cell size distributions in MIZ and open-water blooms.
3. Quantify the export flux of organic carbon associated with MIZ and open-water blooms in deeper waters (outer-shelf/slope), and link carbon export to primary production and benthic oxygen utilization to assess the efficiency of pelagic-benthic coupling associated with seasonal and interannual changes in sea ice extent.

To achieve these objectives we have used the following methods:

- Obj 1. Net primary production was estimated by use of ^{14}C incubations. Taxon-specific carbon turnover will be estimated from samples taken in collaboration with Ray Sambrotto that will be sorted on a flow cytometer after returning to BIOS.
- Obj 2. We are documenting phytoplankton community structure by several methods. Microplankton ($>10\mu\text{m}$) are being quantified using traditional Utermohl setting chambers. We are estimating the C content of each cell (and after multiplying by cell abundance - Carbon in each population or functional group) using the Strathman equations and the Montagnes et al. equations using approximate geometric shapes for each species. Pico and Nano-plankton, and heterotrophic bacteria are being quantified using flow cytometry. Cell abundance is calculated based upon the volume analyzed method. Lastly, we are taking samples for HPLC analysis that will be converted to approximate taxonomic groups based upon the CHEMTAX algorithms.
- Obj 3. We are quantifying POC fluxes using surface-tethered traps and elemental analysis. We are also collecting samples for pigment analyses from the trap material to attempt to link phytoplankton community structure in the euphotic zone with the composition that is collected in the traps (with all appropriate caveats).

Expected Results are:

1. size-fractionated (whole and $>5\mu\text{m}$) chl *a* (fluorometry) and HPLC pigment concentrations, and size-fractionated rates of primary production.
2. estimates of *Synechococcus* abundance, pico- and nano-eukaryotes, and identification, abundances and carbon content of microphytoplankton.
3. Export flux estimates for POC and HPLC pigments.

Data availability:

1. Chl *a* and HPLC pigments, and primary production will be available by the end of the calendar year 2008.
2. Pico/nano-plankton analyses and a subset of the microplankton analyses will be completed by the end of 2008.

3. No samples for HLY0803 have been analyzed as yet, and therefore nothing to show at this time.

9.4 Nitrogen productivity & cycling – Sambrotto/ LDEO group

Project objectives:

- 1) Measure new (nitrate) and regenerated productivity during summer as a comparison to the spring measurements made on earlier BEST cruises.
- 2) Measure isotopic signatures of particulate carbon and nitrogen to determine their variations with production and regeneration.
- 3) Determine the seasonal development of the production in the regions that supported elevated levels of productivity during the earlier ice edge.

Nitrogen & carbon productivity rates measured:

Station	cast	latitude [degrees north]	longitude [degrees east]	Date
4	7	54.43817	194.8588	07/04/2008
11	12	57.00117	196.0948	07/05/2008
16	18	55.2805	192.0733	07/06/2008
21	24	55.25	192.0705	07/07/2008
29	33	59.89683	190.8008	07/09/2008
37	42	60.171	189.9333	07/10/2008
46	52	62.204	187.2988	07/12/2008
59	67	57.89867	190.7612	07/14/2008
66	76	56.28817	188.9352	07/15/2008
75	85	57.0925	189.8347	07/17/2008
79	90	56.27117	187.4248	07/18/2008
86	102	56.84867	186.6872	07/20/2008
89	107	57.82833	185.0075	07/21/2008
92	114	59.00667	185.9843	07/22/2008
98	122	59.89867	189.0002	07/23/2008
103	131	59.89367	185.4247	07/24/2008
110	140	59.89517	181.1935	07/25/2008
116	147	62.199	184.8383	07/26/2008
126	159	61.552	186.3098	07/27/2008
143	176	59.616	189.0673	07/28/2008

Expected results:

- 1) Spatial pattern of productivity.
- 2) Vertical pattern of productivity – Particularly the productivity associated with sub-surface Chl a layers.
- 3) Pattern of isotopic fractionation of marine particulate material and its relationship to productivity and regeneration.

9.5 Particle flux – URI group

Project Objectives:

- 1) Quantify the flux of particulate organic carbon (POC) from the surface water to the deep waters of the Bering Sea using ^{234}Th as a tracer of particle export.
- 2) Determine POC/ ^{234}Th ratio and phytoplankton community values for particles collected in drifting sediment traps at five different depths (25, 40, 50, 60, and 100m).
- 3) Estimate particle export using measurements of total ^{234}Th – ^{238}U disequilibrium in the water column determined using small-volume thorium extractions with a manganese oxide precipitate.
- 4) Estimate gross primary production within the surface mixed layer using triple-oxygen isotope method.
- 5) Estimate cross-shelf exchange and self-break front isopycnal mixing using short-lived radium isotopes ($^{223}, ^{224}\text{Ra}$).

Samples Collected:

Drifting sediment traps

Station	Depths (m)	POC/ ^{234}Th samples	Pigment samples
PIT 1	25,40,50,60,100	2 per depth	2 per depth
PIT 2	25,40,50,60,100	2 per depth	2 per depth
PIT 3	25,40,50,60,100	2 per depth	2 per depth

Small Volume ^{234}Th

Station	Depths (m)
4-UP3	10, 20, 30, 40, M60, 75, 100, 131
16-PITD-1	6, 12, 19, 24, 30, 46, 70, 100, 150, 200
29-MN3	10, 20, 30, 40
46-SL8	6, 18, 25, 30, 35, 45, 50
59-NP7	10, 20, 30, 40, 50, 62
67-PITD-2	10, 25, 40, 50, 60, 80, 100, 125, 150, 175, 200, 215
83-NP11	10, 20, 30, 40, 50, 66
86-LS1-6	10, 20, 30, 40, 50, 75, 100, 125, 161
87-PITD-3	10, 25, 40, 50, 60, 75, 100, 150, 170
92-P14-3	10, 20, 30, 40, 50, 75, 100, 120
103-MN12	10, 20, 30, 40, 50, 75, 107
111-MN20	10, 20, 30, 40, 50, 75, 100, 200, 300, 400, 500
116-SL14	10, 20, 30, 40, 50, 76
143-70M36	10, 20, 30, 38, 50, 67

Triple-oxygen isotopes

Date	Station	Depth (m)
7/4/08 13:07	4-UP3	10
7/5/08 11:47	10-UAP3	20
7/6/08 1:53	17-CN18	18
7/8/08 9:43	24-CNN6	32

7/9/08 20:09	31-MN5	20
7/10/08 13:15	35-W7	10
7/11/08 10:10	41-W3	20
7/12/08 10:06	46-SL8	12
7/12/08 10:06	46-SL8	35
7/13/08 6:37	51-N25	10
7/14/08 14:14	59-NP7	15
7/15/08 6:19	64-NP12	10
7/16/08 1:06	68-NP15	16
7/18/08 11:58	78-NP12.5	10
7/19/08 4:29	81-LS1-1	10
7/20/08 5:56	84-LS1-4	10
7/20/08 8:52	85-LS1-5	10
7/20/08 11:08	86-LS1-6	12
7/21/08 16:24	89-P14-4	15
7/22/08 11:32	92-P14-3	10
7/23/08 10:16	96-MN8	10
7/24/08 9:42	102-MN11	10
7/25/08 8:09	108-MN17	10
7/25/08 13:18	110-MN19	10
7/25/08 21:04	111-MN20	10
7/26/04 12:10	114-SL16	10
7/26/08 23:09	118-SL12	10
7/27/08 12:50	124-70M55	10
7/28/08 4:17	133-70M46	10
7/28/08 13:45	141-70M48	15
7/29/08 5:09	153-70M26	10
7/29/08 9:17	156-70M23	10
7/29/08 16:01	159-70M19	15
7/29/08 22:40	164-70M14	15
7/30/08 0:44	165-70M13	6
7/30/08 5:59	169-70M09	15
7/30/08 9:59	172-70M06	10
7/30/08 16:20	177-70M01	15

Short-lived Radium isotope samples

Station	Depth (m)
S-25	8
C-25	8
72-NP13.5	20
73-NP12.5	39
74-NP11.5	36
76-NP11	69
LS1-4	8

LS1-5	8
LS1-6	8
92-P14-2	26
93-P14-1	25
96-MN8	20
101-MN9B	28
101-MN9B	74

Data Summary

Trap Results:

Measurable quantities of ^{234}Th have been collected in the sediment traps, though further comment is unwarranted due to incomplete analysis. CHN and pigment analysis will be completed upon return to URI-GSO. Samples archived for pigment analysis may also be analyzed for $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ by Dr. Sambrotto, as well as organic biomarkers by Dr. Goes.

Small Volume Results:

Measurable quantities of ^{234}Th have been collected, though further comment is unwarranted due to incomplete analysis.

Triple Oxygen Productivity Results:

Samples will be analyzed post-cruise, no comment is warranted at this time.

Short Lived Radium Results:

Analysis of only sample collected up to this point revealed low activities of ^{223}Ra and ^{224}Ra , typical of open ocean values. Near shore, shallow water column samples had highest activities (~ 2 dpm/100L for ^{224}Ra , and 0.15 dpm/100L for ^{223}Ra). Near-bottom samples collected where the $25.5 \sigma_t$ isopycnal was in contact with the sediments had the next highest activities (0.7-0.8 dpm/100 L for ^{224}Ra , and 0.05 dpm/100 L for ^{223}Ra). Lowest radium activities were found where the $25.5 \sigma_t$ isopycnal shoaled (0.14-0.19 dpm/100L for ^{224}Ra and 0.007 dpm/100L for ^{223}Ra). These values are not corrected for supported values, which will be made after the cruise in the lab. However, these results indicate some advection of radium from the sediments along the $25.5 \sigma_t$ isopycnal. Values for longer-lived ^{228}Ra and ^{226}Ra will be determined at URI-GSO.

9.6 Iron Analysis - UAF group

PI: Jingfeng Wu

Participants: Ana Aguilar-Islas and Robert Rember

Cruise goals

The “Green Belt”, a band of high and sustained spring and summer productivity, is located along the shelf break in the transition zone between the iron-limited HNLC surface waters of the deep basin, and the iron-replete, but nitrate-limited surface shelf waters. Mesoscale eddies and the shelf break front have been suggested as

mechanisms that can promote mixing of outer shelf and slope waters, and lead to the input of nutrients into Green Belt waters. During the summer this additional input of nitrate can only be fully assimilated when enough iron is also available. In summer, surface outer shelf waters contain low (picomolar) surface dissolved iron concentrations, thus external sources of iron are needed to support the high and sustained productivity of the Green Belt. A possible mechanism for the introduction of iron into Green Belt waters is the outcropping of higher iron bottom shelf water via frontal processes.

To test our hypothesis that the shelf break front provides a mechanism for the input of iron into the Green Belt, we collected vertical profiles along this front. Three cross-shelf transects were selected (NP, P-14, and MN lines) to cover southern and northern portions of the shelf break. Our sampling was focused within isopycnals originating on bottom shelf waters and surfacing within the photic zone of the Green Belt.

Methods

Collection of seawater samples was focused from the outer shelf to offshore waters along the NP, P14, and MN lines. Trace metal-clean vertical profiles were collected using our UAF/ATE vane samplers at 15 stations (5 stations at each line). To avoid contamination from the ship, samples were filtered in a class 100 laminar flow hood using trace metal clean procedures. Samples were filtered through 0.4 μm PCTE filters, collecting triplicate 30 ml samples for total dissolved Fe measurements. The remaining unfiltered sample will be used for the analysis of total acid dissolvable Fe (pH 2). Samples will be acidified to pH 2 with the equivalent of 1 ml of 12 M hydrochloric acid (Optima grade, Fisher Scientific Inc.) per liter of sample. Acidified samples will be stored at room temperature for 1 month prior to Fe analysis using $\text{Mg}(\text{OH})_2$ co-precipitation isotope dilution ICPMS. This method has a detection limit of ~ 0.03 nM and a precision of $\sim 3\%$ at 0.3 nM Fe level.

Expected results

A well developed shelf break front, with isopycnals originating in the bottom layer of the outer shelf (near the 80 m isobath) and surfacing in the photic zone of the shelf break, was apparent at the NP, P-14, and MN lines. We expect the bottom waters of the outer shelf to be high in iron due to their proximity to sediment. This high iron water could serve as an iron reservoir throughout the summer. An inshore-offshore gradient in both total dissolved iron and total dissolvable iron (which includes labile particulate iron) is expected along the sampled isopycnals.

The vertical profiles taken along the NP, P-14, and MN lines provided high resolution samples along the isopycnals of interest. Concentrations of the two sampled iron species together with information from short and long lived Radium isotopes (samples collected and measured by the Moran lab) will provide data for quantifying the flux of iron to the Green Belt via the shelf break front. Nutrient and chlorophyll, and species composition data (historical and from this cruise) could then be used to estimate the biological iron requirement for maintaining a phytoplankton bloom in the Green Belt. The importance of the shelf break front as a mechanism for the delivery of additional iron to Green Belt waters can then be evaluated.

Total dissolved iron, and total dissolvable iron data will be available during the winter of 2008.

Sample Station Data Table

Station Name	Sample Depth (m)	Sigma t (Kg m ⁻³)	Total Dissolved Iron (nM)	Total Dissolvable Iron (nM)
P14-4	15 m	25.45		
	20 m	25.57		
	25 m	25.88		
	30 m	26.08		
	40 m	26.17		
	55 m	26.25		

9.7 Euphausiid and macrozooplankton collections – UAF & OSU group

Meso-Zooplankton Distribution and Abundance (A. Pinchuk)

The primary task of the zooplankton component was to assess the abundance, biomass and species composition of the mesozooplankton and micronecton on the shelf-break/outer, middle and inner shelf domains of the southeastern Bering Sea. The data from these samples will aid in determining the fate of new and recycled production on the shelf. A total of 58 MOCNESS tows were taken in shelf-break and outer shelf regime and in the middle and inner domains. We obtained 81 CalVET samples at all CTD stations along NP, MN, SL and W transect lines and at selected locations along 70M line.

The large mesozooplankton component was sampled using a 1-m MOCNESS (Multiple Opening Closing Net and Environmental Sensing System), equipped with 0.5 mm mesh nets. The MOCNESS was equipped with salinity, temperature and fluorescence sensors to provide depth profiles of physical oceanographic data during the tows. Samples were consistently taken in 20 m depth increments from 100 m or the bottom to the surface.

The small mesozooplankton were sampled with a 25 cm CalVET (CalCOFI Vertical Egg Tow) net equipped with 0.15 mm mesh nets. The net was towed vertically from the bottom to the surface and from 100 m to the surface at sites deeper than 100 m. The nets were equipped with General Oceanics digital flow meters to monitor volume filtered. The CTD sample number was recorded with each net to facilitate comparison of CalVET samples with physical oceanographic data.

Samples were preserved in 10% formalin seawater and returned to the lab for processing. Samples will be split and organisms identified to the lowest possible taxonomic category. Copepods will be staged and wet weights will be determined for each species and stage. The above procedure will generate the species composition, abundance and wet weight biomass for all identified taxa from each tow.

Casual observation of the samples indicates that oceanic zooplankton species were common in the shelf-break and outer shelf region, but large copepods were rare or absent from the middle and inner domains stations. It appears that the mesozooplankton community was dominated by medium-sized and small copepods, gelatinous zooplankton and, at some stations, euphausiids and hyperiids. Oceanic *Neocalanus* spp., *Eucalanus bungii* and *Thysanoessa longipes* were abundant on the offshore ends of NP and MN transects indicating advection of oceanic water on the outer shelf (up to ~100 m isobath). *Calanus marschallae* and

Thysanoessa raschii comprised bulk of zooplankton on the middle shelf, while *Sagitta elegans* and small copepod *Pseudocalanus* spp. were especially abundant in the inner domains. Cold water hyperiids *Parathemisto libellula* were common on the northern part of the shelf. Large numbers of juvenile scyphozoan *Chrysaora melanaster* (bell diameter <15 cm) were observed on the southern middle shelf. A detailed assessment of zooplankton abundance, biomass and distribution will be made after the samples have been processed.

Appendix 1 Summary of zooplankton survey during HLY0803

STATION	GEAR	LATITUDE	LONGITUDE	DEPTH(m)
UP1	MOCNESS	54.3166	165.3790	60
UP2	MOCNESS	54.3942	165.1927	100
UP4	MOCNESS	54.4803	164.9563	70
UP3	MOCNESS	54.4315	165.1535	100
UAP8	MOCNESS	54.8728	165.2534	100
UAP7	MOCNESS	55.0818	164.8218	90
UAP6	MOCNESS	55.3016	164.3954	80
UAP5	MOCNESS	55.5296	163.9713	75
UAP4	MOCNESS	55.7538	163.5531	80
UAP3	MOCNESS	55.9769	163.1440	70
CN6	MOCNESS	56.9994	163.8889	55
CN6	CalVET	56.9971	163.9112	68
CN8	CalVET	56.6967	164.5303	70
CN8	MOCNESS	56.6984	164.5296	55
CN10	CalVET	56.4313	165.2817	84
CN12	CalVET	56.1380	166.1061	100
CN14	CalVET	55.8476	166.9038	100
CN14	MOCNESS	55.8376	166.9099	100
CN18	CalVET	55.2916	168.3838	100
CN18	MOCNESS	55.3115	168.4010	100
CN16	MOCNESS	55.5479	167.7002	100
CN16	MOCNESS	55.5639	167.7036	100
CN16	CalVET	55.5611	167.7042	100
X-1	MOCNESS	55.3855	167.8542	100
X-2	MOCNESS	55.2482	167.9574	100
CNN8	CalVET	56.1424	168.7432	100
CNN8	MOCNESS	56.1540	168.7139	100
CNN7	MOCNESS	56.4949	168.2223	100
CNN6	CalVET	56.7338	167.8441	98
CNN6	MOCNESS	56.7602	167.8578	98
MN1	CalVET	59.8976	167.9899	28
MN1	MOCNESS	59.8936	167.9904	15
MN2	CalVET	59.9025	168.5884	38
MN2	MOCNESS	59.8822	168.5699	25
MN3	CalVET	59.8963	169.1956	44
MN3	MOCNESS	59.8871	169.1806	44
MN4	CalVET	59.8917	169.8089	54
MN4	MOCNESS	59.8839	169.8142	40
MN5	CalVET	59.9067	170.3914	60

MN5	MOCNESS	59.8947	170.4189	50
MN6	CalVET	59.8965	170.9852	69
MN6	MOCNESS	59.9006	170.9917	55
MN7	CalVET	59.9001	171.6130	74
MN7	MOCNESS	59.9021	171.6191	55
MN8	CalVET	59.8981	172.2310	72
MN8	MOCNESS	59.8919	172.2408	60
W7	CalVET	59.9920	171.1072	65
W6	CalVET	60.1014	170.4563	60
W3	CalVET	60.4003	168.6142	36
W4b	CalVET	60.2550	169.5659	47
SL6b	CalVET	61.9697	171.9436	54
SL7b	CalVET	62.1230	171.9957	51
SL8b	CalVET	62.2093	172.7101	54
SL5b	CalVET	61.9820	170.6009	48
SL5b	MOCNESS	61.9804	170.5979	48
SL4b	CalVET	61.9154	169.9003	49
SL3b	CalVET	61.7970	169.3037	41
SL3b	MOCNESS	61.8065	169.2862	41
SL2b	CalVET	61.7717	168.4877	33
NP1	CalVET	59.4453	167.7933	38
NP1	MOCNESS	59.4420	167.7869	38
NP2	CalVET	59.1272	168.0030	44
NP3	CalVET	58.8297	168.1597	47
NP3	MOCNESS	58.8304	168.1456	43
NP4	CalVET	58.6029	168.4110	55
NP5	CalVET	58.3596	168.6845	65
NP6	CalVET	58.1274	168.9586	70
NP7	CalVET	57.8982	169.2236	70
NP7	MOCNESS	57.8910	169.2132	70
NP8	CalVET	57.6729	169.5173	70
NP9	CalVET	57.4448	169.7786	64
NP9	MOCNESS	57.4431	169.7757	50
NP10	CalVET	57.2056	170.0244	41
NP10	MOCNESS	57.2030	170.0151	30
NP11	CalVET	56.9732	170.2705	72
NP11	MOCNESS	56.9668	170.2677	72
NP12	CalVET	56.7386	170.5473	100
NP12	MOCNESS	56.7343	170.5479	100
NP13	CalVET	56.5140	170.7914	100
NP13	MOCNESS	56.5017	170.7977	100
NP14	CalVET	56.2876	171.0606	100
NP14	MOCNESS	56.2813	171.0665	100
NP15-B	CalVET	56.0535	171.3025	100
NP 12.5	MOCNESS	56.6226	170.6794	100
NP 11.5	MOCNESS	56.8537	170.4136	85
NP10.5	MOCNESS	57.0871	170.1698	25
NP12.5	CalVET	56.6267	170.7015	100

NP12.5	MOCNESS	56.6222	170.7347	100
LS3	CalVET	56.4464	171.3597	100
LS3	MOCNESS	56.4429	171.3662	100
LS1-6	CalVET	56.8826	173.3031	100
LS1-6	MOCNESS	56.8715	173.3306	100
PI-4-4.5	CalVET	57.8023	175.0217	100
PI-4-4.5	MOCNESS	57.8076	175.0254	100
PI4-2	CalVET	58.9893	173.9856	100
PI4-2	MOCNESS	58.9997	174.0014	100
MN10	CalVET	59.9076	173.4077	84
MN9	CalVET	59.9094	172.7834	70
MN8	CalVET	59.8962	172.1926	70
MN7	CalVET	59.8935	171.5962	71
MN6	CalVET	59.8962	171.0020	70
MN5	CalVET	59.9008	170.4057	62
MN11	CalVET	59.8975	174.0033	100
MN11	MOCNESS	59.8994	173.9972	85
MN12	CalVET	59.8938	174.5808	100
MN13	CalVET	59.9012	175.1943	100
MN14	CalVET	59.8955	175.7924	100
MN15	CalVET	59.9088	176.3594	100
MN16	CalVET	59.9036	176.9836	100
MN17	CalVET	59.9230	177.5628	100
MN18	CalVET	59.8974	178.1806	100
MN19	MOCNESS	59.8858	178.8088	100
MN19	CalVET	59.8926	178.8108	100
MN20	CalVET	59.8940	179.3797	100
SL16	CalVET	62.1996	175.9632	93
SL15	CalVET	62.1920	175.5612	86
SL14	CalVET	62.1953	175.1642	80
SL13	CalVET	62.2132	174.7502	71
SL12	CalVET	62.2028	174.3359	68
SL11	CalVET	62.1971	173.9059	62
SL10	CalVET	62.1952	173.5242	61
SL9	CalVET	62.1958	173.1153	64
SL8	CalVET	62.1967	172.7046	55
70m-56	CalVET	61.9413	174.3645	69
70m-53	CalVET	61.5568	173.7013	69
70m-50	CalVET	61.0766	173.7683	78
70m-47	CalVET	60.5824	173.6563	69
70m-44	CalVET	60.1121	173.3207	69
70m-23	MOCNESS	57.9083	169.0708	55
70m-22	MOCNESS	57.8451	168.8898	55
70m-20	MOCNESS	57.6334	168.8104	55
70m-6	MOCNESS	56.9823	165.0145	55
70m-5	MOCNESS	56.9118	164.8361	55
70m-4	MOCNESS	56.8542	164.5716	55
70m-3	CalVET	56.8321	164.3117	73

70m-2	CalVET	56.8493	164.0662	69
70m-1	CalVET	56.7649	163.9609	69

9.8 Euphausiid rate measurements – Lessard/ Harvey/UW group

The Trophic Role of Euphausiids in the eastern Bering Sea: Ecosystem Responses to Changing Sea-Ice Conditions (Lessard and Harvey)

Lessard field team: Tracy Shaw, Megan Bernhardt and Gigi Engel

Harvey field team: Rachel Pluethner, Karen Taylor and Charlie Morgan

The goal of our project is to understand how climatically-driven changes in sea-ice conditions may affect the ecology and population dynamics of euphausiids in the eastern Bering Sea. Our primary hypothesis is that seasonal and interannual variation in the timing and coverage of sea-ice and associated food resources will lead to differences in age structure, diet history, and nutritional condition for euphausiids, which ultimately translate into differences in production rates and availability as prey to higher trophic levels. To determine diet, nutritional condition, and feeding rates, we are performing shipboard krill feeding experiments to measure ingestion rates of specific prey taxa (phytoplankton, heterotrophic protists, copepods) and we are determining the lipid profiles of both euphausiids and prey field. We are also isolating and culturing specific prey species to identify prey biomarkers. Identifying the lipid profiles and specific biomarkers for different prey taxa (particularly the poorly known heterotrophic protists) will enable us to infer diets from lipid profiles of field-caught euphausiids. We are also measuring euphausiid growth and egg production rates and estimating euphausiid age using the lipofuscin method. Our colleague, Alexei Pinchuk, will conduct laboratory rearing to allow calibration of the lipofuscin aging method when eggs can be collected in the field.

Bongo net tows

We performed 25 Bongo tows (Table 1) to capture live euphausiids for feeding and growth experiments and for lipid, carbon and lipofuscin analyses. The nets were towed obliquely.

Feeding experiments

We performed seventeen feeding experiments (Table 2, Figure 1) across the Bering Sea shelf and slope. For the feeding experiments, we captured live euphausiids with a Bongo net and added known numbers and species to bottles filled with seawater and incubated, along with control bottles without animals, for 24 hours on a rotating wheel in a flowing seawater incubator. The prey for each experiment were unaltered seawater plankton or seawater supplemented with concentrated seawater targeting the addition of protists. Shipboard, herbivorous feeding was assessed by measuring changes in size-fractionated chlorophyll. Samples were also fixed for microscopic counts of phytoplankton and heterotrophic protists to be analyzed back in the laboratory.

Growth experiments

We performed eight growth experiments, assessing instantaneous growth rates on 392 euphausiids (Table 3). Euphausiids were captured with a Bongo net and one euphausiid was added to a sideflask filled with 200µm filtered seawater captured from the CTD. The euphausiids were checked twice a day for molts for two days. We provided >700 euphausiids with species and size determinations, from the feeding and growth experiments, to Harvey for lipid profiles, carbon and lipofuscin content (an index of age).

Starvation experiment

We performed one starvation experiment to assess what lipids are stored or changed within euphausiids over time without the presence of food. Two 10L carboys were filled with filtered seawater and twenty krill were put into each 10L carboy. An additional 10 krill were retained initially for analysis. Carboys were incubated in the dark in a climate control chamber. Each week for two weeks, 5 krill were taken from each carboy. Experiment was to run for 4 weeks but a failed incubator ended the experiment prematurely.

Preliminary findings

Feeding and growth experiments were done under varying conditions, from inner shelf to slope, low and high chlorophyll and varying euphausiid species and sizes. As expected, the dominant euphausiid species on the mid to inner shelf was *Thysanoessa raschii*, with *Thysanoessa inermis* dominating on the outer shelf. Chlorophyll was typically low (<1.5µg/L) throughout our feeding experiments and dominated by small (< 5 µm) pico- and nanoplankton (cyanobacteria, picoeukaryotes, small flagellates), with heterotrophic protists (dinoflagellates and ciliates) present in modest numbers. Based on chlorophyll readings, herbivorous feeding by euphausiids was not always detectable with low initial chlorophyll. When initial chlorophyll was higher (>2µg/L), herbivorous feeding was detected. Experiments conducted with the high initial chlorophyll were mostly diatom dominant communities. Detection of feeding by euphausiids was most evident in the >5µg/L chlorophyll fraction despite total chlorophyll concentration. Final euphausiid feeding rates will be determined with the counting of the fixed samples in the laboratory. This process will take a significant amount of time as each individual organism, both phytoplankton and microzooplankton, will be enumerated and sized for biomass.

Station #	Station Name	Cast #	Latitude (decimal)	Longitude (decimal)	Date (local)	Time (local)	Station Depth	Calculated tow depth	Surface temperature (°C)	Surface chl (µg/l)	Salinity	Air temp (°C)	Feeding expt	IGR
4	UP3	1	54.4225	165.1481	7/4/08	0254	137	85	5.5	0.6	31.98	6.5		
10	UAP3	2	55.9673	163.1394	7/5/08	0400	88	57	7.6	0.6	31.78	6.98		
14	CN12	3	56.1390	166.1075	7/6/08	0250	110	64	7.72	2	31.86	6.9	1	
18	CN16	4	55.5626	167.7029	7/7/08	0310	136	71	7.51	0.8	32.06	7.1	2	11
24	CNN6	5	56.7344	167.8446	7/8/08	0220	100	78	6.9	0.4	31.7	7.2		
28	MN2	6	59.9025	168.5882	7/9/08	0230	41	32	3.4	0.6	31.38	4.1	3	
35	W7	7	60.0033	171.0919	7/10/08	0540	70	60	5.6	0.6	30.86	4.2	4	12
40	W2	8	60.4936	168.0000	7/11/08	0005	30	21	5.9	1.8	31.21	4.5		
41	W3	9	60.4001	168.6204	7/11/08	0232	35	28	4.3	0.7	30.92	4.2	5	13
46	SL8	10	62.2098	172.7108	7/12/08	0245	58	43	5.9	0.2	31.15	5.5		
52	EL2	11	61.0475	167.7863	7/13/08	0200	26	21	5.7	0.8	31.25			
58	NP6	12	58.1306	168.9698	7/14/08	0410	70	39	5.2	0.9	31.58	5.8	6	14
65	NP13	13	56.5145	170.7919	7/15/08	0215	122	71	7	1.8	32.15	7.3	7	
69	EL3	14	56.2783	170.7919	7/16/08	0445	142	85	7.7	1	32.38	5.7		
75	NP10.5	15	57.0929	170.1545	7/17/08	0220	39	30	3.9	2	31.8	5.8		
78	NP12.5	16	56.6268	170.7006	7/18/08	0430	116	85	6.24	3.27	31.84	6.75	8	15
82	LS1-3	17	56.4526	171.3680	7/19/08	0330	152	85	7.7	1.35	32.33	7.4	9	
86	LS1-6	18	56.8822	173.3032	7/20/08	0300	160	92	7.74	1.87	32.09	7.6	10	
89	P14-4	19	57.8024	175.0218	7/21/08	0315	2665	92	7.9	1.26	32.56	8.16	11	16
92	P14-2	20	58.9893	173.9856	7/22/08	0407	126	89	7.1	1.6	31.88	7.69	12	
96	MN8	21	59.8965	172.1976	7/23/08	0249	73	57	6.27	1.45	30.25	5.8	13	17
102	MN11	22	59.8982	174.0092	7/24/08	0308	101	85	7.3	1.51	29.97	6.9	14	
110	MN19	23	59.8975	178.8003	7/25/08	0457	157	85	8.01	1.92	32.29	8.3	15	18
114	SL16	24	62.2005	175.9653	7/26/08	0434	94	78	6.48	1.84	31.69	5.8	16	
123	70m56	25	61.9402	174.3643	7/27/08	0330	74	60	7.06	2.04	31.12	7.06	17	

Table Zoopl.-1

Station #	Station Name	Cast #	Latitude (decimal)	Longitude (decimal)	Date (local)	Time (local)	Station Depth	Calculated tow depth	Surface temperature (°C)	Surface chl (µg/l)	Salinity	Air temp (°C)	Feeding expt	IGR
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14	CN12	3	56.1390	166.1075	7/6/08	0250	110	64	7.72	2	31.86	6.9	1	
18	CN16	4	55.5626	167.7029	7/7/08	0310	136	71	7.51	0.8	32.06	7.1	2	11
24	CNN6	5	56.7344	167.8446	7/8/08	0220	100	78	6.9	0.4	31.7	7.2		
28	MN2	6	59.9025	168.5882	7/9/08	0230	41	32	3.4	0.6	31.38	4.1	3	
35	W7	7	60.0033	171.0919	7/10/08	0540	70	60	5.6	0.6	30.86	4.2	4	12
40	W2	8	60.4936	168.0000	7/11/08	0005	30	21	5.9	1.8	31.21	4.5		
41	W3	9	60.4001	168.6204	7/11/08	0232	35	28	4.3	0.7	30.92	4.2	5	13
46	SL8	10	62.2098	172.7108	7/12/08	0245	58	43	5.9	0.2	31.15	5.5		
52	EL2	11	61.0475	167.7863	7/13/08	0200	26	21	5.7	0.8	31.25			
58	NP6	12	58.1306	168.9698	7/14/08	0410	70	39	5.2	0.9	31.58	5.8	6	14
65	NP13	13	56.5145	170.7919	7/15/08	0215	122	71	7	1.8	32.15	7.3	7	
69	EL3	14	56.2783	170.7919	7/16/08	0445	142	85	7.7	1	32.38	5.7		
75	NP10.5	15	57.0929	170.1545	7/17/08	0220	39	30	3.9	2	31.8	5.8		
78	NP12.5	16	56.6268	170.7006	7/18/08	0430	116	85	6.24	3.27	31.84	6.75	8	15
82	LS1-3	17	56.4526	171.3680	7/19/08	0330	152	85	7.7	1.35	32.33	7.4	9	
86	LS1-6	18	56.8822	173.3032	7/20/08	0300	160	92	7.74	1.87	32.09	7.6	10	
89	P14-4	19	57.8024	175.0218	7/21/08	0315	2665	92	7.9	1.26	32.56	8.16	11	16
92	P14-2	20	58.9893	173.9856	7/22/08	0407	126	89	7.1	1.6	31.88	7.69	12	
96	MN8	21	59.8965	172.1976	7/23/08	0249	73	57	6.27	1.45	30.25	5.8	13	17
102	MN11	22	59.8982	174.0092	7/24/08	0308	101	85	7.3	1.51	29.97	6.9	14	
110	MN19	23	59.8975	178.8003	7/25/08	0457	157	85	8.01	1.92	32.29	8.3	15	18
114	SL16	24	62.2005	175.9653	7/26/08	0434	94	78	6.48	1.84	31.69	5.8	16	
123	70m56	25	61.9402	174.3643	7/27/08	0330	74	60	7.06	2.04	31.12	7.06	17	

Table Zoopl.-2

Expt #	Station Number	Station Name	Start Date	Species	Stages	# days incubated	# animals	# molts	Surface Chl (µg/L)	Surface Temp (°C)	Depth (m)
				<i>T. spinifera</i> &							
11	18	CN16	7/7/08	<i>T. inermis</i>	adults	2	45	10	0.80	7.51	136
12	35	W7	7/10/08	<i>T. raschii</i>	adults	2	50	10	0.60	5.60	70
13	41	W3	7/11/08	<i>T. raschii</i>	adults	2	50	11	0.70	4.30	35
14	58	NP6	7/14/08	<i>T. raschii</i>	adults	2.5	50	5	0.90	5.20	70
15	78	NP12.5	7/18/08	<i>T. inermis</i>	adults	2.5	49	7	3.27	6.24	116
16	89	P14-4	7/21/08	<i>T. inermis</i>	adults	3	50	4	1.26	7.90	2665
17	96	MN8	7/23/08	<i>T. raschii</i>	adults	2.5	50	34	1.45	6.27	73
18	110	MN19	7/25/08	<i>T. inermis</i>	adults	2.5	48	12	1.92	8.01	157

Table Zoopl.-3

9.9 Organic tracers of trophics/ euphausiid age structure – Harvey/ UMD group

Karen Taylor, Charlie Morgan, and Rachel Pleuthner, University of Maryland, Chesapeake Biological Labs, MOGEL Lab

- KRILL GRAZING AND AGING EXPERIMENTS
- **Water Column Particles and Krill Collection**
- Grazing Experiments for Determination of Euphausiid Grazing Rates and Food Source Preferences

Grazing experimental setup is detailed in the report from Lessard. For characterization of food resources and tracking of consumption, euphausiids were collected and incubated over a 24-hour period in CTD water from the same station. Water from designated Niskin bottles was filtered at T_0 through combusted GF/F filters for carbon and detailed lipid analysis to characterize the algal and detrital food available to krill (Table 1). At the conclusion of each of the seventeen grazing experiment conducted by the Lessard group, water from the four bottles containing animals was combined and filtered onto particulate filters for lipid analysis to compare food amounts and potential for selective grazing. Water from each of the control bottles was treated likewise.

Three different grazing experiments aimed to investigate food source preference through the addition of a volume of concentrated protists. Refer to Table 2 for sample inventory.

At the beginning of each grazing experiment, ten animals -with the exception of Experiment #16 which only has 5 animals - were collected from the bongo tow and individually frozen in the -80°C to serve as T_0 samples. Euphausiids incubated for 24 hours were also individually frozen (T_{final} animals). The frozen samples will be taken back to the lab for lipid analysis using a Gas Chromatograph coupled with Mass Spectrometry. The results will illustrate the types and quantities of organisms that euphausiids utilize for food

Lipid Storage and Metabolism Via Starvation Experiments

Long term lipid storage and metabolism will be examined by means of a starvation experiment initiated at the beginning of this cruise. Animals were collected at the beginning of the cruise with the intention of running a long-term incubation in filtered sea water. Ten animals were taken at T_0 and ten sub-sampled weekly to observe the gradual change in euphausiid lipid content. An environmental refrigerator malfunction, coincidentally at the Week 2 time point, increased the temperature to 68°F . Since the animals had survived the temperature increase, the experiment was moved into the refrigerators in the aft staging area. At the Week 3 time point, that refrigerator's temperature had jumped to 50°C . None of the animals survived; hence, the experiment ended prematurely.

The frozen samples will be taken back to the lab for lipid analysis using a Gas Chromatograph coupled with Mass Spectrometry.

Total Energy Content

Animals of a wide range of body lengths were frozen, 95 in total, to measure total carbon content using calorimetry.

Determination of Age in Euphausiids Found in the Bering Sea

Eight growth experiments were performed over the course of the cruise; the animals from these experiments were also used for lipofuscin measurements. These experiments have included animals of a large size range to provide a first estimate of lipofuscin indices in field animals of differing ages. Alexei Pinchuk will conduct growth experiments, when euphausiid eggs are made available, that will span a minimum of two years in order to allow age calibration of the field specimens that have been analyzed.

At the conclusion of the first growth experiment, the experiment animals were sacrificed. The eyes and eye stalks were removed; both the lipofuscin (Part A) and protein content (Part B) in each pair of eyes was extracted and quantified via flow-through fluorescence using an Agilent 1100 HPLC. This preliminary analysis of the first growth experiment could not be completed due to the malfunction of the fluorescence flow cell, for which there was no replacement. Euphausiids from subsequent experiments were frozen in the -80°C chest freezer for future lipofuscin analysis.

Lipofuscin Sample Analysis

High Performance Liquid Chromatography for the Identification and Quantification of Lipofuscin - Part A

Toward the beginning of the HLY0802 cruise, the optimal excitation and emission wavelengths for lipofuscin - an oxidation product that accumulates in euphausiid neural tissue - from *Thysanoessa inermis* was determined by running a three-dimensional fluorescent scan of the extracted product present in a composite of samples of krill neural tissue (see Figure 1). That scan allowed a qualitative identification of lipofuscin for that species, and was used to measure lipofuscin content in euphausiids for the duration of the cruise. A calibration curve using quinine sulfate allowed quantitative measurements of fluorescence intensity to be performed for each run.

Part B

For protein analysis of krill neural tissues, tryptophan fluorescence – a proxy for protein quantification – was measured using known excitation and emission wavelengths. Bovine Serum Albumin (BSA) was used to generate a calibration curve, which acted as a means to quantify protein in the eye tissues.

During protein analysis (Part B) for Growth Experiment 11 (a continuation of the numbering system from HLY0802), the HPLC fluorescence flow cell began to leak. Timing did not allow for the shipment of a replacement, preventing sample runs for the remainder of the cruise. Consequently, all animals used for lipofuscin analysis were frozen.

Using the age calibration established from Alexei's animals, the euphausiid age population can be examined both seasonally (winter and spring) and interannually (2008-2009). This information will bring insight into the condition of the krill population.

ORGANIC TRACERS

Water Column Particles and Sediment Collection

Investigating the input and preservation of bacterial derived organic matter

Sediment cores were collected as detailed in the report from David Shull. Surface sediment (0-2 cm) and downcore samples (sliced in 1 cm increments) were obtained from thirteen stations (Table 4) and frozen in I-Chem jars for the characterization of bacterial derived organic matter using intact bacteriohopanepolyols (BHPs). Intact BHPs are recalcitrant membrane lipids and thus considered ideal biomarkers for tracing microbial sources of organic carbon, and potentially bacterial populations and processes within marine environments. To trace the remnants of the spring bloom and also compare recent inputs and preservation of intact BHPs in the Bering Sea, water sampled below the chlorophyll maximum was taken from Niskin bottles and filtered through combusted GF/F filters for carbon and lipid analysis. Reversed-phase high-performance liquid chromatography-atmospheric pressure chemical ionization tandem mass spectrometry (RP-HPLC-APCI-MSn) will be applied for the analysis of intact BHPs, and all work will be completed at the University of Maryland Center for Environmental Science Chesapeake Biological Laboratory.

FIGURES AND TABLES

Table 1: Water Column Sample Collection for Experiments. This table lists the T₀ and T_f water collection in conjunction with the grazing experiments

Grazing Experiment	Station, No.	T ₀ filter date	T _f filter date	Added Nutrition
1	CN-14, #12	7/6/2008	7/7/2008	None
2	CN-16, #18	7/7/2008	7/8/2008	None
3	MN-2, #28	7/9/2008	7/10/2008	None
4	W-7, #35	7/10/2008	7/11/2008	None
5	W-3, #41	7/11/2008	7/12/2008	None
6	NP-6, #58	7/14/2008	7/15/2008	Protists
7	NP-13, #65	7/15/2008	7/16/2008	None
8	NP-12.5, #78	7/18/2008	7/19/2008	None
9	LS-3, #82	7/19/2008	7/20/2008	None
10	LS1-6, #86	7/20/2008	7/21/2008	Protists
11	P14-4, #89	7/21/2008	7/22/2008	None
12	P14-2, #92	7/22/2008	7/23/2008	None

13	MN-8, #96	7/23/2008	7/24/2008	None
14	MN-11, #102	7/24/2008	7/25/2008	Protists
15	MN-19, #110	7/25/2008	7/26/2008	None
16	SL-16, #114	7/26/2008	7/27/2008	None
17	70m-56, #123	7/27/2008	7/28/2008	None

*All filters frozen in the -80 immediately after particle collection

Table 2: Grazing Experiment Animal Log

This table lists all animals collected in grazing experiments, barring T₀ animals.

Experiment No.	No. Animals in the Incubation	Dominant Species	Storage date
Grazing Experiment #1	17	<i>T. inermis</i>	7/7/2008
Grazing Experiment #2	16	<i>T. spinifera</i>	7/8/2008
Grazing Experiment #3	16	<i>T. raschii</i>	7/10/2008
Grazing Experiment #4	32	<i>T. raschii</i>	7/11/2008
Grazing Experiment #5	31	<i>T. raschii</i>	7/12/2008
Grazing Experiment #6	24	<i>T. raschii</i>	7/15/2008
Grazing Experiment #7	16	<i>T. inermis</i>	7/16/2008
Grazing Experiment #8	16	<i>T. inermis</i>	7/19/2008
Grazing Experiment #9	16	<i>E. pacifica</i>	7/20/2008
Grazing Experiment #10	16	<i>E. pacifica</i>	7/21/2008
Grazing Experiment #11	16	<i>T. inermis</i>	7/22/2008
Grazing Experiment #12	16	<i>T. raschii</i>	7/23/2008
Grazing Experiment #13	16	<i>T. raschii</i>	7/24/2008
Grazing Experiment #14	16	<i>T. raschii</i>	7/25/2008
Grazing Experiment #15	16	<i>T. inermis</i>	7/26/2008
Grazing Experiment #16	16	<i>T. raschii</i>	7/27/2008
Grazing Experiment #17	16	<i>T. raschii</i>	7/28/2008

Table 3: HPLC Sample Run Log

This table lists the full set of HPLC sample runs performed throughout the entire cruise, barring calibration curves. A leaking HPLC flow cell prevented full analysis of Growth Experiment #11 and any experiment thereafter. Refer to Lessard report for the corresponding stations.

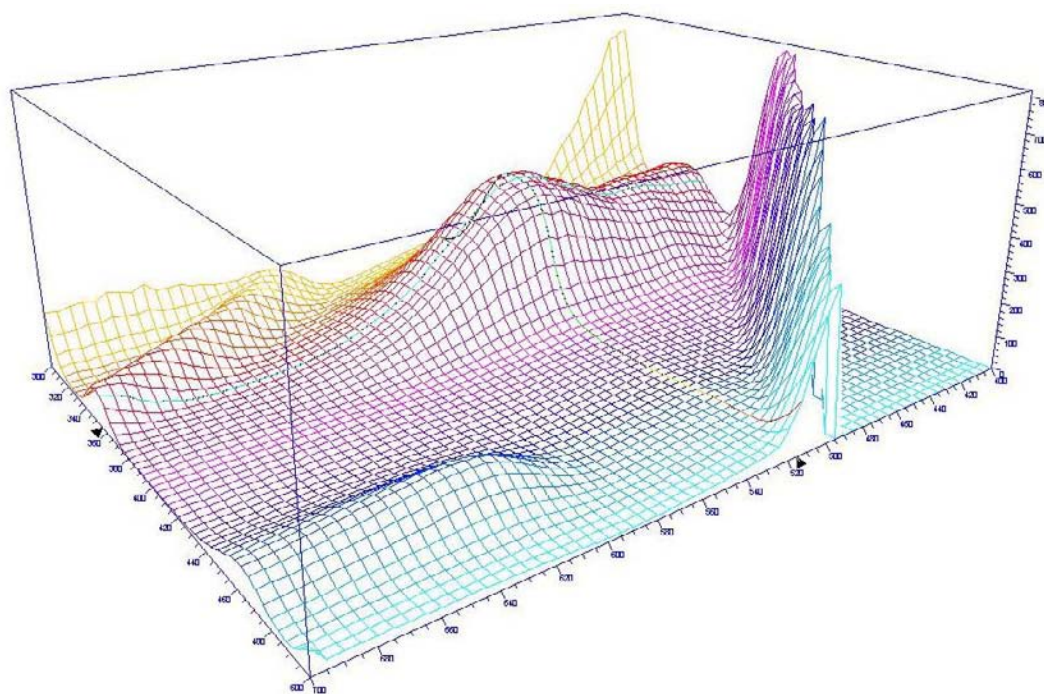
Experiment No.	No. Animals in the Experiment	Dominant Species	Krill Eye Lipofuscin Analysis	Krill Eye Protein Analysis	Whole Samples Frozen	Total Whole Frozen	Storage date
Growth Experiment #11	45	<i>T. inermis</i>	7/9/08	7/10/2008	No	0	7/9/08
Growth Experiment #12	50	<i>T. raschii</i>	N/A	N/A	Yes	50	7/12/08
Growth Experiment #13	50	<i>T. raschii</i>	N/A	N/A	Yes	50	7/13/08
Growth Experiment #14	50	<i>T. raschii</i>	N/A	N/A	Yes	50	7/16/08
Growth Experiment #15	49	<i>T. inermis</i>	N/A	N/A	Yes	49	7/21/08
Growth Experiment #16	50	<i>T. inermis</i>	N/A	N/A	Yes	50	7/24/08
Growth Experiment #17	50	<i>T. raschii</i>	N/A	N/A	Yes	50	7/25/08
Growth Experiment #18	48	<i>T. inermis</i>	N/A	N/A	Yes	48	7/28/08

Table 4: Sediment and particle sample collection for intact BHP analysis

Station Name	Bottom Depth (m)	Depth of sediment (cm)	Vol. water filtered (L)
UAP-3	88	0 - 2	0
CN-10	84	0 - 2	24.2
CN-18	1665	0 - 17	34.2
MN-5	62	0 - 10	22.0
SL-8	58	0 - 2	23.0
NP-7	66	0 - 2	24.0
NP-15	2755	0 - 2	24.0
NP-14	140	0 - 2	20.0
LS16	188	0 - 2	24.0
P14-2	126	0 - 2	24.0
MN-5 (revisit)	62	0 - 2	0
MN-15	145	0 - 10	19.5
SL-14	80	0 - 2	23.0

*All sediment and filters frozen in -80°C immediately following collection and filtration, respectively

Figure 1: Three-dimensional scan of lipofuscin in *Thysanoessa inermis* neural tissues from HLY0802.



9.10 Ichthyoplankton – UAF group

The effect of environmental factors on distribution, abundance, and energy density of late larval and early juvenile stages of walleye pollock (*Theragra chalcogramma*), Pacific cod (*Gadus macrocephalus*), and arrowtooth flounder (*Atheresthes stomias*)

Participating Scientists

Nicola Hillgruber, Ron Heintz, Elizabeth Siddon

Statement of objectives

Our main objectives were to (1) characterize the larval/early juvenile summer fish assemblages in the eastern Bering Sea, (2) determine the effect of environmental factors on the abundance, distribution, and composition of ichthyoplankton, and (3) examine the energy density of larval and early juvenile stages of three target species, namely walleye pollock (*Theragra chalcogramma*), Pacific cod (*Gadus macrocephalus*), and arrowtooth flounder (*Atheresthes stomias*). Our participation on this research cruise was supported by the North Pacific Research Board's Bering Sea Integrated Ecosystem Research Program (BSIERP) and the National Science Foundation's Bering Ecosystem Studies (BEST). Our data will contribute to the collaborative effort of improving the understanding of the effects of climate variability on spatial and temporal variability in abundance, distribution, and condition of the early life stages of these three target fish species in the eastern Bering Sea. Ultimately, these data will allow for the better prediction of transport pathways of early life stages and of production of the three target taxa under changing climatic conditions in the eastern Bering Sea.

Field Methods

Sampling was conducted in collaboration with Alexei Pinchuk. We sampled ichthyoplankton using a 1 m² MOCNESS (Multiple Opening Closing Net and Environmental Sensing System) equipped with nine 500 µm mesh nets. In addition, the MOCNESS was equipped with sensors for conductivity, temperature, and fluorescence, thus recording data to characterize the sampled water column. Sampling was conducted obliquely in 20-m increments from 100 m depth (or 15 m above the bottom) to the surface. After retrieval of the gear, all net bags were carefully rinsed down, codends were detached, and samples were preserved in 10% formalin seawater. Samples will be brought to the lab for further processing. In addition, the contents of the drogue net of the MOCNESS were examined on board and all larval and early juvenile fishes were removed for identification. Early life stages of target taxa were counted and frozen at -80 °C for further analysis of energy density in the lab.

During the first leg of HLY0803, a total of 45 stations on 7 transects were sampled (Table 1). While not all scheduled stations could be sampled due to time constraints, the realized sampling effort still provides a large spatial coverage of data on ichthyoplankton assemblages from the eastern Bering Sea. In addition, MOCNESS sampling is continuing on leg 2 of HLY0803 and stations collected as part of that effort will further increase the spatial extent of the ichthyoplankton data.

The examination of the drogue net of the MOCNESS resulted in the collection of early life stages of all three target taxa. Overall, a total of 440 *T. chalcogramma*, 24 *G. macrocephalus*, and 9 *Atheresthes* spp. were frozen for future analysis of energy density. Highest abundances of *T. chalcogramma* were observed in the vicinity of St. Paul Island; at all other stations, numbers of *T. chalcogramma* were substantially lower. Future lab analysis will provide data on the horizontal and vertical distribution, size, and condition of early life stages of the target fish taxa.

MOCNESS sampling benefitted greatly from the generous help provided by the participating PolarTrec teacher-at-sea, Ms. Jillian Worssam.

Table 1. Summary of MOCNESS samples collected during leg 1 of HLY0803 and number of specimens frozen for further analysis

Station ID	MOCNESS #	# Samples	<i>Theragra chalcogramma</i>	<i>Gadus macrocephalus</i>	<i>Atheresthes stomias</i>
UP 1	MOC 1	4	0	0	0
UP 2	MOC 2	5	0	0	0
UP 4	MOC 3	4	0	3	0
UP 3	MOC 4	5	2	9	0
UAP 8	MOC 5	5	0	0	0
UAP 7	MOC 6	5	0	0	0
UAP 6	MOC 7	4	0	0	1
UAP 5	MOC 8	4	0	0	0
UAP 4	MOC 9	4	0	3	0
UAP 3	MOC 10	4	0	1	0
CN 6	MOC 11	3	0	2	0
CN 8	MOC 12	3	0	0	0
CN 14	MOC 13	5	0	0	3
CN 18	MOC 14	5	0	0	0
CN 16	MOC 15	5	0	0	1
X 1	MOC 16	5	0	0	2
X 2	MOC 17	5	0	0	0
CNN 8	MOC 18	5	0	0	1
CNN 7	MOC 19	5	0	2	0
CNN 6	MOC 20	4	0	0	0
MN 1	MOC 21	1	0	0	0
MN 2	MOC 22	2	0	0	0
MN 3	MOC 23	2	0	0	0
MN 4	MOC 24	2	0	0	0
MN 5	MOC 25	3	0	0	0
MN 6	MOC 26	3	0	0	0
MN 7	MOC 27	3	0	0	0
MN 8	MOC 28	3	0	1	0
SL 7B	MOC 29	2	0	1	0
SL 5B	MOC 30	2	0	1	0
SL 3B	MOC 31	2	0	0	0
NP 1	MOC 32	1	3	0	0
NP 3	MOC 33	2	2	0	0
NP 5	MOC 34	3	0	1	1
NP 7	MOC 35	3	34	0	0
NP 9	MOC 36	3	34	0	0
NP 10	MOC 37	2	66	0	0
NP 11	MOC 38	3	155	0	0
NP 12	MOC 39	4	47	0	0
NP 13	MOC 40	5	1	0	0
NP 14	MOC 41	5	0	0	0
NP 15	MOC 42	5	0	0	0
NP 12.5	MOC 43	5	0	0	0
NP 11.5	MOC 44	4	78	0	0
NP 10.5	MOC 45	2	18	0	0

9.11 Microzooplankton grazing – UMD group

Project: Microzooplankton abundance, biomass and grazing impact on phytoplankton during summer in the eastern Bering Sea (funded by BSIERP)

PI: Diane Stoecker; Research Assistant: Kristen Blattner

Science Goals for Cruise:

1. Obtain samples for determination of microzooplankton (MZ) abundance, biomass and composition in the mixed layer and chlorophyll maximum layer.
2. Estimate the grazing impact of microzooplankton on phytoplankton in the mixed layer.

Methods:

Water samples were collected on “Prod” and some regular CTD casts from the mixed layer and chlorophyll maximum layer, if present, and preserved with 10% acid Lugol’s solution. These samples will be shipped to HPL for enumeration of the larger microzooplankton (>20 micron microzooplankton). In addition, 20 ml samples were preserved with alternative fixatives including Basic Lugol’s solution, acid Lugol’s solution and 1 % glutaraldehyde. These smaller samples were observed onboard to characterize the phytoplankton and microzooplankton communities used in the dilution experiments and to make note of the presence or absence of plastids and fluorescence of the larger protists. This information will be useful in identifying and characterizing microzooplankton when the samples are counted in the laboratory. It has also been useful in understanding optical signals, particularly orange fluorescence, detected by in situ sensors (Dr. Goes and colleagues). Water samples for microzooplankton enumeration were collected in association with the dilution experiments and at additional selected stations (refer to tables).

Two point dilution grazing experiments (with and without the addition of N) were conducted to estimate grazing rates of microzooplankton assemblages on phytoplankton. Water for the experiments was collected from the mixed layer. Grazing on total chlorophyll and chlorophyll in the >20 micron size was determined from 24 h incubations conducted in a flowing seawater on deck incubator screened to achieve an irradiance of ~50% I₀. Experiments were conducted at 21 stations (refer to table). In conjunction with these experiments, Dr. Goes has been measuring variable fluorescence (an indicator of phytoplankton health and potential productivity) in control and N enriched treatments. Treatments from the experiments are being analyzed by Dr. Sambrotto to investigate the role of microzooplankton grazing in urea production. These collaborative efforts should provide some data on nutrient regeneration by microzooplankton and phytoplankton physiology in the Bering Sea during summer.

Expected Results:

We will have data on the abundance, biomass, and taxonomic composition of microzooplankton (primarily ciliates and dinoflagellates) from different regimes. We observed and sampled a diversity of phytoplankton and microzooplankton communities, some containing unusual organisms. In low nutrient waters, the phytoplankton was dominated by very small cells and microzooplankton (primarily

ciliates), not phytoplankton, were the dominant microplankton (20-200 micron fraction). At many stations, the phytoplankton were N limited and their growth was stimulated by the addition of nitrate: at other stations, although the phytoplankton appeared to be nutrient limited, they did not grow in response to added nitrate in the incubations. Nutrient regeneration by microzooplankton appeared to be important to phytoplankton growth in many experiments. Cryptophytes, single cell *Phaeocystis* and other small flagellates were usually the dominant phytoplankton in low chlorophyll waters. These small cells made up the chlorophyll maximum at some stations at the edge of the shelf.

Twenty-one dilution grazing experiments were conducted. Microzooplankton grazing was a large source of mortality to phytoplankton at most stations (see example of data from experiment 18). Photosynthetic mixotrophic ciliates appeared to be the dominant large micrograzers in low chlorophyll waters. Large non- thecate dinoflagelates were important grazers in chlorophyll maximum layers dominated by diatom chains. They were a source of fecal particles containing diatom frustules. At the stations centered around SL12, we found a interesting diverse assemblage containing some unexpected organisms, including large cyanobacteria chains (see micrographs below).

Abundance and biomass data, expected availability: June 2009. Estimates of microzooplankton grazing on phytoplankton, expected availability: October-November 2008.

List of Microzooplankton Grazing Experiments Conducted, Summer 2008

Experiment#	Date	Station	Sample Depth (m) *
1	4-July	UP-3	5
2	5-July	CN-6	7
3	6-July	Pit 1	6
4	7-July	Sta 21	5
5	9-July	MN-3	3
6	10-July	C-55	4
7	11-July	W-4b	4
8	12-July	SL8	4
9	14-July	NP-7	7
10	15-July	NP-14	4
11	18-July	NP-14	5
12	19-July	NP-11	3
13	20-July	LS1-6	5
14	21-July	P14.4.5	7
15	22-July	P14.2	7
16	23-July	MN-6	5
17	24-July	MN-12	7
18	25-July	MN-19	9

19	26-July	SL-14	6
20	27-July	70m-53	7
21	28-July	70m-36	5
*Correponds to 55% Io, in mixed layer.			

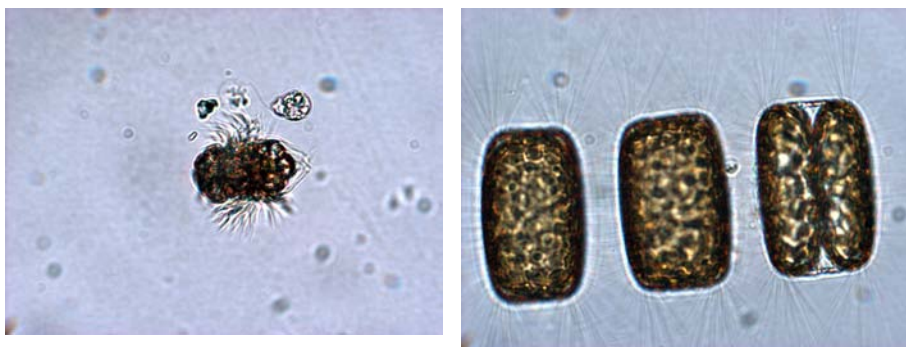
List of Microzooplankton Samples Collected, Summer 2008

Date	Station	Mixed layer sample depth (m)	Chl. Max Sample Depth (m)
4-July	UP-3	5	10
5-July	CN-6	7	29
6-July	Pit 1	6	19
7-July	Sta 21	5	NA
9-July	MN-3	3	NA
	MN-6	10	39
10-July	C-55	4	NA
11-July	W-4b	4	NA
	N-55	20	40
12-July	SL8	4	34
13-July	NP-1	5	NA
	NP-2	5	NA
	NP-3	5	NA
	NP-4	5	28
14-July	NP-5	5	20
	NP-6	5	NA
	NP-7	7	NA
	NP-8	5	NA
	NP-9	5	30
	NP-10	5	NA
	NP-11	5	NA
	NP-12	5	NA
	NP-13	5	20
15-July	NP-14	4	NA
	NP-15	5	NA
18-July	NP-14	5	15
19-July	NP-11	3	17
20-July	LS1-6	5	NA
21-July	P14.4.5	7	NA
22-July	P14.2	7	27
23-July	MN-6	5	NA
	MN-9	10	29
24-July	MN-12	7	33
25-July	MN-19	9	34, 39, 43

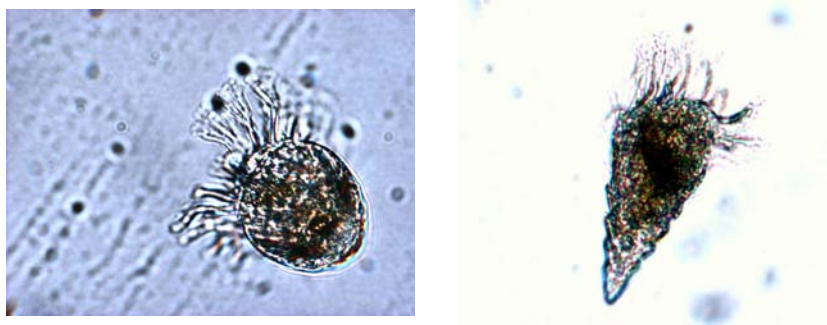
26-July	SL-14	6, 10	30, 30
	SL13	NA	30
	SL-12	NA	30
27-July	70m-53	7	32
	70m-50	10	20, 30,40,50
	70m-47	NA	20,30
28-July	70m-36	5	25
	70m-27	10	30
29-July	70m-17	10	25
	70m-13	6	14
NA=Not Applicable, no strong Chl. Max or sample not available			

Sample results (photomicrographs taken by DS and John Casey):

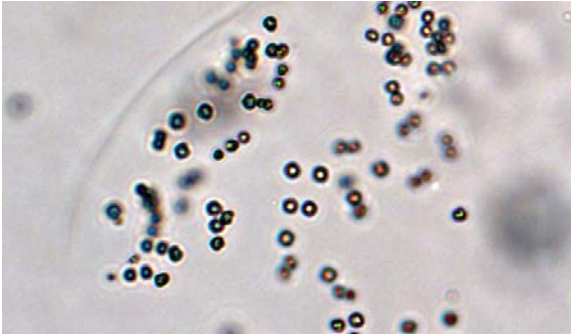
In the chlorophyll maximum layer on the mid-shelf, often the ciliate *Myrionecta rubra* and small phytoflagellates, particularly cryptophytes, were common along with large chain forming diatoms (example, station N-55, midshelf south of St Matthews Island). This mixture of diatoms, phytoflagellates and microzooplankton may be important as a concentrated source of food for macrozooplankton.



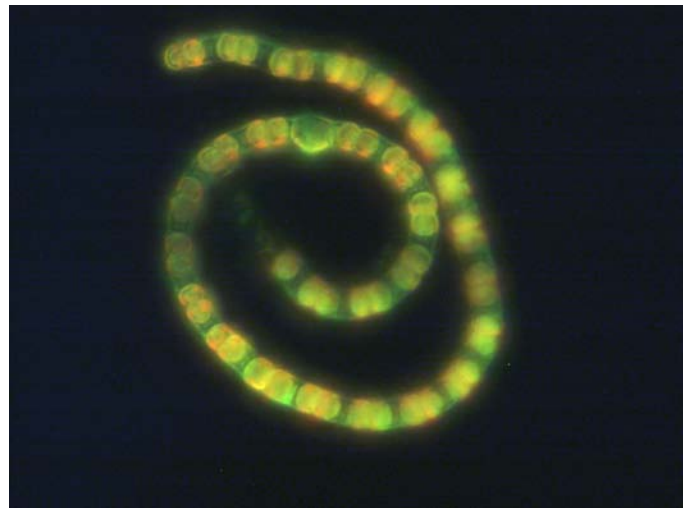
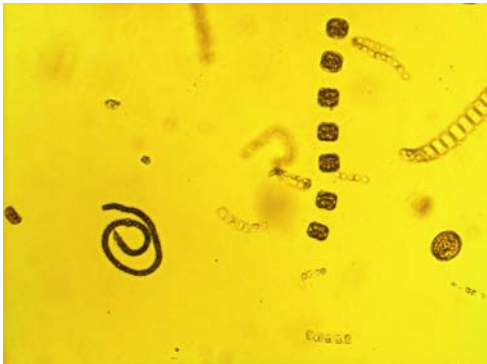
At many shelf stations, chlorophyll values were low and most of the phytoplankton were small (>90% of chl in < 20 micron size class). At these stations, ciliates were very abundant and large photosynthetic ciliates were very common in the mixed layer. Ciliates may be important prey for macrozooplankton and as well as important as photosynthetic microplankton in low chlorophyll waters. For example, *Laboea strobila* (a mixotrophic ciliate) and heterotrophic ciliates were common in the mixed layer at station NP-2 located on the mid shelf.



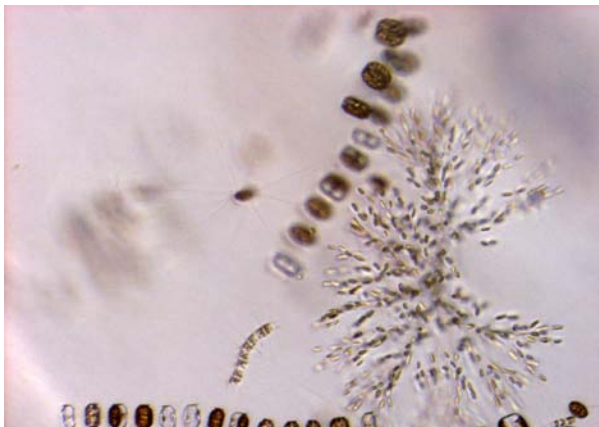
At some stations, the mixed layer was dominated by *Phaeocystis* and small diatoms. At these stations a rich mixture of ciliate and dinoflagellate grazers was observed (examples stations NP-14 & 15 near the shelf edge).



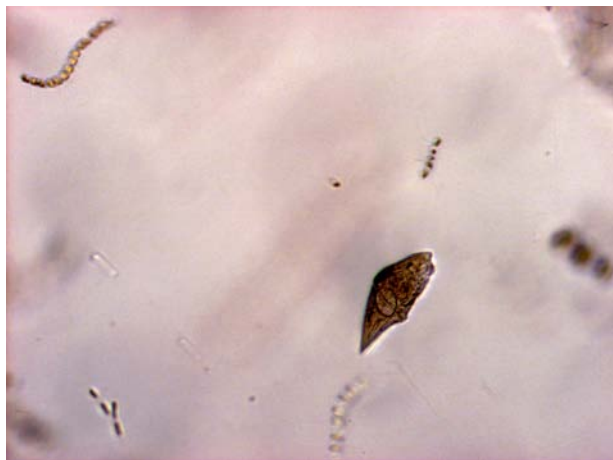
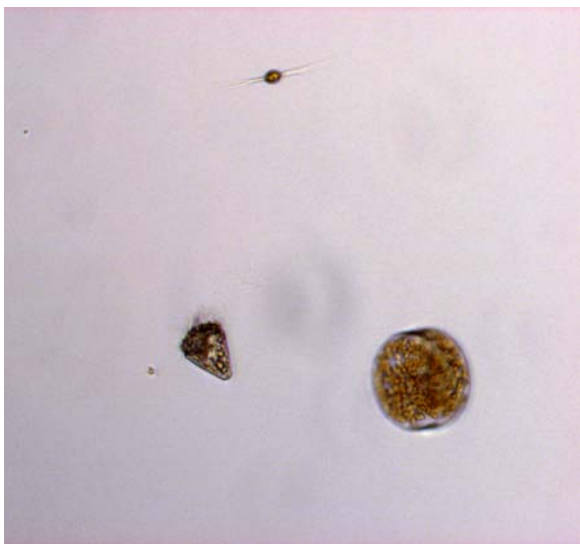
At midshelf stations SL-12 and SL-13, located in the cold pool area southwest of St. Matthews Island, cyanobacterial chains with heterocysts (tentatively identified as *Anabaena* sp.) were present in the chl. max. layer. The chains appeared to be healthy. This observation is very surprising. Do they survive year round in Bering Sea waters or are they seeded by river discharge? Are they fixing N?



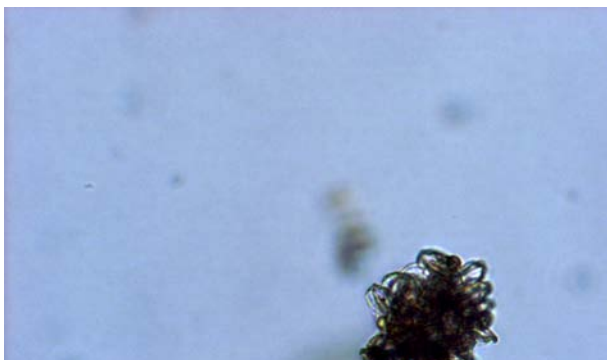
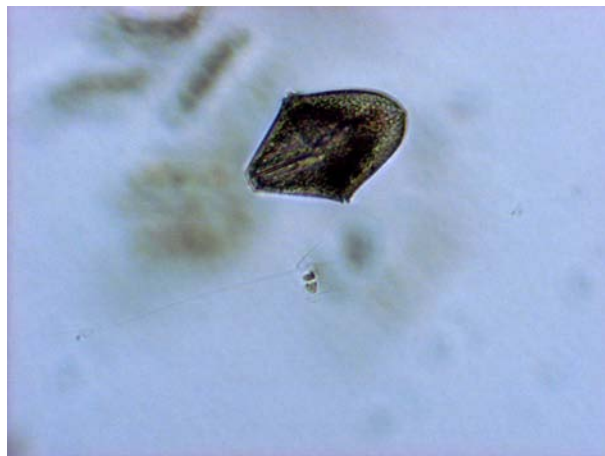
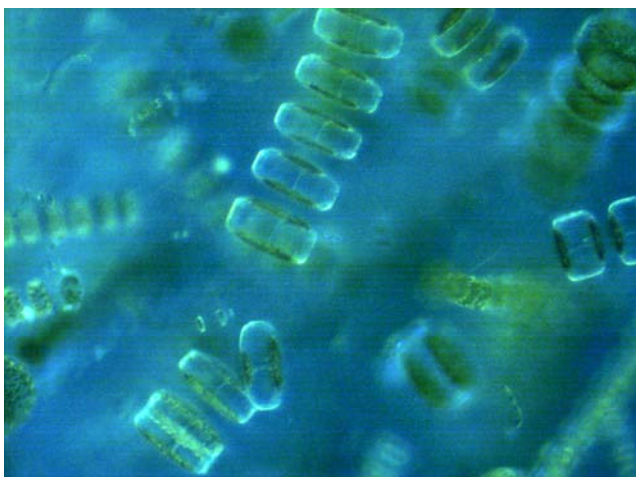
The phytoplankton at SL-12 and S-13 was rich mixture of diatoms and flagellates. Colonies of the mixotrophic chrysophyte, *Dinobryon*, were common.



Microzooplankton were abundant and included photosynthetic and non-photosynthetic ciliates and small and large heterotrophic dinoflagellates. This was a rich and diverse assemblage of microplankton.



At some mid shelf stations, a chlorophyll max dominated by large diatom chains was present. Large non-thecate heterotrophic dinoflagellates were grazing on the diatoms and producing fecal pellets. An example is mid shelf station 70m-47.



Example of data from a dilution experiment, these results are preliminary. The data have not undergone statistical analyses.

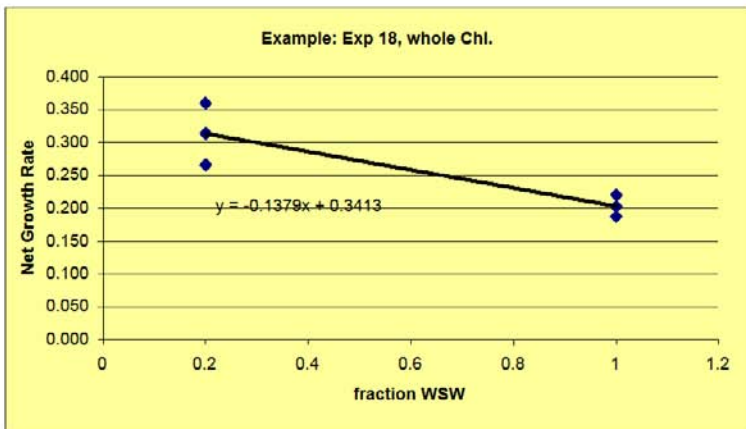
Experiment #18 25-Jul-08 Sta110 MN-19 9m Preliminary data

Pt=final chl
Po= initial chl
K=net growth rate
 μ = intrinsic growth rate
g=grazing rate

Treatment	Pt (t=24)	Po (t=0)	Whole Chl's			μ	g	g/u
WSW	0.463	0.371	0.220	1		0.341	-0.138	0.40
	0.448	0.371	0.187	1				
	0.455	0.371	0.202	1				
20% WSW	0.107	0.075	0.360	0.2				
	0.102	0.075	0.314	0.2				
	0.098	0.075	0.266	0.2				
WSW + N	0.449	0.371	0.190	1		0.243	-0.063	0.26
	0.453	0.371	0.199	1				
	0.431	0.371	0.150	1				
20% WSW + N	0.091	0.075	0.201	0.2				
	0.095	0.075	0.238	0.2				
	0.096	0.075	0.252	0.2				

Control
Plus N

Treatment	Pt (t=24)	Po (t=0)	<20 μ m chl's			μ	g	
WSW	0.429	0.359	0.178	1		0.316	-0.127	0.40
	0.440	0.359	0.204	1				
	0.431	0.359	0.185	1				
20% WSW	0.098	0.074	0.281	0.2				
	0.101	0.074	0.316	0.2				
	0.097	0.074	0.274	0.2				
WSW + N	0.474	0.359	0.279	1		0.288	-0.012	0.04
	0.466	0.359	0.261	1				
	0.478	0.359	0.288	1				
20% WSW + N	0.100	0.074	0.302	0.2				
	0.098	0.074	0.288	0.2				
	0.096	0.074	0.267	0.2				



The results of this experiment indicate several things. At this station, most of the chlorophyll is < 20 micron phytoplankton. The growth rate of the phytoplankton (μ), was not stimulated the addition of nitrate; the phytoplankton do not appear to be N limited. In the treatments without added nutrients, the grazing coefficient was 0.34 d⁻¹

and about 40% of the daily phytoplankton production was grazed by microzooplankton. For some unknown reason, the addition of nitrate appeared to inhibit grazing. It will be interesting to see if the variable fluorescence is consistent with the phytoplankton responses observed. It will also be interesting to determine if the phytoplankton growth rates correlate with measurements of primary productivity conducted simultaneously on the cruise and whether the microzooplankton grazing rates correlate with urea production.

The results from this grazing experiment will be compared to those from the other 20 stations to determine if grazing coefficients or % of production or standing stock grazed per day are related to phytoplankton or microzooplankton community composition, nutrient availability or domain.

9.12 Benthic biological characterization and fluxes – WWU group

Shull/Devol Cruise report

The primary goal of the benthic biogeochemistry group is to measure rates of sedimentary organic carbon remineralization, benthic denitrification rates, nutrient fluxes, and sediment bioirrigation rates and to evaluate the role of the benthos in the nitrogen cycle of the Bering Sea. A secondary goal is to determine gas exchange rates for calculating water column primary productivity along with measurements of the triple isotopes of dissolved oxygen.

Core samples

Fifteen stations have been sampled during this cruise using an Ocean Instruments MC-800 multicorer equipped with eight 10-cm diameter polycarbonate core tubes. Two drops were made at each station resulting in as many as sixteen cores per station. The actual number of usable samples generally averaged approximately twelve. Cores were processed on deck and, depending upon the number of usable cores recovered, were allocated as follows:

- 2 - 3 flux cores (incubated for ca. 5d and overlying water sampled for, N₂/Ar, O₂/Ar, nitrate, nitrite, ammonium, phosphate, and silicate). Following flux measurements, these were frozen for later CT-scanning of burrow distributions
- 1 squeeze core
 - Profiles of dissolved oxygen measured by microelectrode and by optode
 - Profiles of dissolved nutrients (nitrate, nitrite, ammonia) by whole-core squeezing
- 2 section cores cut at 0.5-cm or 1-cm intervals and centrifuged for pore-water nutrients, nitrate, nitrite, ammonium, phosphate, silicate, dissolved iron and manganese to 20 cm. Remaining sediment was reserved and frozen for later measurements of solid-phase elements (Fe, Mn, Al, C, N, Pb-210)
- 2-3 cores sectioned at 2-cm intervals for measurement of Rn-222/Ra-226 disequilibrium
- 2-3 cores sieved over 0.5-mm sieve and preserved in 10% formalin for later enumeration of benthic infauna

Water-column sampling

At nine stations vertical profiles of $^{222}\text{Rn}/^{226}\text{Ra}$ were measured. The $^{222}\text{Rn}/^{226}\text{Ra}$ measurements will be used for determining gas exchange rates and, combined with oxygen isotope data, rates of net primary production.

Multicore locations

Station data						Measurements			
Stn	Name	Date	Latitude	Longitude	Depth (m)	[O ₂]pw	Flux	[Nut]pw	^{222}Rn Benthos
4	UP-3	7/4/2008	54° 25.3' N	165° 8.4' W	133	X	X	X	X
9	UAP-4	7/5/2008	55° 45.7' N	169° 31.6' W	94	X	X	X	X
17	CN-18	7/6/2008	55° 18.4' N	168° 23.5' W	1615	X	X	X	X
24	CNN6	7/8/2008	56° 45.68' N	167° 51.77' W	100		X	X	X
		X							
31	MN-5	7/9/2008	59° 54' N	170° 23.8' W	63	X	X	X	X
46	SL-8	7/12/2008	62° 12.1' N	172° 42.83' W			59	X	X
		X	X						
59	NP-7	7/14/2008	57° 53.6' N	169° 13.8' W	66		X	X	X
		X							
68	NP-15	7/15/2008	56° 3.5' N	171° 18.6' W	2766	X	X	X	X
79	NP-14	7/18/2008	56° 17.21' N	171° 2.85' W	136		X	X	X
		X							
86	LS1-6	7/20/2008	56° 50.0' N	173° 18.9' W	160			X	
92	P-14-2	7/22/2008	59° 00.6' N	174° 00.52' W		W	123	X	X
		X	X						
99	MN-5	7/23/2008	59° 54.1' N	170° 23.6' W	64		X	X	X
		X							
102	MN-11	7/24/2008	59° 54.0' N	173° 59.9' W	101		X	X	X
		X							
106	MN-15	7/25/2008	59° 54.3' N	176° 23.3' W	139		X	X	X
		X							
116	SL-14	7/26/2008	62° 11.5' N	175° 10.7' W	80		X	X	X
		X							

Expected results

Our measurements of oxygen and nitrogen changes in flux cores will enable us to determine the rate of organic matter supply to the benthos and the rate of denitrification. Nutrient measurements in flux cores will enable us to calculate rates of nutrient cycling between sediments and overlying water. We will calculate rates of sediment bioirrigation from the pore water profiles of Rn-222. Profiles of dissolved oxygen and nutrients will enable us to infer the biogeochemical processes leading to the observed nutrient and gas fluxes. Samples preserved for identification and enumeration of benthic organisms along with CT scans of flux core burrow distributions will enable us to examine how benthic organisms influence nutrient cycling in Bering Shelf sediments.

Many of these measurements will not be completed until after the Healy returns to port in October. At that time, we will be able to offload samples and equipment necessary to complete this work. I expect that most data will be available by July 2009.

Example of data from flux core experiments and sediment profiles, Stn 24

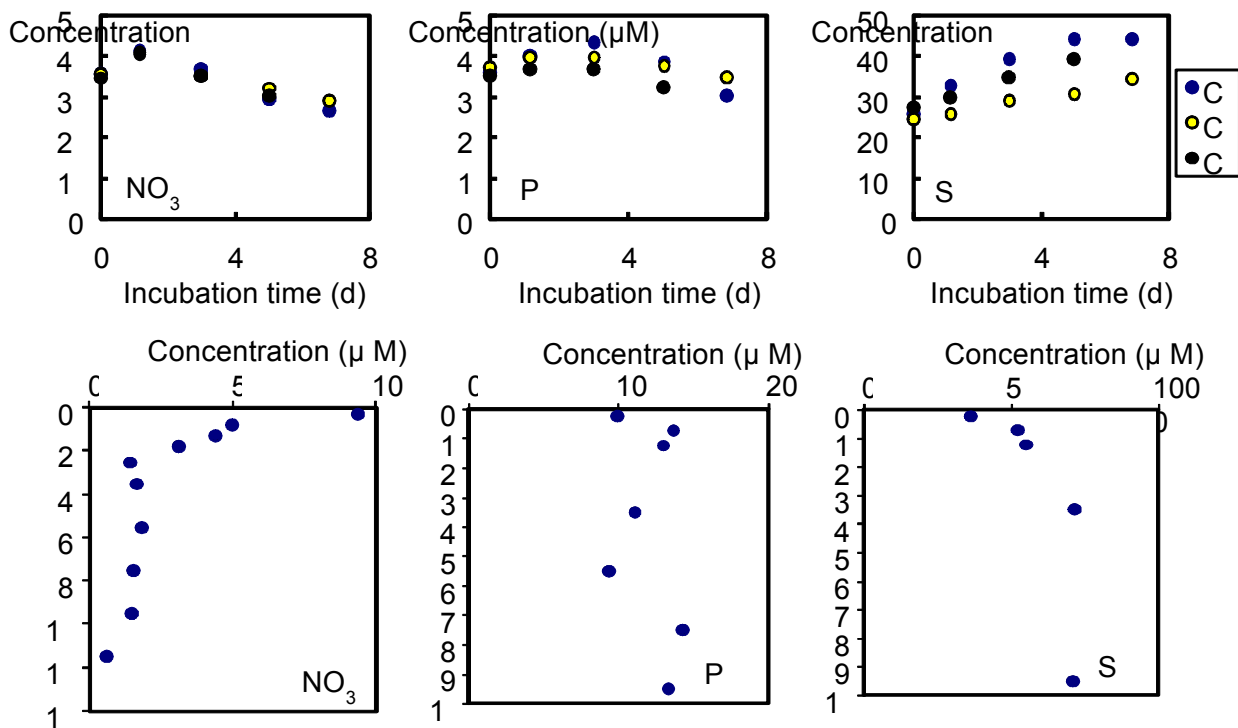


Figure 9-12.1. Changes in concentration of nitrate, phosphate and silicate in flux-core experiments (top row) and corresponding profiles in sediment from the same station (bottom row). Station 24, 100 m water depth.

9.13 Bird distribution and abundance – US Fish & Wildlife group

Seabird and Marine Mammal Surveys aboard HLY0803

Kathy J. Kuletz and Elizabeth A. Labunski

U.S. Fish and Wildlife Service, Migratory Bird Management, MS-201, 1011 E. Tudor Rd., Anchorage, Alaska 99507

Project:

As part of the HLY0803 cruise we surveyed marine birds and mammals in conjunction with the National Science Foundation 2008 BEST cruise onboard the USCGC Healy July 3 - 31, 2008. The seabird and marine mammal data will be archived in the North Pacific Pelagic Seabird Database (U.S. Fish and Wildlife Service, Anchorage, Alaska) and are part of the Bering Sea Ecosystem Integrated Research Program funded by the North Pacific Research Board (Anchorage, Alaska).

Methods:

We surveyed marine birds and mammals from the port side of the bridge (22m above the sea surface), using standard survey protocol during daylight hours while the vessel was underway at cruising speeds over 5 knots. One observer scanned the water ahead of the ship using hand-held 10x binoculars and recorded all birds

and mammals within a 300-m arc, extending 90° from the bow to the beam. Birds on the water or foraging were counted continuously. Flying birds were counted during 'snapshots' at intervals that varied with ship speed, typically about once every minute. We recorded the animal's behavior as flying, on water, on scan, or actively foraging. We used strip transect methodology with three distance bins extending from the vessel: 0-100 m, 101- 200 m, 201-300 m. We determined the distance to bird sightings using geometric and laser hand-held rangefinders. Unusual sightings beyond the 300 m strip transect were also recorded for rare birds, for large bird flocks, and mammals.

We used the DLOG2 data entry program (Ford Ecological Consultants, Inc.) to record observations directly into a laptop computer interfaced with the Healy's global positioning system. Every entry by the observer was recorded with location, date, and time stamps, along with associated environmental and observer variables. Location data were also automatically written to the data file in 20 second intervals, and allowed us to simultaneously record changing weather conditions, Beaufort Sea State, ice type and coverage, and glare conditions. We recorded other environmental variables at the beginning of each transect, including wind speed and direction, air temperature, and sea surface temperature.

Preliminary Results and Discussion:

From July 3 - 31, we surveyed 115 transects totaling 3,515 km. During this time we recorded a total of 24,184 birds, of which 95 % were identified to species for a total of 29 species (Table 1). The majority of birds observed were Short-tailed Shearwaters (*Puffinus tenuirostris* and Northern Fulmars (*Fulmarus glacialis*) which were 77 % of the on-transect bird observations. Six species of seabirds comprised 92 % of the identified birds; these were Short-tailed Shearwaters, Northern Fulmar, Fork-tailed Storm-petrel, Thick-billed Murre, Common Murre, and Black-legged Kittiwake (Table 1).

A preliminary examination of the distribution of the two prevalent species of marine birds (short-tailed shearwater and northern fulmars) illustrate that Short-tailed Shearwaters were concentrated near Unimak Pass, while Northern Fulmars were the most ubiquitous of the prevalent seabird species throughout the survey region (Figure 1). Planktivorous Fork-tailed Storm-petrels were the third most prevalent seabird observed during the survey. The majority of these surface feeding Storm-petrels were observed along the Bering Sea shelf edge and southern Bering Sea, while piscivorous Murre spp. (Common & Thick-billed) were more commonly observed on the shelf (Figure 2). Other marine bird observations of interest included 3 species of albatross (Black-footed, Laysan and Short-tailed), Mottled Petrels, and Red-legged Kittiwakes.

Marine Mammals: We recorded 230 marine mammals of 7 identified species (Table 2). Of those, 117 were 'on transect'. Dall's Porpoise were the most common

and widely distributed marine mammal encountered, followed by Northern Fur Seals (Figure 3). We also observed 4 whale species: Fin, Minke, Humpback, and Killer Whales. Humpback whales were observed in the southern Bering Sea north of Unimak Pass, while observations of Fin, Minke and Killer whales extended into the central and north Bering Sea (Figure 3).

Table 1. Marine bird observations during the BEST cruise on the USCGC Healy, July 3 - 31, 2008

Species	Total Count (n)	Proportion of Total (%)	Overall Abundance (# / 100km)
Ancient Murrelet	52	0.22	1.48
Arctic Tern	5	0.02	0.14
Barn Swallow	1	0.00	0.03
Black-footed Albatross	1	0.00	0.03
Black-legged Kittiwake	332	1.37	9.45
Cassin's Auklet	3	0.01	0.09
Common Murre	878	3.63	24.98
Crested Auklet	42	0.17	1.19
Fork-tailed Storm-petrel	1372	5.67	39.03
Glaucous Gull	3	0.01	0.09
Glaucous-winged Gull	1	0.00	0.03
Harlequin Duck	5	0.02	0.14
Herring Gull	5	0.02	0.14
Horned Puffin	97	0.40	2.76
Laysan Albatross	4	0.02	0.11
Least Auklet	111	0.46	3.16
Leach's Storm-petrel	1	0.00	0.03
Mottled Petrel	6	0.02	0.17
Northern Fulmar	5046	20.87	143.56
Parakeet Auklet	65	0.27	1.85
Parasitic Jaeger	2	0.01	0.06
Pigeon Guillemot	3	0.01	0.09
Pomarine Jaeger	19	0.08	0.54
Red Phalarope	25	0.10	0.71
Red-legged Kittiwake	63	0.26	1.79
Short-tailed Albatross	2	0.01	0.06
Short-tailed Shearwater	13599	56.23	386.88
Thick-billed Murre	1131	4.68	32.18
Tufted Puffin	109	0.45	3.10
Unid. Alcid	21	0.09	0.60
Unid. Dark Shearwater	987	4.08	28.08
Unid. Gull	1	0.00	0.03
Unid. Kittiwake	6	0.02	0.17
Unid. Murre	181	0.75	5.15
Unid. Shearwater	1	0.00	0.03
Unid. Small Dark Alcid	4	0.02	0.11
Total	24184	100.00	688.02

Table 2. Marine mammal observations during the BEST cruise on the USCGC Healy, July 3- 31, 2008. Off transect counts were observations > 300 m from the ship's center line.

Species	On transect counts			Off transect (> 300 m)
	Total Count (n)	Proportion of Total (%)	Overall Abundance (# / 100km)	Total Count (n)
Dall's Porpoise	61	52.14	1.74	61
Fin Whale	3	2.56	0.09	14
Humpback Whale	2	1.71	0.06	6
Killer Whale	0	0.00	0.00	12
Minke Whale	2	1.71	0.06	7
Northern Fur Seal	45	38.46	1.28	11
Pacific Walrus	0	0.00	0.00	1
Unid. Seal	1	0.85	0.03	0
Unid. Whale	1	0.85	0.03	1
Unid. Pinniped	2	1.71	0.06	0
Total	117	100	3.33	113

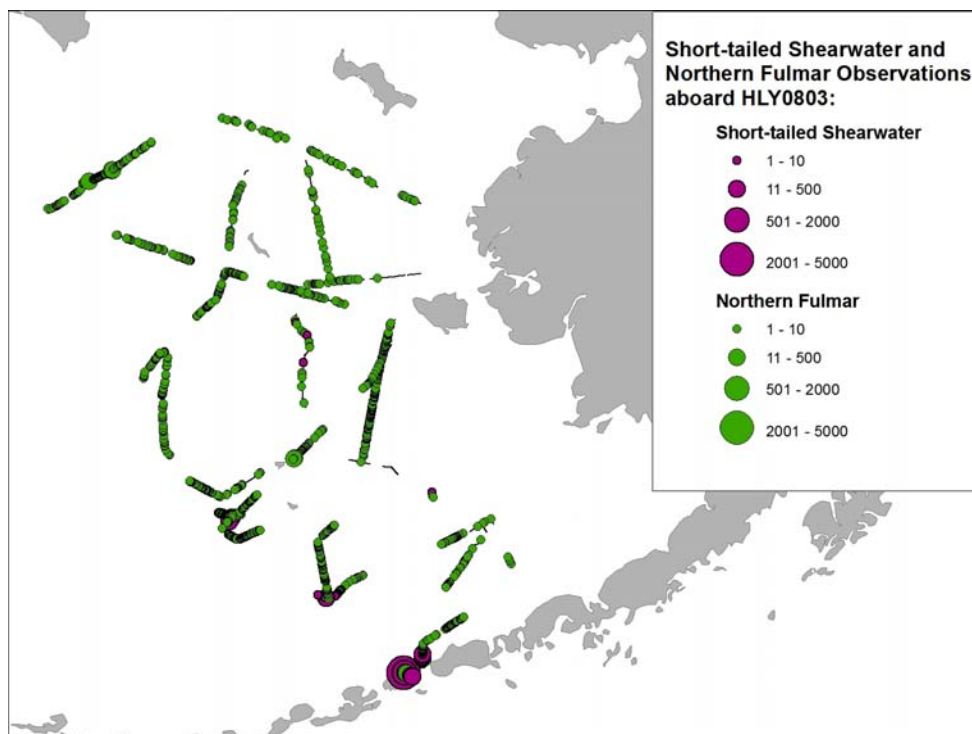


Figure 9-13.1. Short-tailed Shearwater and Northern Fulmar observations during surveys from the Healy, 3 – 31 July, 2008. Black lines indicate survey track lines.

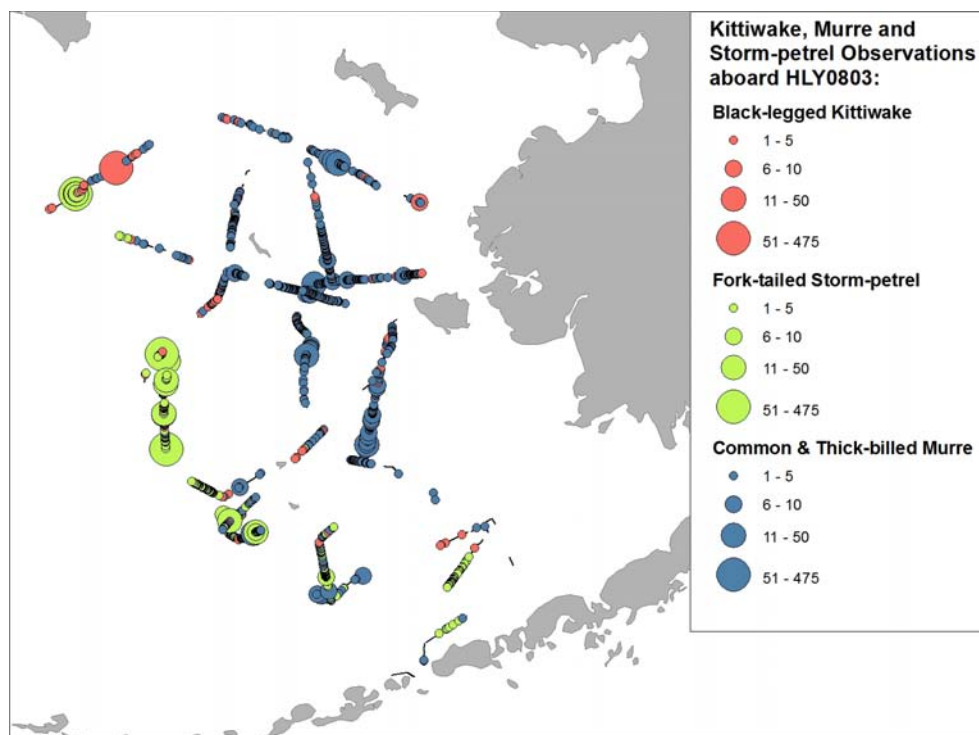


Figure 9-13.2. Black-legged Kittiwake, Fork-tailed Storm-petrel and Common & Thick-billed Murre observations during surveys from the Healy, 3 – 31 July, 2008. Black lines indicate survey track lines.

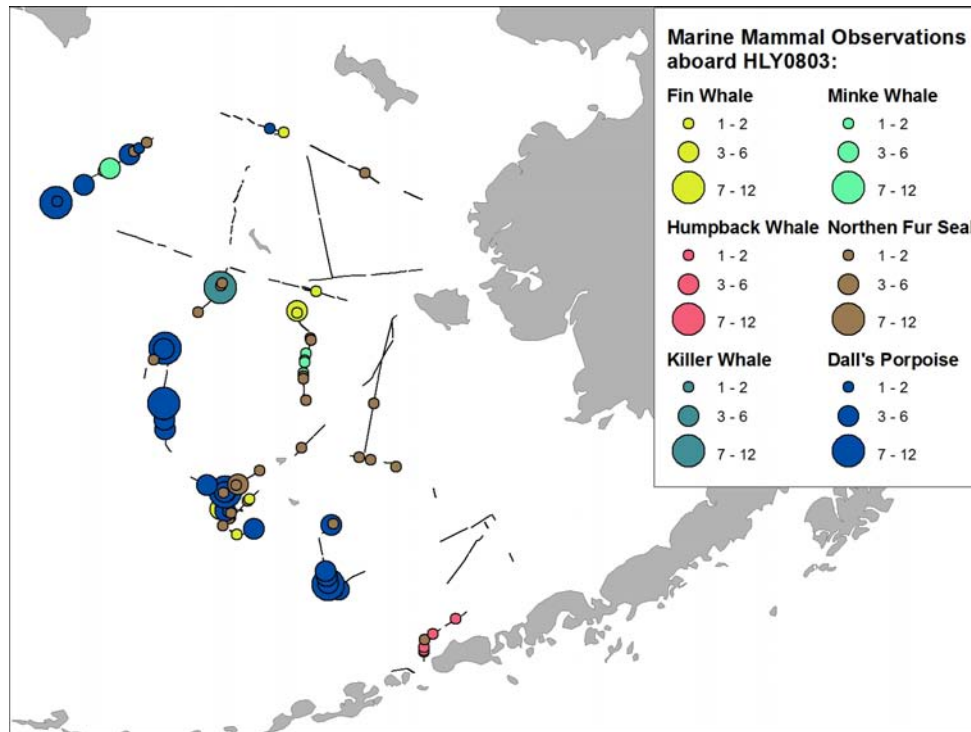


Figure 9-13.3. Marine Mammal observations during surveys from the Healy, 3 – 31 July, 2008. Black lines indicate survey track lines.

9.14 Marine mammal distribution and abundance – NOAA group

Overall goal

This 2008 survey is part of the North Pacific Right Whale Tagging and Foraging Ecology Study and was conducted aboard the USCGC Healy along pre-determined transect lines. Photographic and sighting data were collected on a “not-to-interfere basis” with the oceanographic survey. Right whales were the priority species for this survey but all cetaceans and pinnipeds were of interest and all sightings save fur seals (*Callorhinus ursinus*) near the Pribilof Islands were included in the effort. The primary objective of this portion of the right whale ecology study was to aid in the location of right whales prior to the departure of the R/V “Ocean Olympic” at the beginning of August. This latter cruise will focus on areas of past sightings and carry out a dedicated program of radio tagging, photogrammetry, acoustic monitoring, and biopsy of these animals to further understand their biology and the present state of the health of this remaining stock in the eastern portion of the whales range.

Procedures

The visual data was collected by a researcher positioned in the ship’s Aloft Conning Tower (“aloft con”) with a height of eye of 33 meters using hand held 10 power Zeiss binoculars. Sightings were recorded on WinCruz software during daylight transit periods

between stations. Effort was halted whenever viewing conditions dropped below one nautical mile of visibility generally due to fog or fog and rain. The ship traveled at between 11 and 15 nautical miles per hour and ocean conditions were generally favorable.

Digital photos taken of marine mammals and seabirds were taken by a Canon EOS 30D with a Canon 100-400 image stabilized zoom lens.

All sightings recorded during transit were input into WinCruz which accepts relative angle and distance (reticule or estimated in nautical miles).

Communication between the bird observers working a similar schedule was made possible via three ships VHF radios.

Results and Data Distribution

Daily summaries will accompany the WinCruz data files and be made available to the Chief Scientist of the HLY0803 Cruise as well as members of the NMFS- Northwest Marine Mammal Laboratory in Seattle.

Sample Summary:

July 04- Began survey after morning station and continued throughout day in between stations, working two hours on and ~ one hour off.

16 sightings:

11 of *M. novaeangliae*, (24 individuals)

1 of *P. dalli* porpoise (5 individuals)

1 of *B. physalus* (12 individuals)

2 of unidentified large whales (3 individuals)

Two fur seals were seen by the birders.

Much of the day was spent on a good ocean (Beaufort 2-3) with some fog and rain. The rain became more of a problem later in the day as visibility decreased. After the third station (1900) fog descended and no further effort was possible although just as we arrived at the 2130 station the fog lifted to reveal 8 more fin whales (included in count above).

Sample Win Cruz readout for same day:

```
022*.092405 070408 N54:33.95 W165:10.03
023*.092605 070408 N54:34.36 W165:10.14
024S.092702 070408 N54:34.55 W165:10.19 001 001 2 1 280 8.00 076
025A.092702 070408 N54:34.55 W165:10.19 001 N N 070
1 OBS BEST HIGH LOW %S1 %S2 %S3
026*.092805 070408 N54:34.76 W165:10.25
027C.092941 070408 N54:35.09 W165:10.34 Large whale breaching near horizon
028*.093005 070408 N54:35.17 W165:10.35
029S.093125 070408 N54:35.44 W165:10.40 002 001 6 2 270 2.00 076
030A.093125 070408 N54:35.44 W165:10.40 002 N N 076
1 OBS BEST HIGH LOW %S1 %S2 %S3
031*.093205 070408 N54:35.58 W165:10.42
032C.093236 070408 N54:35.69 W165:10.43 3 humpback, one cow/calf
033*.093405 070408 N54:35.98 W165:10.48
034S.093418 070408 N54:36.02 W165:10.49 003 001 6 2 270 1.40 076
035A.093418 070408 N54:36.02 W165:10.49 003 N N 076
1 OBS BEST HIGH LOW %S1 %S2 %S3
```

036C.093537 070408 N54:36.29 W165:10.54 4 M. novaeangliae traveling slowly in one direction
 037*.093605 070408 N54:36.38 W165:10.55
 038S.093650 070408 N54:36.53 W165:10.59 004 001 3 1 273 0.90 076
 039A.093650 070408 N54:36.53 W165:10.59 004 N N 076
 1 OBS BEST HIGH LOW %S1 %S2 %S3
 040C.093802 070408 N54:36.77 W165:10.64 2 M. novaeangliae slow travel towards stern
 041*.093805 070408 N54:36.79 W165:10.64
 042C.093927 070408 N54:37.07 W165:10.71 Increased bird activity

Photographs:

All pictures of cetaceans will be placed on the “shared site” on the public drive and be available there as well as files that will accompany the data and summaries going to NMML.

Comments on Operations

The USCGC Healy is an outstanding platform for numerous oceanographic observations and experiments. These comments will be restricted to this past survey and future possible marine mammal sighting surveys.

Aloft-Con:

This area of the ship, which is used as a steering station when the ship is transiting ice, is an extremely good area for marine mammal sighting. Protected from the environment and with near 360 degree viewing at the commanding height of 99 feet makes it a natural for line transect or behavioral work. Suggestions to improvement:

- 1) Purchase two pairs of Zeiss 20X48 Image Stabilized binoculars and mount them in the front windows using a sling made of shock cord suspended from the overhead to a small board. This would provide flexibility to move from front to side windows and remove some of the possible vibration encountered at some shaft RPM.
- 2) It would be nice to get the heater working!

Flying Bridge:

This vessel is very stable and the height of the flying bridge would lend itself for “big eye” type line-transect work as well. The ship has two, port and starboard, mounting platforms for pedestals and the 75X150 Fujinon binoculars used on other ships for this type of work. A major draw back would be the environment this ship generally works. Mounting the binoculars and stands would be relatively easy due to the afore mentioned mounts but to make observing from this area feasible some sort of protection from the elements would be required. Room for a structure to enclose the binoculars is restricted by two items on both port and starboard. An antennae/receiver and spotlight surround the present pedestal mounts and would present difficulties to positioning the standard big-eyes and/or a weather housing. Communication would need to be provided with VOX style headsets between port and starboard and a recorder (most likely stationed in the “aloft-con”).

9.15 Water column bio-optics – Bigelow Lab. Group

Cruise Report for HLY0803

Project: Spatial and Temporal Variability in phytoplankton biomass and primary production in the Bering Sea linked to Climate Change (funded by NASA, IPY)

Team Members:

Joaquim I. Goes and Helga Gomes (Bigelow Laboratory for Ocean Sciences)
Eurico D'Sa and Puneeta Naik (Louisiana State University)

Science Goals for the Cruise:

Our major goal in this study was to collect bio-optical data in conjunction with measurements of chlorophyll, rates of primary productivity, CDOM, suspended matter, coccolithophore counts and calcium carbonate in support of NASA's efforts to develop robust empirical and semi-analytic algorithms for ocean color products of the Bering Sea. This effort is part of longer strategic objective of understanding the impacts of changing climate on biological oceanographic processes in the Bering Sea using ocean color satellite data.

An additional goal was to investigate the distribution of major phytoplankton groups and their physiological status in relation to nitrate and iron availability in the water column.

Methods and Approach:

Data required to meet our project goals was acquired from three different streams: 1) Profiling with a bio-optical package 2) Underway measurements with a prototype Automatic Laser Fluorometer and 3) Laboratory based analysis of samples obtained from discrete depths in the water column.

1) Bio-optical Profiling:

A bio-optical package put together prior to this cruise was used to attain some of the core measurements required to meet our project objectives. This bio-optical profiling package comprised of the following instruments: i) **CTD**, ii) **ECO-Triplet** (WETLABS chlorophyll, CDOM fluorometer with scattering sensor), iii) **ac-s** (WETLABS hyperspectral absorption-attenuation meter), iv) **Hyper OCI** (Satlantic hyperspectral downwelling irradiance sensor), v) **Hyper OCR** (Satlantic hyperspectral upwelling radiance sensor), vi) **VSF-3** (WETLABS three-angle, three wavelength volume scattering function meter) and a **Fast**



Figure 1: Bio-optical package before deployment at a station in the Bering Sea during USCG Healy cruise 0803.

Repetition Rate Fluorometer (FRRF, JAPAN, for estimating quantum efficiencies (variable fluorescence) of ambient phytoplankton populations, rates of photosynthesis, and parameters (α and P^B_{max}) of the photosynthesis versus light response curve required for modeling of water column primary production (Fig.1). Preliminary results obtained with the bio-optical package and processed are given in Figs. 2 to 4; This data can be used to model column primary production. Our plan is to test model derived measurements of primary production with those made using standard deck-based incubation methods (Fig. 1).

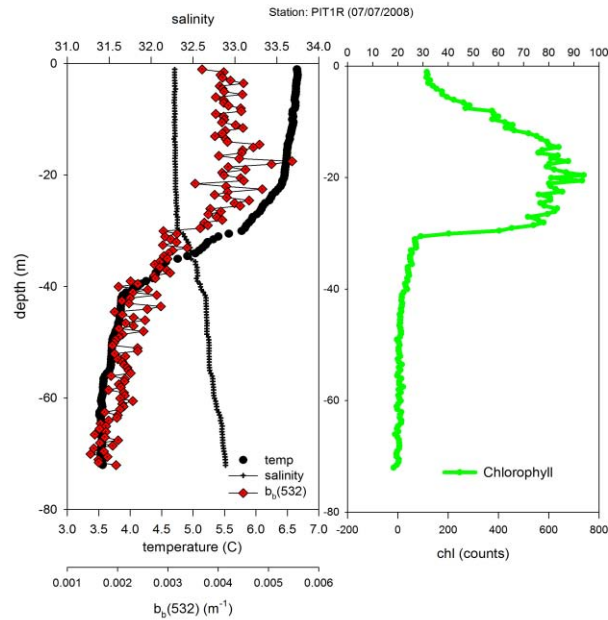


Figure 2. Vertical profiles of water column properties obtained from the optical profiler at station PIT1R

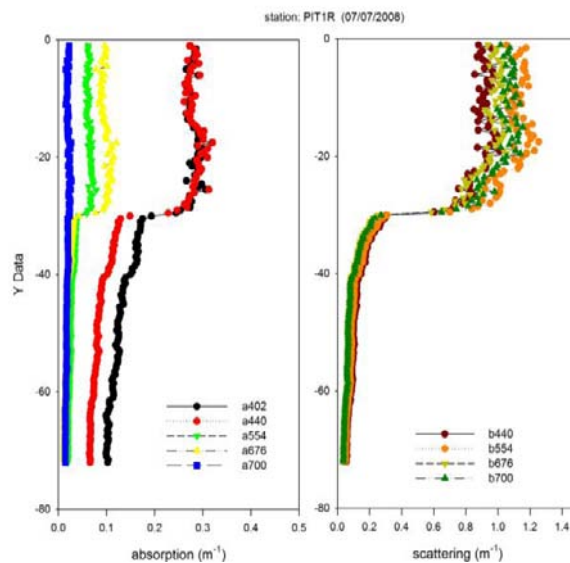


Figure 3. Vertical profiles of spectral absorption and scattering estimated at station PIT1R

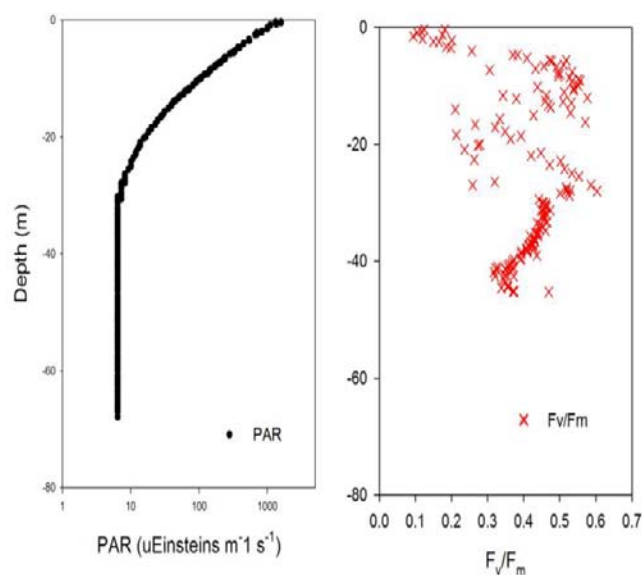


Figure 4. PAR and phytoplankton physiological response F_v/F_m measured by the Fast Repetition Rate Fluorometer (FRRF) on the optical profiler at station PIT1R

On July 8th 2008, during normal shipside operations at station NP2 to the southeast of Nunivak Island the bio-optical package was lost after we had acquired data at 10 stations. By the 19th of July 2008,

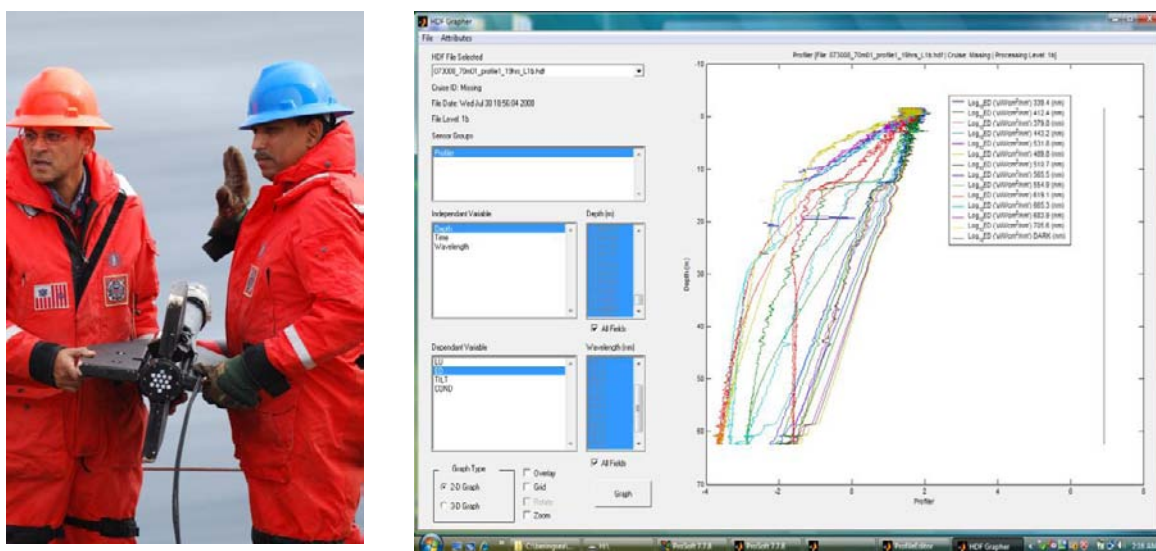


Figure 7. Hand-deployment of Satlantic 14 channel radiometer for measurements of downwelling irradiance and upwelling radiance. Screen capture of the radiometer data is also shown.

with the exception of the profiling FRRF, we were able to obtain replacement instruments to continue our measurements. Following the loss of the bio-optical package, replacement equipment were sent to St. Paul when both new scientific crew and USCG member were picked up in the island. Dr. Ian Walsh from Wetlabs put together and rushed a basic optical package to St. Paul. Dr. Ajit Subramaniam from LDEO shipped the Satlantic Profiling and surface reference multi-channel radiometers (Figure 6). These

equipments were instrumental in our group being able to continue making optical measurements in the Bering Sea from 22 July till the end of the cruise. Over the period of the cruise we carried out around 46 optical deployments.

Operations with the replacement instruments commenced on 20th of July, St. LS1 onwards and continued until the 30th of July 2008. In addition to water column profiles of underwater light, one of the instruments that was part of the replacement was able to provide us with on-deck measurement of incoming solar irradiance at 14 different wavelengths.

2) Underway measurements:

A dual automated blue and green laser induction fluorometer (ALF) being developed at Lamont Doherty

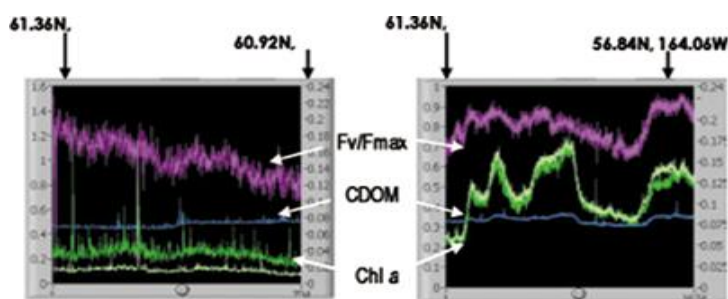


Figure 6. Screen capture of data generated by the ALF in underway mode along the 70m isobath line.

Earth Observatory by Dr. A. Chekalyuk, was serially interfaced with several other instruments fitted to the ship's underway system to obtain measurements of phytoplankton chlorophyll, a measure of major phytoplankton groups (diatoms, dinoflagellates, cryptophytes, cyanobacteria etc. based on specific pigment fluorescence signatures of each group) and

variable fluorescence as the ship transected through different water types in the Bering Sea. The instrument also provided trends in Colored Dissolved Organic Matter (CDOM) along the cruise track when it was operated in underway mode. An illustration of data sets along the 70m-isobath is shown in Figure 6.

At a few select stations, the ALF was operated in discrete mode for quantifying the quantum efficiencies for photosynthesis of phytoplankton populations with depth in the water column. Figure 7 is an illustration dataset for Station SL14. These datasets will be processed further for identification of the major phytoplankton groups. At a few stations these data will be compared with HPLC pigment analysis and phytoplankton absorption spectra determined spectrophotometrically.

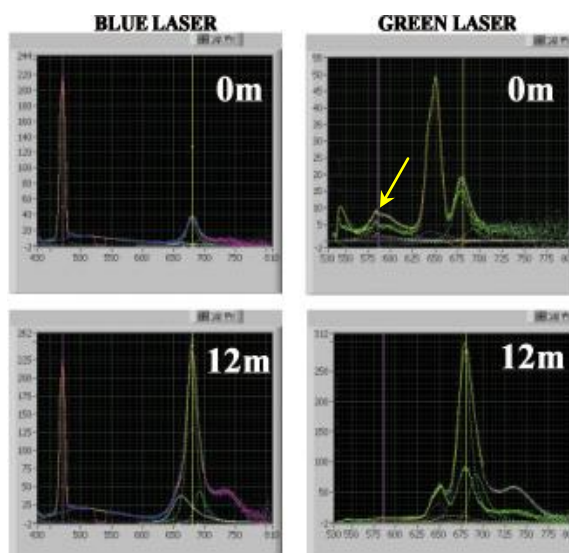


Figure7. Screen capture of data generated by the ALF for samples obtained at discrete depths at Station SL14. The arrow indicates the presence of phycoerythrin containing pigmented organisms in surface waters.

be

3) Discrete stations:

At select survey stations, water samples were collected for the following measurements: 1) Spectral absorption of CDOM using a shipboard hyperspectral waveguide capillary system 2) spectral absorbance by phytoplankton and detrital particles, 3) coccolithophore counts and calcium carbonate concentrations in separate aliquots of seawater samples filtered onto 0.4 μ m polycarbonate filters using microscopy and the inductively-coupled optical emission spectrometric technique respectively, 4) Pigment analysis by HPLC 5) suspended matter and POC estimates of material collected on pre-weighed GF/F filters (see Fig. 8 for sample data).

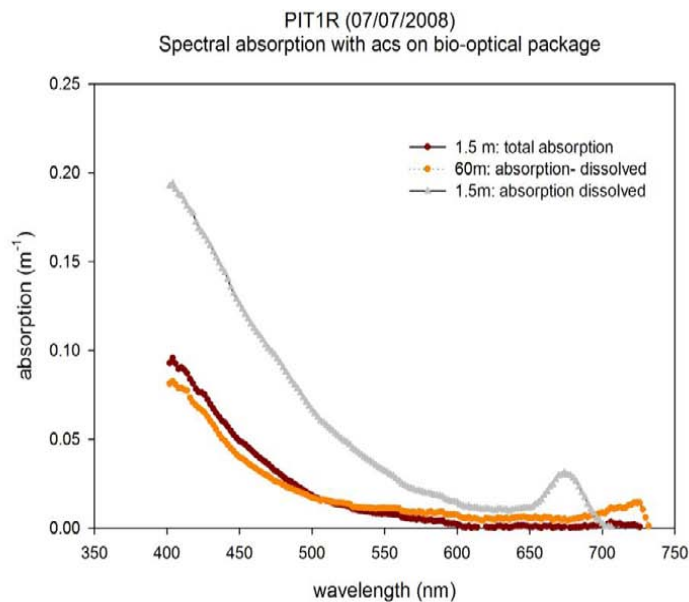


Figure 8. Absorption spectra of total particulate and CDOM measured by the ac-s at 1.5 and 60 m depths.

Anticipated outcome::

- 1) Development of more robust algorithms for estimating chlorophyll, primary productivity, CDOM, total suspended matter in the Bering Sea from satellite fields of ocean color data.
- 2) Maps showing the distribution of major summer-time phytoplankton communities and their physiological status.

9.16 Science support activities - Steve Roberts and Tom Bolmer

This is a brief summary of the performance of the Underway Science systems during the research cruise HLY0803 on the USCGC Healy, 07/03/08 – 07/31/08 from Dutch Harbor to Dutch Harbor, AK. A more complete log of events that affected the recording of data can be seen in the ELOG entries by the shipboard technicians for this leg. The Data Synopsis Report for HLY0803 has additional information.

Acoustic Data

SeaBeam 2112 Multibeam Sonar

The SeaBeam worked well for this leg. Frequent CTD's allowed us to keep the sound velocity profile valid. However, much of the cruise was in shallow water (less than 100 meters deep). This water depth is less than optimal for the SeaBeam system. This data should be aggressively edited for use in mapping. The Center Beam data that was averaged in the 1-minute average file is a good summary of that data. Similar to an event in HLY0802 a brief power outage appears to have corrupted the Magneto Optical(MO) disk and the system was down for a couple of hours to identify the problem and replace the disk. These failures should serve as a reminder as to how fragile this system has become and the need for it to be replaced.

Knudsen 320BR Sub-Bottom Profiler

The Knudsen was run in the Low Frequency "CHIRP" (3 - 6 KHz) mode for most of the cruise. These data look good. The system was run briefly in 12KHz "CHIRP" mode to aide in the search for the lost optics package but in the few passes over the search area no indications of the package were observed. Again, care must be taken when using this data, particularly if the desire is to use it for water depth. We do not recommend using subbottom profiler data for bathymetry. For this cruise the multibeam data is a better choice..

The Knudsen "KEL" formatted file saved in the SCS data directory Knudsen has the wrong internal time. The Knudsen adds about 22.8 seconds to it's internal clock each day. The time to use for this data is the SCS time stamp in the first columns of the file. The depth and location in the file are right.

We occasionally operated this system in 12kHz pinger mode to allow accurate depth determination of the multicorer.

ADCP 75

Due to the cruise being primarily in shallow water and issue with both ADCP's interfering with each other the ADCP 75 was not operated for this leg.

ADCP150

The ADCP 150 was operated for the whole leg.

Navigation

POS/MV-320

The POSMV recorded the ship's position, heading, pitch and roll with no known issues during the cruise. It was noted that DGPS was utilized by the POS/MV for most of the cruise except for the extreme Northwest region.

Ashtech ADU5

The ADU5 operated well except for an occasional drop outs which are mostly logged in the ELOG. There were also events where the receiver stopped producing heading and attitude data even though the data streams remained active. Partway into the cruise antenna cabling problems caused the system to stop generating output for a short period of time. Be sure to check the ELOG entries if you are using this data.

Sperry Gyro compasses

Two new Sperry Gyroscopes were added to the Healy to replace the old Sperry MK27s prior to this season. They have been up to 1.5 degree different from the POSMV and the ADU5 and show surprisingly large "wander" in heading. With its current behavior the systems have been shown to not be an acceptable fall back in the event of a problem with the POSMV. We do not recommend using this data. The ETs have done several tests and adjustments trying to improve the quality of the data during this cruise. We have been monitoring and generating plots for the ETs during this period.

Sea Water Flow Through data Uncontaminated Sea Water

This seemed to work satisfactorily for HLY0803.

Thermosalinographs

New primary and a spare TSGs were installed by SIO/ODF (Scott Hiller) was installed for this season in the Biochem Lab. These appeared to operate satisfactorily. Be sure to look in the ELOG for ties when the Salinity values were adjusted.

Dissolved Oxygen, Fluorometer, and Flowmeter

In addition to temperature and salinity, dissolved oxygen, fluorescence and the rate of flow of the water through the TSG were also recorded. It appears that these systems worked satisfactorily.

Meteorological Sensors

New Meteorological sensors were installed for this season by SIO/ODF (Scott Hiller.) The sensors were operated in addition to the ship's existing sensors. These sensors operated satisfactorily for the leg. For the wind speed and direction 2D ultrasonic instruments were installed on the Yard Arm and the Jack Staff.

Mapserver

A web-based real-time GIS system (Mapserver) was actively maintained and kept up-to-date with the most current science cruise data and information. Included in this activity was the monitoring, downloading and processing of near real-time MODIS chlorophyll imagery.

Gravity

Two Bell BGM-3 marine gravity meters were installed in IC/Gyro prior to this season and appeared to operate satisfactorily.

Data Logging

LDS (Lamont Data System)

The LDS data logging system was run to record and store underway data for the leg. This system logged the Navigation, SeaBeam, the SIO MET data, gravity, and web camera images.

Underway Data Distribution

At the end of the cruise a USB disk containing all the underway data along with various documentation was created and provided to the chief scientist.

Data QC

Continuously monitored all underway data streams and addressed anomalies as they became apparent.

Terrascan

Monitored and maintained the Terrascan system plus a separate laptop with a second Terrascan license. This second laptop was used to generate various ice imagery for general science use and inclusion into the Mapserver. Since we were operating in the Fairbanks, Alaska station range circle all DMSP data was collected in unencrypted mode.

Web Cameras

Web cameras were operated in Aloft Con, Aft Con and the Board of Lies. Images from the cameras were logged on LDS. In addition once an hour an image from Aloft Con was emailed to shore for use in a web site there. The Aloft Con web camera was replaced with a wider angled lens camera mid way through the cruise. This replacement is working satisfactorily

9.17 Educational Activities

The two teachers on board, Jillian Worssam (a high school teacher from Arizona supported by PolarTrek) and John Karavias (a high school science teacher from New York supported by Armada) managed several educational activities. By 'joining' various research groups, they familiarized themselves with science questions and methods. They then developed educational activities around these experiences. Some of these educational materials will require further development after the cruise. Some were shared with teachers on-shore via webinars originated

from the ship. The teachers also directed the outreach activities on St. George Island on July 11. The arrangements were made beforehand with Michele Ridgeway, a marine geophysicist who has developed a summer Ocean Science Camp on the island (see letter in Appendix). The teachers and several scientists from the Healy took part in presentations and discussions with the students during a several hour visit.

10 Appendix – Event Log; Niskin water budget; Sample depths; Letter from Michele Ridgeway

Station Stn.# Name	Event Type	Cast#	CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
1 UP1	CTD	1	Standard	7/4/08 2:08	7/4/08 2:21	54.30889	165.3536	54.31361	165.3634	80	78	Unimak Pass 1/start; end
1 UP1	BIO-OPTICS			7/4/08 2:50	7/4/08 3:05	54.3087	165.3533	54.3075	165.3538	78	76	
1 UP1	MOC			7/4/08 3:22	7/4/08 3:56	54.3166	165.379	54.3082	165.3549	60	60	
2 UP2	CTD	2	Standard	7/4/08 6:18	7/4/08 6:41	54.38532	165.1988	54.39121	165.1966	174	174	UP2 (Unimak Pass); Unimak Pass
2 UP2	CTD			7/4/08 6:55	7/4/08 7:33	54.3942	165.1927	54.4095	165.177	100	100	
3 UP4	CTD	3	Standard	7/4/08 8:40	7/4/08 8:50	54.48513	164.9593	54.48359	164.9572	86	76	start; end
3 UP4	MOC			7/4/08 9:02	7/4/08 9:33	54.4803	164.9563	54.4679	164.972	70	70	
4 UP3	CTD	4	Standard	7/4/08 10:20	7/4/08 10:35	54.41903	165.139	54.4216	165.1465	135	135	start intermediate station; end
4 UP3	BONGO	1		7/4/08 10:54	7/4/08 11:04	54.4225	165.1481	54.4225	165.1481	137	137	Dark. Oblique tow.
4 UP3	CTD	5	Zoop	7/4/08 11:14	7/4/08 11:22	54.42841	165.1503	54.43097	165.156	135	135	Zooplankton; end
4 UP3	MOC			7/4/08 11:38	7/4/08 12:20	54.4315	165.1535	54.4261	165.1534	100	100	
4 UP3	CTD	6	Radon	7/4/08 13:07	7/4/08 13:24	54.42849	165.1432	54.44013	165.1576	130	133	sta 4:start radon cast; end
4 UP3	MCORE	1		7/4/08 14:15	7/4/08 14:25	54.4217	165.1387	54.4216	165.1394	132	133	Eight tubes rigged.... very coarse sandy sediment... some shell hash...20m/min up and down. Time on Deck / End Position = Time / Position of core on bottom !
4 UP3	MCORE	2		7/4/08 14:52	7/4/08 15:00	54.4219	165.1406	54.4217	165.1412	133	133	Time / Position of core on bottom !
4 UP3	CTD	7	Prod	7/4/08 15:36	7/4/08 15:54	54.43815	165.1411	54.44498	165.143	123	122	start; end
4 UP3	BIO-OPTICS			7/4/08 16:50	7/4/08 17:10	54.352	165.138	54.3517	165.1375	138	135	
5 UAP8	CTD	8	Standard	7/4/08 19:05	7/4/08 19:25	54.86988	165.2357	54.87356	165.2335	124	125	start; end nsta 05
5 UAP8	BIO-OPTICS			7/4/08 19:29	7/4/08 19:50	54.8759	165.232	54.876	165.2477	124	125	
5 UAP8	MOC			7/4/08 20:25	7/4/08 20:56	54.8728	165.2534	54.8608	165.244	100	100	
6 UAP7	MOC			7/4/08 23:13	7/4/08 23:42	55.0818	164.8218	55.0894	164.8065	90	90	
7 UAP6	CTD	9	Standard	7/5/08 1:35	7/5/08 1:47	55.30527	164.3981	55.30404	164.3975	100	100	start nsta 07
7 UAP6	BIO-OPTICS			7/5/08 1:55	7/5/08 2:20	55.3008	164.3963	55.2993	164.3958	100	102	no ctd at uap7
7 UAP6	MOC			7/5/08 2:31	7/5/08 3:05	55.3016	164.3954	55.3138	164.363	80	80	
8 UAP5	MOC			7/5/08 5:19	7/5/08 5:48	55.5296	163.9713	55.5491	163.9496	75	75	
9 UAP4	MOC			7/5/08 7:37	7/5/08 8:08	55.7538	163.5531	55.773	163.5388	80	80	
9 UAP4	MCORE	1		7/5/08 8:39	7/5/08 8:43	55.7617	163.5311	55.7617	163.5312	92	92	8 good cores ... ~20cm penetration..well sorted fine grained silty sand...SOG=0.2kts...2900lb pullout at 20m/min in and out. Time on Deck / End Position = Time / Position of core on bottom !
9 UAP4	MCORE	2		7/5/08 9:24	7/5/08 9:29	55.7592	163.5263	55.7592	163.5264	93	93	bottom !
10 UAP3	CTD	10	Standard	7/5/08 11:38	7/5/08 11:52	55.96388	163.1384	55.96687	163.139	87	87	Standard cast Station 10; end sta 10
10 UAP3	BONGO	2		7/5/08 12:00	7/5/08 12:10	55.9673	163.1394	55.9673	163.1394	88	88	Oblique tow. Dark but with visible horizon.
10 UAP3	MOC			7/5/08 12:27	7/5/08 12:59	55.9769	163.144	55.9933	163.1484	70	70	
11 CN6	CTD	11	Standard	7/5/08 19:30	7/5/08 19:41	57.00016	163.8993	57.00043	163.9005	70	70	start sta 12; end
11 CN6	BIO-OPTICS			7/5/08 19:55	7/5/08 20:20	57.001	163.9025	56.9991	163.904	69	70	
11 CN6	MOC			7/5/08 20:28	7/5/08 20:55	56.9994	163.8889	57.0135	163.8901	55	55	
11 CN6	CTD	12	Prod	7/5/08 21:18	7/5/08 21:32	57.00125	163.9052	57.00023	163.9107	70	70	Start Prod cast; the real end for ctd12
11 CN6	CALVET			7/5/08 21:53	7/5/08 21:54	56.9971	163.9112	56.9972	163.9113	68	68	
12 CN8	CTD	13	Standard	7/6/08 0:16	7/6/08 0:27	56.70394	164.511	56.70218	164.5149	70	70	Start station 12; ctd13; end ctd 13
12 CN8	BIO-OPTICS	2		7/6/08 0:35	7/6/08 1:00	56.7021	164.5152	56.6968	164.53	75	80	
12 CN8	CALVET			7/6/08 1:19	7/6/08 1:20	56.69666	164.5303	56.69695	164.5302	70	70	
12 CN8	MOC			7/6/08 1:22	7/6/08 1:50	56.69845	164.5296	56.71634	164.5288	55	55	
13 CN10	CTD	14	Standard	7/6/08 5:02	7/6/08 5:17	56.43049	165.3034	56.43142	165.3025	85	85	start ctd14; end ctd 14
13 CN10	BIO-OPTICS			7/6/08 5:25	7/6/08 5:50	56.4301	165.3011	56.4323	165.294	85	83	
13 CN10	CALVET			7/6/08 6:07	7/6/08 6:12	56.43129	165.2817	56.43156	165.2809	84	84	
13 CN10	MCORE	1		7/6/08 6:41	7/6/08 6:44	56.4301	165.2765	56.4301	165.2766	84	84	8 tubes rigged - no useable samples... disturbed and uneven core penetration (max=~12cms).. possible hard pan / dense clay layer beneath. Time on Deck / End Position = Time / Position of core on bottom !
13 CN10	MCORE	2		7/6/08 7:08	7/6/08 7:12	56.4297	165.2767	56.4297	165.2767	83	83	bottom !
14 CN12	CTD	15	Standard	7/6/08 9:56	7/6/08 10:13	56.13798	166.1061	56.13811	166.106	109	109	start sta014; end sta014
14 CN12	BONGO	3		7/6/08 10:50	7/6/08 11:00	56.1389	166.1075	56.1389	166.1075	110	110	Dark with visible horizon
14 CN12	CTD	16	Zoop	7/6/08 10:59	7/6/08 11:04	56.145667	166.1115	56.145667	166.1115	136	136	start/log overwritten; from ctd file;
14 CN12	CALVET			7/6/08 11:10	7/6/08 11:20	56.13798	166.1061	56.13798	166.1061	100	100	end/log overwritten; from ctd file
15 CN14	CTD	17	Standard	7/6/08 13:33	7/6/08 13:53	55.84761	166.9038	55.84165	166.9062	136	136	start sta015; end sta 015
15 CN14	CALVET			7/6/08 13:55	7/6/08 14:05	55.84761	166.9038	55.84761	166.9038	100	100	
15 CN14	MOC			7/6/08 14:17	7/6/08 14:53	55.8376	166.9099	55.8552	166.9353	100	100	
16 PIT1D	SEDTRAPS			7/6/08 19:24	7/7/08 19:20	55.2878	167.9415	55.2213	168.0373	330	877	First PIT deployment
16 PIT1D	CTD	18	Prod	7/6/08 19:48	7/6/08 20:17	55.28054	167.9276	55.26851	167.9217	368	368	start ctd18 prod at PIT1; end
16 PIT1D	BIO-OPTICS			7/6/08 20:30	7/6/08 21:00	55.2627	167.9188	55.2472	167.9097	370	375	
17 CN18	CTD	19	Standard	7/6/08 23:06	7/7/08 0:42	55.30736	168.4149	55.3037	168.4242	1665	1665	First deep cast; end ctd 19
17 CN18	BIO-OPTICS			7/7/08 1:20	7/7/08 1:50	55.3013	168.4012	55.3017	168.383	1658	1655	
17 CN18	CTD	20	Radon	7/7/08 1:53	7/7/08 2:09	55.29724	168.3904	55.29489	168.3832	1602	1602	Start radon; end ctd 20 radon
17 CN18	CTD			7/7/08 1:53	7/7/08 2:09	55.298	168.3918	55.298	168.3918	1614	1614	Radon and O2 isotopes. Cast to 70m (10 bottles for Rn; 1 for O2; one other)
17 CN18	CALVET			7/7/08 2:22	7/7/08 2:26	55.29159	168.3838	55.29067	168.3862	100	100	
17 CN18	CTD	21	Standard	7/7/08 2:52	7/7/08 3:07	55.2979	168.3954	55.29782	168.3953	1602	1602	start shallow part of standard cast; end shallow part CN18

Station Stn.# Name	Event Type	Cast#	CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
17 CN18	MCORE	1		7/7/08 3:25	7/7/08 4:15	55.3047	168.3974	55.3061	168.3922	1625	1622	Eight tubes rigged.... eight good samples... approx. 25-30 cms penetration. Time on Deck / End Position = Time / Position of core on bottom ! Eight good samples... approx. 25-30 cms penetration. Wire out at core trip = 1680 meters. Time on Deck / End Position =
17 CN18	MCORE	2		7/7/08 5:12	7/7/08 5:58	55.3058	168.3984	55.3075	168.3973	1634	1630	Time / Position of core on bottom !
17 CN18	MOC			7/7/08 7:07	7/7/08 7:26	55.3115	168.401	55.32093	168.4038	100	100	
18 CN16	MOC			7/7/08 10:05	7/7/08 10:42	55.5479	167.7002	55.56395	167.7036	100	100	
18 CN16	CTD	22	Standard	7/7/08 10:46	7/7/08 11:04	55.56375	167.7035	55.5622	167.7029	137	137	start; end
18 CN16	BONGO	4		7/7/08 11:10	7/7/08 11:20	55.5626	167.7023	55.5626	167.7029	136	136	Dark with visible horizon
18 CN16	CALVET			7/7/08 11:19	7/7/08 11:24	55.5614	167.7037	55.56114	167.7042	100	100	
18 CN16	CTD	23	Zoop	7/7/08 11:44	7/7/08 11:50	55.56882	167.7133	55.56878	167.7142	138	137	start; end
19 X1	MOC			7/7/08 13:17	7/7/08 13:47	55.36894	167.8376	55.38552	167.8542	100	100	
20 X2	MOC			7/7/08 15:06	7/7/08 15:34	55.23441	167.9428	55.24823	167.9574	100	100	
20 X2	CTD	24	Prod	7/7/08 15:48	7/7/08 16:30	55.25	167.9295	55.24462	167.9624	502	511	start; end
21 PIT1R	BIO-OPTICS			7/7/08 19:55	7/7/08 20:15	55.2226	168.0087	55.2213	167.9895	740	668	
21 PIT1R	BIO-OPTICS			7/7/08 20:25	7/7/08 20:44	55.2226	168.0087	55.2213	167.9895	740	668	
22 CNN8	CTD	25	Standard	7/8/08 1:38	7/8/08 2:23	56.1495	168.70367	56.1495	168.70367	457	457	start; end
22 CNN8	BIO-OPTICS			7/8/08 2:31	7/8/08 3:20	56.1435	168.7227	56.1412	168.7408	484	501	
22 CNN8	BIO-OPTICS			7/8/08 3:00	7/8/08 3:20	56.1435	168.7227	56.1411	168.7408	484	501	
22 CNN8	CALVET			7/8/08 3:33	7/8/08 3:39	56.14161	168.7432	56.14241	168.7432	100	100	
22 CNN8	MOC			7/8/08 3:50	7/8/08 4:24	56.14563	168.7358	56.15405	168.7139	100	100	
23 CNN7	MOC			7/8/08 6:44	7/8/08 7:53	56.4523	168.2826	56.49494	168.2223	100	100	
24 CNN6	CTD	26	Standard	7/8/08 9:43	7/8/08 9:37	56.73652	167.8463	56.7351	167.8456	95	95	start standard cast; end
24 CNN6	CALVET			7/8/08 10:04	7/8/08 10:08	56.73436	167.8446	56.73382	167.8441	98	98	
24 CNN6	BONGO	5		7/8/08 10:20	7/8/08 10:30	56.7344	167.8446	56.7344	167.8446	100	100	Time of tow close to local midnight. Light level more like twilight.
24 CNN6	MOC			7/8/08 10:51	7/8/08 11:06	56.75302	167.855	56.76021	167.8578	98	98	
24 CNN6	MCORE	1		7/8/08 11:28	7/8/08 11:32	56.7623	167.8619	56.7618	167.8619	99	99	Six tubes rigged.... six good samples... approx. 25-30 cms penetration. 1465 lb pullout. End time/position = time / position of core on the bottom.
24 CNN6	MCORE	2		7/8/08 11:45	7/8/08 11:48	56.7613	167.8627	56.7613	167.8628	100	100	Six tubes rigged.... six good samples... approx. 25-30 cms penetration. 2087 lb pullout. End time/position = time / position of core on the bottom.
25 NP2/S40	CTD	27	Standard	7/8/08 22:01	7/8/08 22:07	59.13982	167.9795	59.13965	167.9794	41	41	start ctd27 mooring S40; end ctd27
25 NP2/S40	MOORING	27	Standard	7/8/08 22:23	7/8/08 22:23	59.1363	167.9823	59.1363	167.9823	42	42	Mooring Deployment; recover in 2009
26 S25	CTD	28	Standard	7/9/08 6:17	7/9/08 6:21	59.85111	167.7826	59.85105	167.7823	27	27	start ctd 28 mooring 25; end ctd 28
26 S25	MOORING			7/9/08 6:30	7/9/08 6:30	59.8515	167.7814	59.8515	167.7814	26	26	Recover in 2009
27 MN1	CTD	29	Standard	7/9/08 7:28	7/9/08 7:33	59.89789	167.9956	59.89762	167.9937	29	29	start ctd29; end ctd 29
27 MN1	CALVET			7/9/08 7:39	7/9/08 7:43	59.89764	167.9911	59.89763	167.9899	28	28	
27 MN1	MOC			7/9/08 7:53	7/9/08 8:04	59.89611	167.99	59.89355	167.9904	15	15	
28 MN2	CTD	30	Standard	7/9/08 10:08	7/9/08 10:14	59.90182	168.5977	59.90231	168.5934	39	39	start ctd 30; end ctd 030
28 MN2	CALVET			7/9/08 10:20	7/9/08 10:23	59.90252	168.5898	59.9025	168.5884	38	38	
28 MN2	BONGO	6		7/9/08 10:30	7/9/08 10:40	59.9025	168.5882	59.9025	168.5882	41	41	Close to local midnight; but horizon easily visible.
28 MN2	CTD	31	Zoop	7/9/08 10:47	7/9/08 10:51	59.89662	168.582	59.89631	168.5806	40	40	Start zoop cast; end zoop cast
28 MN2	MOC			7/9/08 10:59	7/9/08 11:19	59.8951	168.579	59.88216	168.5699	25	25	
29 MN3	CTD	32	Standard	7/9/08 13:33	7/9/08 13:41	59.89606	169.2007	59.8961	169.1987	45	45	start: ctd032; end ctd032
29 MN3	CALVET			7/9/08 13:51	7/9/08 13:55	59.89625	169.1964	59.8963	169.1956	44	44	
29 MN3	MOC			7/9/08 14:09	7/9/08 14:20	59.89219	169.1879	59.88707	169.1806	44	44	
29 MN3	CTD	33	Prod	7/9/08 14:57	7/9/08 15:08	59.89681	169.1991	59.89642	169.1982	45	45	start ctd033; end ctd033
30 MN4	CTD	34	Standard	7/9/08 17:06	7/9/08 17:14	59.89571	169.8055	59.89319	169.8073	52	52	start ctd 34; end ctd 034
30 MN4	CALVET			7/9/08 17:25	7/9/08 17:35	59.89245	169.8086	59.89167	169.8089	54	54	
30 MN4	MOC			7/9/08 17:43	7/9/08 17:59	59.8923	169.8087	59.88385	169.8142	40	40	
31 MN5	CTD	35	Radon	7/9/08 20:09	7/9/08 20:21	59.90223	170.3931	59.90411	170.3922	64	64	Start radon cast; end ctd 34 radon
31 MN5	CALVET			7/9/08 20:27	7/9/08 20:31	59.90531	170.3923	59.90674	170.3914	60	60	
31 MN5	MCORE	1		7/9/08 20:59	7/9/08 21:01	59.8992	170.3979	59.8999	170.3971	62	63	6 tubes rigged.. 5 samples recovered.. one empty...SOG>1.6kts during cast...core probably tipped over during pullout with wire angle to starboard. 1918 lb pullout .. ~25cm uneven penetration. Time / pos'n on deck = time / pos'n of core on bottom.
31 MN5	MCORE	2		7/9/08 21:20	7/9/08 21:24	59.9017	170.3946	59.9025	170.3945	62	63	8 tubes rigged.. 8 samples recovered.SOG=0.4kts during cast...good wire angle 2157 lb pullout .. 28cm penetration into soft mud / clay. Time / pos'n on deck = time / pos'n of core on bottom.
31 MN5	MOC			7/9/08 22:05	7/9/08 22:17	59.89476	170.4081	59.89469	170.4189	50	50	
32 MN6	CTD	36	Standard	7/10/08 0:28	7/10/08 0:37	59.89837	170.9988	59.89635	170.9931	71	71	Start ctd36; end cast 36
32 MN6	CALVET			7/10/08 0:45	7/10/08 0:56	59.89633	170.9883	59.8965	170.9852	69	69	
32 MN6	MOC			7/10/08 1:21	7/10/08 1:39	59.90061	170.9917	59.90846	171.0057	55	55	
33 MN7	CTD	37	Standard	7/10/08 4:05	7/10/08 4:16	59.90108	171.6033	59.89986	171.6068	70	70	start ctd37; end ctd 37
33 MN7	CALVET			7/10/08 4:31	7/10/08 4:37	59.90009	171.613	59.90077	171.6138	74	68	
33 MN7	MOC			7/10/08 4:47	7/10/08 5:09	59.90214	171.6191	59.90751	171.6445	55	55	
34 MN8	CTD	38	Standard	7/10/08 8:36	7/10/08 8:48	59.90001	172.2239	59.89899	172.2275	72	72	start CTD38; end ctd 038
34 MN8	CALVET			7/10/08 8:58	7/10/08 9:04	59.89809	172.231	59.89725	172.2303	72	72	
34 MN8	MOC			7/10/08 9:15	7/10/08 9:39	59.89194	172.2408	59.88066	172.2545	60	60	
35 W7	CTD	39	Standard	7/10/08 13:15	7/10/08 13:26	60.0043	171.0909	60.0018	171.093	71	71	start ctd 039; end ctd 039
35 W7	BONGO	7		7/10/08 13:40	7/10/08 13:50	60	171.0919	60.0033	171.0919	70	70	Pretty light out.
35 W7	CALVET			7/10/08 13:55	7/10/08 14:02	59.99196	171.1072	59.99191	171.1074	65	65	
35 W7	CTD	40	Zoop	7/10/08 14:05	7/10/08 14:13	59.99116	171.1085	59.98862	171.1096	71	71	start ctd 040; end ctd 40
36 W6	CTD	41	Standard	7/10/08 16:35	7/10/08 16:46	60.10216	170.4615	60.10133	170.4579	62	62	start ctd 41; end ctd 41
36 W6	CALVET			7/10/08 16:52	7/10/08 17:04	60.10144	170.4563	60.10144	170.4563	60	60	
37 W5/C55	CTD	42	Prod	7/10/08 18:29	7/10/08 18:39	60.17075	170.0675	60.17102	170.0667	56	56	start ctd 042; end ctd 042
37 W5/C55	MOORING			7/10/08 19:23	7/10/08 19:23	60.1723	170.0885	60.1723	170.0885	55	55	Mooring was recovered and re-deployed as station 43. Use those data for actual position
38 W4/C40	CTD	43	Standard	7/10/08 23:16	7/10/08 23:21	60.33943	169.0222	60.33942	169.0213	40	40	start ctd43 mooring; end c-40 ctd

Station	Event Type	Cast#	CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
38 W4/C40	MOORING	43	Standard	7/10/08 23:34	7/10/08 23:34	60.3367	169.0211	60.3367	169.0211	40.5	40.5	Deployment; recover in 2009 start ctd44 mooring c25; end ctd44 m
39 W1/C25	CTD	44	Standard	7/11/08 5:04	7/11/08 5:08	60.68194	167.336	60.68235	167.3368	26	26	c25
39 W1/C25	MOORING	44	Standard	7/11/08 5:16	7/11/08 5:16	60.6827	167.3376	60.6827	167.3376	25	25	Mooring Deployment; recover in 2009
40 W2	CTD	45	Standard	7/11/08 7:46	7/11/08 7:52	60.49339	168	60.49289	167.9998	30	30	start ctd45; end ctd45
40 W2	BONGO	8		7/11/08 8:05	7/11/08 8:15	60.4936	168	60.4936	168	30	30	Essentially daylight
41 W3	CTD	46	Standard	7/11/08 10:10	7/11/08 10:16	60.40015	168.6195	60.40018	168.6174	36	36	start ctd 46; end ctd 46
41 W3	CALVET			7/11/08 10:24	7/11/08 10:29	60.40029	168.6142	60.39956	168.6119	36	36	
41 W3	BONGO	9		7/11/08 10:32	7/11/08 10:42	60.4001	168.6204	60.4001	168.6204	35	35	Dark
41 W3	CTD	47	Zoop	7/11/08 10:50	7/11/08 10:54	60.39548	168.6011	60.39529	168.5996	36	36	start zoops cast (46); end zoops cast
42 W4.5	CTD	48	Prod	7/11/08 14:07	7/11/08 14:17	60.25412	169.5699	60.25451	169.5684	48	48	start ctd 48/prod also; end ctd 48
42 W4.5	CALVET			7/11/08 14:29	7/11/08 14:35	60.25502	169.5659	60.25751	169.564	47	47	
43 C55	MOORING			7/11/08 19:51	7/11/08 19:51	60.1719	170.0895	60.1719	170.0895	55	55	Mooring will be recovered in 2009
44 SL6.5/N5 CTD		49	Standard	7/12/08 4:32	7/12/08 4:40	61.96651	171.9437	61.96926	171.9417	55	55	start ctd49(moor n55); end ctd49
44 SL6.5/N5 CALVET				7/12/08 4:48	7/12/08 4:52	61.96975	171.9436	61.96989	171.9429	54	54	
44 SL6.5/N5 MOORING		49	Standard	7/12/08 5:31	7/12/08 5:31	61.9619	171.9748	61.9619	171.9748	54	54	Mooring Deployment; recover in 2009
45 SL7.5	CTD	50	Standard	7/12/08 7:02	7/12/08 7:10	62.12497	172.0001	62.12396	171.9976	52	53	start: ctd50/sta 45; end
45 SL7.5	CALVET			7/12/08 7:16	7/12/08 7:19	62.12297	171.9957	62.12254	171.9939	51	51	
45 SL7.5	MOC			7/12/08 7:34	7/12/08 7:53	62.11471	171.9844	62.10329	171.9797	51	51	
46 SL8	CTD	51	Standard	7/12/08 10:06	7/12/08 10:19	62.20331	172.7012	62.20625	172.7052	58	58	start ctd 51; end ctd 51/sta 46
46 SL8	CTD	51	Radon	7/12/08 10:08	7/12/08 10:08	61.2033	172.7012	61.2033	172.7012	58	58	Radon and triple O2 isotopes collected
46 SL8	CALVET			7/12/08 10:33	7/12/08 10:40	62.20931	172.7101	62.20932	172.7101	54	54	during standard cast
46 SL8	BONGO	10		7/12/08 10:45	7/12/08 10:55	62.2098	172.7108	62.2098	172.7108	58	58	Basically twilight
												Eight tubes rigged... extremely erratic pullout wire angle to starboard.. 7 knot wind on starboard beam. all eight samples disturbed -- discarded eight samples for re-try.End time/pos'n =
46 SL8	MCORE	1		7/12/08 11:15	7/12/08 11:19	62.2018	172.7127	62.2088	172.7138	58	58	time/position of core on bottom.
												Eight tubes rigged. very slight pullout wire angle to starboard. 7 knot wind on starboard beam. One minute bottom time with 6 of 8 suitable samples. 1684lb pullout - 1 min on bottom. 25 -30 cms penetration. End time/pos'n = time/pos'n
46 SL8	MCORE	2		7/12/08 11:41	7/12/08 11:44	62.2033	172.7133	62.2038	172.7136	58	58	of core on bottom.
												Eight tubes rigged. very slight pullout wire angle to starboard. 7 knot wind on starboard beam. One minute bottom time with 6 of 8 suitable samples. 2297 lb pullout - 1 min on bottom. 25 -30 cms penetration. End time/pos'n = time/pos'n
46 SL8	MCORE	3		7/12/08 12:00	7/12/08 12:03	62.205	172.7096	62.2054	172.7088	58	58	of core on bottom.
46 SL8	CTD	52	Prod	7/12/08 12:19	7/12/08 12:34	62.20405	172.701	62.20543	172.6964	58	58	start ctd 52/sta 46; end: ctd 52/sta 46
47 SL5.5	CTD	53	Standard	7/12/08 18:19	7/12/08 18:27	61.98258	170.6022	61.98217	170.6015	49	49	start ctd 053/sta 47; end ctd 53
47 SL5.5	CALVET			7/12/08 18:34	7/12/08 18:38	61.98202	170.6009	61.98222	170.6006	48	48	
47 SL5.5	MOC			7/12/08 18:50	7/12/08 19:07	61.98039	170.5979	61.97137	170.5923	48	48	
48 SL4.5	CTD	54	Standard	7/12/08 21:12	7/12/08 21:19	61.91449	169.9004	61.91449	169.9004	46	46	start ctd54; end ctd54
48 SL4.5	CALVET			7/12/08 21:23	7/12/08 21:27	61.91539	169.9003	61.91568	169.9003	49	49	
49 SL3.5/N4 CTD		55	Standard	7/12/08 23:44	7/12/08 23:50	61.79715	169.3074	61.79716	169.3057	42	42	start ctd55 moor n-40; end ctd 55
49 SL3.5/N4 CALVET				7/12/08 23:57	7/13/08 0:00	61.79704	169.3037	61.79707	169.3027	41	41	
49 SL3.5/N4 MOORING		44	Standard	7/13/08 0:30	7/13/08 0:30	61.8055	169.2787	61.8055	169.2787	40	40	Mooring Deployment; recover in 2009
49 SL3.5/N4 MOC				7/13/08 0:37	7/13/08 0:53	61.8065	169.2862	61.80906	169.3031	41	41	
50 SL2.5	CTD	56	Standard	7/13/08 3:12	7/13/08 3:17	61.77159	168.4956	61.77159	168.4956	36	36	start ctd 56; end ctd 56
50 SL2.5	CALVET			7/13/08 3:23	7/13/08 3:26	61.7717	168.4877	61.77182	168.4853	33	33	
51 SL1.5/N2 CTD		57	Standard	7/13/08 6:38	7/13/08 6:42	61.699833	167.45617	61.699833	167.45617	30	30	start ctd 57; end ctd 57
51 SL1.5/N2 MOORING				7/13/08 6:54	7/13/08 6:54	61.7001	167.4519	61.7001	167.4519	25.6	25.6	Mooring will be recovered in 2009
												Dark as it's getting. No other events at
52 B1	BONGO	11		7/13/08 10:00	7/13/08 10:10	61.0475	167.7863	61.0475	167.7863	26	26	this station.
53 NP1	CTD	58	Standard	7/13/08 17:12	7/13/08 17:18	59.44604	167.7969	59.44578	167.7949	37	37	start ctd058; end ctd 58
53 NP1	CALVET			7/13/08 17:23	7/13/08 17:26	59.44534	167.7933	59.44505	167.7923	38	38	
53 NP1	MOC			7/13/08 17:37	7/13/08 17:48	59.44201	167.7869	59.43741	167.7799	38	38	
54 NP2/S40 CTD		59	Standard	7/13/08 20:52	7/13/08 20:57	59.12659	168.0044	59.12701	168.0039	41	41	start ctd 59; end ctd 59
54 NP2/S40 CALVET				7/13/08 21:07	7/13/08 21:13	59.12724	168.003	59.1269	168.0025	44	44	
55 NP3	CTD	60	Standard	7/14/08 2:50	7/14/08 2:56	58.8278	168.1587	58.82911	168.1591	46	46	start ctd 60; end ctd 60
55 NP3	CALVET			7/14/08 3:01	7/14/08 3:05	58.8297	168.1597	58.83028	168.1597	47	47	
55 NP3	MOC			7/14/08 3:18	7/14/08 3:30	58.83043	168.1456	58.82474	168.1388	43	43	
56 NP4/S55 CTD		61	Standard	7/14/08 5:48	7/14/08 5:57	58.60581	168.4185	58.60447	168.4135	55	55	start ctd 61 moor s55; end ctd 61
56 NP4/S55 CALVET				7/14/08 6:03	7/14/08 6:08	58.60288	168.411	58.60157	168.4092	55	55	
56 NP4/S55 MOORING				7/14/08 6:40	7/14/08 6:40	58.5896	168.3941	58.5896	168.3941	55.3	55.3	Mooring will be recovered in 2009
57 NP5	CTD	62	Standard	7/14/08 8:34	7/14/08 8:46	58.36314	168.6973	58.36138	168.6916	68	68	start ctd 62; end ctd 62
57 NP5	CALVET			7/14/08 8:58	7/14/08 8:58	58.35956	168.6845	58.35954	168.6845	65	65	
57 NP5	MOC			7/14/08 9:06	7/14/08 9:30	58.35503	168.6835	58.34164	168.6791	50	50	
58 NP6	CTD	63	Standard	7/14/08 11:29	7/14/08 11:39	58.12969	168.968	58.1284	168.9622	68	68	start ctd63; end ctd 63
58 NP6	CALVET			7/14/08 11:47	7/14/08 11:51	58.12744	168.9586	58.12629	168.9561	70	70	
58 NP6	BONGO	12		7/14/08 12:10	7/14/08 12:20	58.1306	168.9698	58.1306	168.9698	70	70	Dark & foggy
58 NP6	CTD	64	Zoop	7/14/08 12:14	7/14/08 12:20	58.11594	168.9456	58.11538	168.942	70	70	start ctd 64; end ctd 64
59 NP7	CTD	65	Radon	7/14/08 14:12	7/14/08 14:26	57.90056	169.2341	57.89914	169.2269	70	70	start ctd 65; end ctd 65
												10 bottles for radon; one bottle for triple O2 isotopes (15m); one bottle for Karen
59 NP7	CTD	65	Radon	7/14/08 14:12	7/14/08 14:28	57.9003	169.2318	57.9003	169.2318	66	66	(50m)
59 NP7	CALVET			7/14/08 14:32	7/14/08 14:36	57.8982	169.2236	57.89782	169.2216	70	70	
59 NP7	CTD	66	Standard	7/14/08 15:00	7/14/08 15:09	57.89304	169.2106	57.89104	169.2072	70	70	start ctd66; end ctd66
59 NP7	MOC			7/14/08 15:23	7/14/08 15:45	57.891	169.2132	57.89988	169.2323	70	70	
59 NP7	CTD	67	Prod	7/14/08 16:05	7/14/08 16:17	57.89865	169.2389	57.89625	169.2357	66	68	start prod cast; end ctd 67
												8 tubes rigged.. 1 suitable sample recovered.SOG=0.7kts during cast...port wire angle with 1242 lb pullout . <10cms penetration into clayey silt. Time / pos'n
59 NP7	MCORE	1		7/14/08 16:38	7/14/08 16:43	57.8941	169.2317	57.8939	169.231	66	66	on deck = time / pos'n of core on bottom.

Station Stn.# Name	Event Type	Cast#	CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
59 NP7	MCORE	2		7/14/08 17:15	7/14/08 17:19	57.8924	169.2304	57.8924	169.2303	66	66	6 tubes rigged... 5 suitable samples recovered.SOG=0.2kts during cast...good wire angle with 2711 lb pullout . ~18cms penetration into clayey silt. 1.5 min bottom time. Time / pos'n on deck = time / pos'n of core on bottom.
59 NP7	MCORE	3		7/14/08 17:36	7/14/08 17:39	57.8931	169.2316	57.8933	169.2321	66	66	6 tubes rigged... 5 suitable samples recovered.SOG=0.2kts first then 0.9kts at windy pullout... port wire angle with 4041 lb pullout ! .. ~18cms penetration into clayey silt. 1.5 min bottom time. Time / pos'n on deck = time / pos'n of core on bottom.
60 NP8	CTD	68	Standard	7/14/08 19:40	7/14/08 19:50	57.67154	169.5188	57.67255	169.5182	71	71	start ctd68; end ctd 68
60 NP8	CALVET			7/14/08 19:57	7/14/08 20:02	57.67292	169.5173	57.67295	169.5168	70	70	
61 NP9	CTD	69	Standard	7/14/08 22:07	7/14/08 22:16	57.44349	169.7836	57.44464	169.7806	66	66	67 start ctd 69; end ctd 69
61 NP9	CALVET			7/14/08 22:23	7/14/08 22:28	57.44478	169.7786	57.44516	169.777	64	64	
61 NP9	MOC			7/14/08 22:36	7/14/08 22:52	57.44315	169.7757	57.43768	169.7737	50	55	
62 NP10	CTD	70	Standard	7/15/08 0:56	7/15/08 1:01	57.20703	170.0329	57.20631	170.0299	39	39	start ctd 70; end ctd 70
62 NP10	CALVET			7/15/08 1:10	7/15/08 1:12	57.20555	170.0244	57.20535	170.0226	41	41	
62 NP10	MOC			7/15/08 1:23	7/15/08 1:35	57.20297	170.0151	57.20026	170.0079	30	30	
63 NP11	CTD	71	Standard	7/15/08 3:37	7/15/08 3:48	56.9757	170.2814	56.97448	170.2733	72	72	72 start ctd 71; end ctd 71
63 NP11	CALVET			7/15/08 3:55	7/15/08 3:59	56.97318	170.2705	56.97198	170.2701	72	72	
63 NP11	MOC			7/15/08 4:10	7/15/08 4:26	56.96676	170.2677	56.9593	170.2645	72	72	
64 NP12	CTD	72	Standard	7/15/08 6:19	7/15/08 6:32	56.74408	170.5502	56.74024	170.5478	110	110	start ctd72; end ctd 72
64 NP12	CALVET			7/15/08 6:38	7/15/08 6:44	56.73859	170.5473	56.73786	170.5479	100	100	
64 NP12	MOC			7/15/08 6:51	7/15/08 7:24	56.73428	170.5479	56.71418	170.5385	100	100	
65 NP13	CTD	73	Standard	7/15/08 9:48	7/15/08 10:02	56.51832	170.793	56.51553	170.7924	121	121	start ctd73; end ctd 73
65 NP13	CALVET			7/15/08 10:12	7/15/08 10:18	56.51398	170.7914	56.51004	170.7934	100	100	
65 NP13	BONGO	13		7/15/08 10:15	7/16/08 10:25	56.5145	170.7919	56.5145	170.7918	122	122	
65 NP13	CTD	74	Zoop	7/15/08 10:38	7/15/08 10:43	56.50388	170.7974	56.50303	170.7971	122	122	start zoops cast; end ctd 074
65 NP13	MOC			7/15/08 10:47	7/15/08 11:33	56.50172	170.7977	56.47804	170.8188	100	100	
66 NP14	CTD	75	Standard	7/15/08 13:40	7/15/08 13:58	56.28513	171.0565	56.28712	171.0589	136	136	start ctd75; end ctd75
66 NP14	CALVET			7/15/08 14:05	7/15/08 14:11	56.28756	171.0606	56.28807	171.0618	100	100	
66 NP14	CTD	76	Prod	7/15/08 14:27	7/15/08 14:51	56.28816	171.0648	56.28502	171.0713	136	136	start prod cast; end prodcast
66 NP14	MOC			7/15/08 15:01	7/15/08 15:33	56.28132	171.0665	56.27402	171.0399	100	100	
67 PIT 2	SEDTRAPS			7/15/08 17:35	7/16/08 17:52	56.244	171.1084	56.2755	171.2362	300	140	Second Sediment Trap deployment start ctd after Pat's Sediment Trap; end
67 PIT2D	CTD	77	Standard	7/15/08 17:59	7/15/08 18:24	56.25576	171.1246	56.25936	171.129	235	215	PITD1
68 NP15	CTD	78	Standard	7/15/08 20:10	7/15/08 21:20	56.0551	171.3073	56.06511	171.3189	2755	2755	start ctd 78; end ctd 78
68 NP15	MOC			7/15/08 22:27	7/15/08 22:55	56.12275	171.3723	56.13996	171.3754	100	100	
68 NP15	CALVET			7/15/08 22:58	7/15/08 23:06	56.14155	171.3758	56.14485	171.3773	100	100	
68 NP15	CALVET			7/15/08 23:21	7/15/08 23:22	56.14909	171.3813	56.14929	171.3814	100	100	
68 NP15-B	CALVET			7/16/08 0:43	7/16/08 0:48	56.05348	171.3025	56.05376	171.3028	100	100	
68 NP15	CTD	79	Radon	7/16/08 1:06	7/16/08 1:20	56.0556	171.3025	56.05777	171.302	2751	2751	start radon cast; end radon cast
68 NP15	MCORE	1		7/16/08 1:34	7/16/08 3:03	56.0573	171.3028	56.0587	171.3121	2749	2768	8 tubes rigged. 7 suitable samples recovered. Tube 4 missing at recovery (bellyband / failed glue) SOG=0.2kts. good wire angle with 3550 lb pullout ! . 46cms penetration into clay. 1.5 min bottom time. Time / pos'n on deck = time / pos'n of core on bottom.
68 NP15	MCORE	2		7/16/08 4:52	7/16/08 6:10	56.0575	171.301	56.0653	171.3047	2742	2745	8 tubes rigged. 8 suitable samples recovered. SOG=0.4kts. Good wire angle w/ 3328 lb pullout ! Delayed recovery - wire jumped sheave at 186m wire out. 46cms penetration into clay. 1.5 mins bottom time. Time / pos'n on deck = time / pos'n of core on bottom.
69 EL3	BONGO	14		7/16/08 12:45	7/16/08 12:55	56.2783	171.1634	56.2783	171.1634	142	142	Bongo only at this station
71 NP14.5	CTD	80	Standard	7/16/08 23:24	7/16/08 23:56	56.173667	171.20867	56.173667	171.23233	2130	2130	start ctd 80; end ctd 80
72 NP13.5	CTD	81	Standard	7/17/08 1:51	7/17/08 2:11	56.403	170.92667	56.40284	170.9264	126	126	late start ctd081; end ctd81
73 NP12.5	CTD	82	Standard	7/17/08 3:58	7/17/08 4:16	56.62959	170.6742	56.62567	170.6732	114	114	start ctd82; end ctd 82
73 NP12.5	MOC			7/17/08 4:26	7/17/08 4:54	56.62262	170.6794	56.61428	170.6999	100	100	
74 NP11.5	CTD	83	Radium	7/17/08 6:43	7/17/08 6:58	56.86089	170.4196	56.85623	170.413	100	100	start Radium cast 3; end ctd 83 radium
74 NP11.5	MOC			7/17/08 7:05	7/17/08 7:35	56.8537	170.4136	56.84085	170.4281	85	85	
75 NP10.5	CTD	84	Radium	7/17/08 9:33	7/17/08 9:41	57.09328	170.1561	57.09285	170.1538	41	41	start ctd84 radium; end ctd84 no radium
75 NP10.5	CTD	84	Zoop	7/17/08 9:54	7/17/08 10:03	57.09249	170.15	57.0933	170.1486	41	41	combined std and zoops cast; end ctd 85
75 NP10.5	BONGO	15		7/17/08 10:20	7/17/08 10:30	57.0929	170.1545	57.0929	170.1545	39	39	
75 NP10.5	MOC			7/17/08 10:34	7/17/08 10:49	57.0871	170.1698	57.08587	170.1838	25	25	
75 NP10.5	CTD	85	Prod	7/17/08 11:14	7/17/08 11:18	57.09255	170.1642	57.09291	170.1634	41	41	prod cast; 3 AM; end prod cast
76 NP11	CTD	86	Standard	7/18/08 6:23	7/18/08 6:35	56.97878	170.2791	56.97728	170.2768	73	73	start ctd86; end ctd 86
77 NP11.5	CTD	87	Fe	7/18/08 7:40	7/18/08 8:45	56.85	170.4167	56.85	170.4167	102	102	Done in conjunction with CTD 87
77 NP11.5	CTD	87	Standard a	7/18/08 9:00	7/18/08 9:10	56.83773	170.411	56.83773	170.411	100	100	start ctd 87; end ctd 87
78 NP12.5	CTD	88	Fe	7/18/08 10:50	7/18/08 11:40	56.6333	170.6667	56.6333	170.6833	114	114	Done in conjunction with CTD88
78 NP12.5	CTD	88	Standard a	7/18/08 11:58	7/18/08 12:12	56.63053	170.6937	56.62717	170.6976	115	115	start ctd 88; end ctd 88
78 NP12.5	CALVET			7/18/08 12:20	7/18/08 12:25	56.62665	170.7015	56.62665	170.7015	100	100	
78 NP12.5	BONGO	16		7/18/08 12:30	7/18/08 12:40	56.6267	170.7006	56.6267	170.7006	116	116	
78 NP12.5	CTD	89	Zoop	7/18/08 12:53	7/18/08 12:59	56.62286	170.7256	56.62237	170.7268	115	115	start ctd 89; end ctd 89
78 NP12.5	MOC			7/18/08 13:08	7/18/08 13:30	56.62223	170.7347	56.6178	170.7569	100	100	
79 NP 14	CTD	90	Fe	7/18/08 15:45	7/18/08 16:40	56.2833	171.0667	56.2833	171.0667	138	138	In conjunction with CTD 90
79 NP14	CTD	90	Prod	7/18/08 17:00	7/18/08 17:22	56.27102	171.0751	56.26771	171.0803	140	140	start ctd 90; end; ctd 90
79 NP14	CTD	91	Standard	7/18/08 18:16	7/18/08 18:34	56.27932	171.0497	56.27402	171.0535	137	137	start; ctd 91; end; ctd 91
79 NP14	MCORE	1		7/18/08 19:07	7/18/08 19:13	56.287	171.0472	56.2868	171.0477	140	140	6 tubes rigged. 6 suitable samples recovered. SOG=0.3kts. Good wire angle w/ 4240 lb pullout ! 25cms penetration into clay. 1.5 mins bottom time. Time / pos'n on deck = time / pos'n of core on bottom.

Station Stn.# Name	Event Type	Cast# CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
79 NP14	MCORE	2	7/18/08 19:31	7/18/08 19:38	56.2877	171.0479	56.2872	171.0471	137	139	8 tubes rigged. 8 suitable samples recovered. SOG=0.4kts. Good wire angle w/ 4595 lb pullout ! 15 cms penetration into clay. 1.5 mins bottom time. Time / pos'n on deck = time / pos'n of core on bottom.
80 NP15.5	CTD	92 Fe	7/18/08 22:30	7/18/08 23:35	55.9167	171.45	55.9167	171.45	3032	3032	In conjunction with CTD 92
80 NP15.5	CTD	92 Standard	7/18/08 23:51	7/19/08 0:25	55.92019	171.4473	55.9241	171.4391	3031	3031	start ctd 92; end ctd 92
81 LS1_1	CTD	93 Standard	7/19/08 4:29	7/19/08 4:49	56.09095	170.3484	56.08728	170.3541	150	150	start; end
82 LS1_3	CTD	94 Standard	7/19/08 10:46	7/19/08 10:56	56.45	171.3622	56.45	171.3622	152	152	start; ctd94; end; ctd94
82 LS1_3	CALVET		7/19/08 11:10	7/19/08 11:15	56.44639	171.3597	56.44642	171.3598	100	100	
82 LS1_3	BONGO	17	7/19/08 11:30	7/20/08 11:40	56.4526	171.368	56.4513	171.368	152	152	
82 LS1_3	CTD	95 Zoop	7/19/08 11:43	7/19/08 11:50	56.44761	171.3636	56.44521	171.3625	154	154	start; ctd 95; end; ctd 95
82 LS1_3	MOC		7/19/08 12:01	7/19/08 12:27	56.4429	171.3662	56.4433	171.3904	100	100	
83 NP11	CTD	96 Prod	7/19/08 17:23	7/19/08 17:37	56.9795	170.2918	56.9808	170.2915	70	70	start; ctd 96; end; ctd 96
83 NP11	CTD	97 Standard	7/19/08 18:35	7/19/08 18:46	56.98674	170.2901	56.9881	170.2899	67	67	start; ctd 97; end
84 LS1_4	CTD	98 Standard	7/20/08 5:50	7/20/08 5:50	56.569	172.675	56.569	172.675	153	153	start; end
85 LS1_5	CTD	99 Standard	7/20/08 9:01	7/20/08 9:01	56.697	173.213	56.697	173.213	155	155	start; end
86 LS1_6	CALVET		7/20/08 10:35	7/20/08 10:42	56.88256	173.3031	56.87917	173.3048	100	100	
86 LS1_6	BONGO	18	7/20/08 11:00	7/20/08 11:10	56.8822	173.3032	56.8822	173.3032	160	160	
86 LS1_6	CTD	100 Zoop	7/20/08 11:08	7/20/08 11:08	56.875	173.324	56.875	173.324	175	175	start; end
86 LS1_6	MOC		7/20/08 11:20	7/20/08 11:44	56.87146	173.3306	56.86508	173.3544	100	100	
86 LS1_6	CTD	101 Standard	7/20/08 12:24	7/20/08 12:24	56.884	173.307	56.884	173.307	163	163	start; end
86 LS1_6	CTD	102 Fe	7/20/08 13:13	7/20/08 14:10	56.865	173.315	56.865	173.315	188	188	In conjunction with CTD 102
86 LS1_6	CTD	102 Prod	7/20/08 14:36	7/20/08 14:36	56.852	173.314	56.852	173.314	174	174	start; end
86 LS1_6	BIO-OPTICS		7/20/08 15:23	7/20/08 15:43	56.8835	173.3008	56.8858	173.302	178	175	
86 LS1_6	BIO-OPTICS		7/20/08 15:50	7/20/08 16:14	56.8742	173.2983	56.8745	173.2985	157	159	
86 LS1_6	MCORE	1	7/20/08 17:00	7/20/08 17:08	56.8419	173.3095	56.8417	173.3093	163	163	6 tubes rigged..SOG >1.7kts at pullout with sudden jerk to 4000lbs. Only one suitable sample .. five disturbed samples. Time / pos'n at end = time / pos'n of core on bottom
86 LS1_6	MCORE	2	7/20/08 17:45	7/20/08 17:49	56.8336	173.3156	56.833	173.3156	163	163	8 tubes rigged..SOG=0.5kts with low 2850 lb pullout at 10m/min wire speed. At least six disturbed samples. Very shallow uneven core penetration. Time / pos'n at end = time / pos'n of core on bottom
87 PIT3	SEDTRAPS		7/21/08 3:01	7/22/08 20:00	58.42	174.4686	58.465	174.565	210	200	Third trap deployment
87 PIT3D	CTD	103 Standard	7/21/08 3:17	7/21/08 3:17	58.417	174.453	58.417	174.453	176	176	start; end
88 P14_5	CTD	104 Standard	7/21/08 8:19	7/21/08 8:19	57.494	175.243	57.494	175.243	3493	3493	start; end
89 P14_4.5	CALVET		7/21/08 11:05	7/21/08 11:15	57.80229	175.0217	57.80024	175.0257	100	100	
89 P14_4.5	BONGO	19	7/21/08 11:15	7/21/08 11:25	57.8024	175.0218	57.8024	175.0218	2665	2665	
89 P14_4.5	CTD	105 Zoop	7/21/08 11:44	7/21/08 11:44	57.797	175.043	57.797	175.043	2568	2568	start; end
89 P14_4.5	MOC		7/21/08 12:00	7/21/08 12:29	57.80764	175.0254	57.80767	175.0254	100	100	
89 P14_4.5	CTD	106 Fe	7/21/08 12:43	7/21/08 13:38	57.81	175	57.81	175	2371	2371	Prior to CTD 106
89 P14_4.5	CTD	106 Standard	7/21/08 14:06	7/21/08 14:06	57.812	174.983	57.812	174.983	2327	2327	start; end
89 P14_4.5	CTD	107 Prod	7/21/08 15:07	7/21/08 15:07	57.828	174.994	57.828	174.994	2250	2250	start; end
89 P14_4.5	CTD	108 Radon	7/21/08 16:25	7/21/08 16:45	57.8253	174.9427	57.8253	174.9427	2195	2195	Radon and triple O2 isotopes collected
89 P14_4.5	CTD	108 Radon	7/21/08 16:32	7/21/08 16:32	57.825	174.944	57.825	174.944	2194	2194	start; end
89 P14_4.5	BIO-OPTICS		7/21/08 16:55	7/21/08 17:12	57.788	174.9088	57.818	174.8943	2210	2271	
89 P14_4.5	BIO-OPTICS		7/21/08 17:21	7/21/08 17:46	57.8217	174.9088	57.822	174.9095	2191	2196	
89 P14_4.5	CTD	109 Standard	7/21/08 18:44	7/21/08 18:44	57.817	174.892	57.817	174.892	2450	2450	start; end
90 P14_4	CTD	110 Standard	7/21/08 21:41	7/21/08 21:41	57.999	174.851	57.999	174.851	3005	3005	start; end
90 P14_4	MCORE	1	7/21/08 23:05	7/22/08 0:15	58.007	174.873	57.9963	174.8524	3008	3010	8 tubes rigged..SOG=0.9kts with 4487 lb pullout at 20m/min after 2 min settling at 60 meters off bottom. Added four 75lb CTD weights to multicore frame for 50m/min descent. All eight tubes overpenetrated. Time/pos'n at end = time / pos'n of core on bottom
91 P14_3	CTD	111 Fe	7/22/08 5:12	7/22/08 6:08	58.4867	174.5417	58.4867	174.5417	236	236	In conjunction with CTD 111
91 P14_3	CTD	111 Standard	7/22/08 6:22	7/22/08 6:22	58.485	174.518	58.485	174.518	200	200	start; end
92 P14_2	CTD	112 Fe	7/22/08 10:30	7/22/08 11:15	58.9983	173.99	58.9983	173.99	126	126	In conjunction with CTD 112
92 P14_2	CTD	112 Standard	7/22/08 11:29	7/22/08 11:40	58.99261	173.9858	58.99074	173.9855	126	126	start; ctd 112; end; ctd 112
92 P14_2	CALVET		7/22/08 11:48	7/22/08 11:54	58.98932	173.9856	58.98786	173.9861	100	100	
92 P14_2	BONGO	20	7/22/08 12:07	7/22/08 12:17	58.9893	173.9856	58.9893	173.9856	126	126	
92 P14_2	MOC		7/22/08 12:26	7/22/08 12:56	58.99967	174.0014	59.01491	174.0112	100	100	
92 P14_2	CTD	113 Zoop	7/22/08 13:24	7/22/08 13:30	59.01328	174.0076	59.01243	174.0078	123	123	start; ctd 113; end; ctd 113
92 P14_2	MCORE	1	7/22/08 13:45	7/22/08 13:50	59.0108	174.0083	59.0103	174.0086	123	123	6 tubes rigged. 4 suitable samples. SOG=0.4kts with good 2069 lb pullout and 10cms penetration into dense clay. Time/pos'n end = time/pos'n of core on the bottom.
92 P14_2	MCORE	2	7/22/08 14:08	7/22/08 14:14	59.0092	174.0098	59.0089	174.0103	125	125	6 tubes rigged. 4 suitable samples. SOG=0.4kts with good 1579 lb pullout and 10cms penetration into dense clay. Time/pos'n end = time/pos'n of core on the bottom.
92 P14_2	MCORE	3	7/22/08 14:35	7/22/08 14:41	59.0075	174.0124	59.0076	174.0126	125	125	4 tubes rigged. 4 suitable samples. SOG=0.4kts with good 2028 lb pullout and 15cms penetration into dense clay. Time/pos'n end = time/pos'n of core on the bottom.
92 P14_2	MCORE	4	7/22/08 15:00	7/22/08 15:06	59.007	174.0139	59.0068	174.0143	124	124	4 tubes rigged. 4 suitable samples. SOG=0.3kts with good 2287 lb pullout and 15cms penetration into dense clay. Time/pos'n end = time/pos'n of core on the bottom.
92 P14_2	CTD	114 Prod	7/22/08 15:21	7/22/08 16:14	59.00651	174.0148	59.00709	174.0154	124	124	start; ctd 114; end; ctd 114
92 P14_2	BIO-OPTICS		7/22/08 17:11	7/22/08 17:20	59.0083	174.011	59.0048	174.0098	120	122	
92 P14_2	BIO-OPTICS		7/22/08 17:25	7/22/08 17:45	59.0053	174.0117	59.0048	174.0098	120	23	
92 P14_2	CTD	115 Radium	7/22/08 17:53	7/22/08 18:02	59.00492	174.0093	59.00481	174.0081	124	124	start; ctd 115; end; ctd 115
93 P14_1	CTD	116 Fe	7/22/08 21:50	7/22/08 22:50	59.4533	173.4167	59.4533	173.4167	103	103	Done in conjunction with CTD 116

Station	Event Type	Cast#	CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
93 P14_1	CTD	116	Standard	7/22/08 23:06	7/22/08 23:19	59.4562	173.4191	59.45715	173.4196	104	104	start; end
93 P14_1	BIO-OPTICS			7/22/08 23:30	7/22/08 23:45	59.4578	173.4212	59.4593	173.4267	102	106	
93 P14_1	BIO-OPTICS			7/22/08 23:50	7/23/08 0:15	59.4578	173.4245	59.4613	173.0433	103	102	
94 MN10	CTD	117	Standard	7/23/08 3:08	7/23/08 3:20	59.90043	173.4002	59.90061	173.4013	87	87	start; end
94 MN10	BIO-OPTICS			7/23/08 3:26	7/23/08 3:35	59.9007	173.4015	59.9032	173.4043	86	87	
94 MN10	BIO-OPTICS			7/23/08 3:44	7/23/08 4:02	59.9035	173.4032	59.905	173.4042	85	87	
94 MN10	CALVET			7/23/08 4:25	7/23/08 4:31	59.90755	173.4077	59.90826	173.4078	84	84	
95 MN9	CTD	118	Fe	7/23/08 6:54	7/23/08 6:54	59.9117	172.795	59.9067	172.795	74	72	Done in Conjunction with CTD 118
95 MN9	CTD	118	Standard	7/23/08 7:41	7/23/08 7:51	59.90901	172.7864	59.90939	172.7845	75	75	start ctd 118; end ctd
95 MN9	CALVET			7/23/08 7:58	7/23/08 8:02	59.90937	172.7834	59.90923	172.7829	70	70	
95 MN9	BIO-OPTICS			7/23/08 8:10	7/23/08 8:30	59.9095	172.7837	59.925	172.7517	72	73	
96 MN8	CTD	119	Standard	7/23/08 10:16	7/23/08 10:26	59.89646	172.1972	59.89622	172.1943	73	73	start; ctd 119; end; ctd 119
96 MN8	CALVET			7/23/08 10:32	7/23/08 10:39	59.89624	172.1926	59.89584	172.1918	70	70	
96 MN8	BONGO	21		7/23/08 10:49	7/23/08 10:59	59.8965	172.1976	59.8965	172.1976	73	73	
96 MN8	CTD	120	Zoop	7/23/08 11:11	7/23/08 11:18	59.90215	172.211	59.90155	172.2099	73	73	start; ctd 120; end; ctd 120
96 MN8	MOC			7/23/08 11:24	7/23/08 11:34	59.90124	172.209	59.90124	172.209	55	56	
97 MN7	CTD	121	Standard	7/23/08 13:32	7/23/08 13:43	59.89647	171.5962	59.89456	171.5959	70	70	start; ctd 121; end; ctd 121
97 MN7	CALVET			7/23/08 13:53	7/23/08 13:57	59.89348	171.5962	59.89301	171.5965	71	71	
98 MN6	CTD	122	Prod	7/23/08 15:57	7/23/08 16:10	59.89937	170.9994	59.89737	171.0011	69	69	start; ctd 122; end; ctd 122
98 MN6	CALVET			7/23/08 16:19	7/23/08 16:26	59.89616	171.002	59.89511	171.0024	70	70	
98 MN6	BIO-OPTICS			7/23/08 16:28	7/23/08 16:45	59.8922	171.0045	59.8923	171.0042	69	68	
98 MN6	BIO-OPTICS			7/23/08 16:50	7/23/08 17:12	59.8925	171.0042	59.8922	171.0045	69	70	
98 MN6	BIO-OPTICS			7/23/08 17:20	7/23/08 17:41	59.889	171.0098	59.8892	171.0099	70	71	
98 MN6	CTD	123	Standard	7/23/08 17:30	7/23/08 17:40	59.888	171.0115	59.88724	171.0143	69	69	start; ctd 123; end; ctd 123
99 MN5	CTD	124	Standard	7/23/08 19:45	7/23/08 19:53	59.89987	170.4004	59.89919	170.4019	62	62	start; end
99 MN5	CALVET			7/23/08 20:08	7/23/08 20:15	59.90083	170.4057	59.90084	170.4057	62	62	
99 MN5	BIO-OPTICS			7/23/08 20:30	7/23/08 20:45	59.9037	170.3932	59.9033	170.392	62	61	
99 MN5	BIO-OPTICS			7/23/08 20:50	7/23/08 21:06	59.9033	170.392	59.9035	170.3913	60	61	
99 MN5	BIO-OPTICS			7/23/08 21:10	7/23/08 21:23	59.903	170.3918	59.9022	170.3923	63	64	
99 MN5	MCORE	1		7/23/08 21:38	7/23/08 21:41	59.9012	170.3945	59.9012	170.394	64	64	6 tubes rigged. 6 suitable samples. SOG=0.2kts with good 1654 lb pullout and 30cms penetration into soft clay/mud. Time/pos'n end = time/pos'n of core on the bottom.
99 MN5	MCORE	2		7/23/08 22:03	7/23/08 22:06	59.9018	170.3958	59.9017	170.3958	64	64	8 tubes rigged. 8 suitable samples. SOG=0.2kts with good 2569 lb pullout and 38cms penetration into soft clay/mud. Time/pos'n end = time/pos'n of core on the bottom.
100 70M39	MOORING	125	Standard	7/24/08 2:29	7/24/08 5:53	59.9077	171.7074	59.9077	171.7074	71	71	08-BSV-SA
100 70M39	CTD	125	Standard	7/24/08 2:40	7/24/08 2:50	59.909	171.7147	59.90718	171.7174	70	70	start CTD @ Mooring M5; end CTD
101 MN9	CTD	126	Standard	7/24/08 5:52	7/24/08 6:04	59.89777	172.8112	59.89769	172.8111	74	74	start; end
101 MN9	CTD	127	Standard	7/24/08 6:40	7/24/08 6:52	59.8982	172.8089	59.89868	172.8082	75	75	start; end
102 MN11	CTD	128	Standard	7/24/08 9:42	7/24/08 9:52	59.89822	174.0084	59.89781	174.0056	104	104	start; ctd 128; end; ctd 128
102 MN11	CALVET			7/24/08 9:59	7/24/08 10:06	59.89755	174.0033	59.89749	174.0003	100	100	
102 MN11	BONGO	22		7/24/08 10:08	7/24/08 10:18	59.8982	174.0092	59.8982	174.0092	101	101	Lots of Chrysaeora
102 MN11	CTD	129	Zoop	7/24/08 10:28	7/24/08 10:34	59.89873	173.9766	59.89849	173.9752	103	103	start; ctd 129; end; ctd 129
102 MN11	MOC			7/24/08 10:45	7/24/08 11:04	59.89944	173.9972	59.89946	173.9975	85	85	
102 MN11	MCORE	1		7/24/08 11:30	7/24/08 11:05	59.8994	173.9981	59.8996	173.9981	104	104	6 tubes rigged. 4 suitable samples. SOG=0.1kts with good 1857 lb pullout and 20cms penetration into silty clay. Time/pos'n end = time/pos'n of core on the bottom.
102 MN11	MCORE	2		7/24/08 11:53	7/24/08 11:58	59.9005	173.9984	59.9006	173.9983	102	102	the bottom.
103 MN12	CTD	131	Fe	7/24/08 14:00	7/24/08 14:50	59.8967	174.5933	59.8967	174.5933	109	111	Done in conjunction with CTD 131
103 MN12	CTD	130	Standard	7/24/08 15:06	7/24/08 15:21	59.89434	174.5836	59.89403	174.5818	114	114	start; ctd 130; end; ctd 130
103 MN12	CALVET			7/24/08 15:29	7/24/08 15:35	59.89378	174.5808	59.89346	174.5785	100	100	
103 MN12	CTD	131	Prod	7/24/08 15:59	7/24/08 16:11	59.89378	174.5751	59.89433	174.5736	110	110	start; ctd 131; end; ctd 131
103 MN12	BIO-OPTICS			7/24/08 16:15	7/24/08 16:28	59.8938	174.5725	59.8943	174.5723	110	108	
103 MN12	BIO-OPTICS			7/24/08 16:36	7/24/08 16:55	59.8935	174.5685	59.8942	174.5687	107	108	
104 MN13	CTD	132	Standard	7/24/08 18:37	7/24/08 18:53	59.90152	175.1928	59.9012	175.194	120	120	start; ctd 132; end; ctd 133
104 MN13	CALVET			7/24/08 19:00	7/24/08 19:08	59.90122	175.1943	59.90088	175.194	100	100	
105 MN14	CTD	133	Standard	7/24/08 20:51	7/24/08 21:09	59.89901	175.7969	59.89659	175.7932	132	132	start; end
105 MN14	CALVET			7/24/08 21:17	7/24/08 21:25	59.89553	175.7924	59.8956	175.7925	100	100	
106 MN15	CTD	134	Fe	7/24/08 23:15	7/25/08 0:10	59.9033	176.3917	59.9033	176.3917	145	141	Done in conjunction with CTD 134
106 MN15	CTD	134	Standard	7/25/08 0:08	7/25/08 0:28	59.90678	176.3715	59.90822	176.363	140	140	start; end
106 MN15	CALVET			7/25/08 0:36	7/25/08 0:44	59.90879	176.3594	59.90919	176.3561	100	100	
106 MN15	BIO-OPTICS			7/25/08 0:50	7/25/08 1:04	59.9092	176.3558	59.9113	176.3463	140	142	
106 MN15	BIO-OPTICS			7/25/08 1:10	7/25/08 1:25	59.9098	176.343	59.9108	176.3362	139	143	
106 MN15	BIO-OPTICS			7/25/08 1:30	7/25/08 1:45	59.911	176.3358	59.9113	176.3367	142	140	
106 MN15	MCORE	1		7/25/08 2:15	7/25/08 2:23	59.9028	176.3931	59.9027	176.3928	141	141	6 tubes rigged. 5 suitable samples. SOG=0.2kts with a starboard angle 2366 lb pullout and 25cms penetration into silty clay. Time/pos'n end = time/pos'n of core on the bottom.
106 MN15	MCORE	2		7/25/08 2:42	7/25/08 2:55	59.9025	176.3889	59.9057	176.3881	140	140	8 tubes rigged. 8 suitable samples. SOG=0.2kts with a vertical 2814 lb pullout and 21cms penetration into silty clay. Let core settle out 40 mab for 2 minutes. Time/pos'n end = time/pos'n of core on the bottom.
107 MN16	CTD	135	Standard	7/25/08 4:47	7/25/08 5:05	59.90092	176.9963	59.90294	176.9872	137	137	start; end
107 MN16	CALVET			7/25/08 5:12	7/25/08 5:16	59.90362	176.9836	59.90407	176.9807	100	100	
108 MN17	CTD	136	Fe	7/25/08 7:00	7/25/08 8:00	59.9117	177.58	59.9117	177.58	137	138	Done in conjunction with CTD 136
108 MN17	CTD	136	Standard	7/25/08 8:09	7/25/08 8:24	59.91773	177.5716	59.92162	177.565	138	138	start; ctd 136; end; ctd 136
108 MN17	CALVET			7/25/08 8:29	7/25/08 8:34	59.92303	177.5628	59.92444	177.5606	100	100	
109 MN18	CTD	137	Standard	7/25/08 10:24	7/25/08 10:41	59.89869	178.1952	59.89771	178.1844	143	143	start; ctd 137; end; ctd 137
109 MN18	CALVET			7/25/08 10:47	7/25/08 10:52	59.89739	178.1806	59.89726	178.1773	100	100	
110 MN19	BONGO	23		7/25/08 12:57	7/25/08 13:07	59.8975	178.8003	59.8975	178.8003	157	157	
110 MN19	MOC			7/25/08 13:08	7/25/08 13:58	59.8858	178.8088	59.87362	178.8344	100	100	

Station	Event Type	Cast#	CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
110 MN19	CTD	138	Zoop	7/25/08 13:18	7/25/08 15:43	59.88924	178.805	59.89476	178.8097	171	171	start; ctd 138; end; ctd 138
110 MN19	CTD	139	Fe	7/25/08 14:15	7/25/08 15:15	59.895	178.81	59.895	178.81	167	169	in conjunction with CTD 139
110 MN19	CALVET			7/25/08 15:50	7/25/08 15:56	59.89262	178.8108	59.89261	178.8107	100	100	
110 MN19	CTD	139	Standard	7/25/08 16:16	7/25/08 16:20	59.89387	178.807			167	167	start; ctd 139; end
110 MN19	CTD	140	Prod	7/25/08 16:23	7/25/08 16:33	59.894	178.807	59.894	178.807	165	165	start; end
110 MN19	BIO-OPTICS			7/25/08 16:45	7/25/08 16:52	59.8995	178.8062	59.8993	178.8063	161	159	
111 MN20	CTD	141	Fe	7/25/08 20:00	7/25/08 20:59	59.8967	179.3933	59.8967	179.3933	2705	2705	Done in conjunction with CTD 141
111 MN20	CTD	141	Standard	7/25/08 21:04	7/25/08 21:44	59.89454	179.3884	59.89402	179.3803	2718	2718	start; end
111 MN20	CALVET			7/25/08 21:58	7/25/08 22:09	59.89402	179.3797	59.90501	179.3471	100	100	
112 DL1	CTD	142	Standard	7/26/08 3:33	7/26/08 3:52	60.80879	177.9759	60.80934	177.9734	154	154	start; end
112 DL1	BIO-OPTICS			7/26/08 4:00	7/26/08 4:15	60.812	177.9687	60.8117	177.9692	151	149	
112 DL1	BIO-OPTICS			7/26/08 4:20	7/26/08 4:42	60.8115	177.9677	60.81	177.9683	149	147	
113 DL2	CTD	143	Standard	7/26/08 8:14	7/26/08 8:25	61.4853	176.998	61.48532	176.9958	120	120	start; ctd 143; end; ctd 143
114 SL16	CTD	144	Standard	7/26/08 12:10	7/26/08 12:20	62.20081	175.9662	62.2001	175.9643	94	94	start; ctd 144; end; ctd 144
114 SL16	CALVET			7/26/08 12:26	7/26/08 12:30	62.19962	175.9632	62.19919	175.9628	93	93	
114 SL16	BONGO	24		7/26/08 12:34	7/27/08 12:44	62.2005	175.9653	62.2005	175.9655	94	94	
114 SL16	CTD	145	Zoop	7/26/08 12:53	7/26/08 12:58	62.19515	175.9815	62.19468	175.9816	94	94	start; ctd 145; end; ctd 145
115 SL15	CTD	146	Standard	7/26/08 14:17	7/26/08 14:29	62.19558	175.5603	62.19342	175.5609	85	85	start; ctd 146; end; ctd 146
115 SL15	CALVET			7/26/08 14:36	7/26/08 14:41	62.19202	175.5612	62.19105	175.5618	86	86	
116 SL14	CTD	147	Prod	7/26/08 16:02	7/26/08 16:18	62.19967	175.1612	62.19651	175.1633	80	80	start; ctd 147; end; ctd 147
116 SL14	CALVET			7/26/08 16:25	7/26/08 16:31	62.19534	175.1642	62.19312	175.1663	80	80	
116 SL14	BIO-OPTICS			7/26/08 16:45	7/26/08 17:00	62.186	175.1808	62.1932	175.1815	79	80	
116 SL14	CTD	148	Standard	7/26/08 17:28	7/26/08 17:40	62.18567	175.1743	62.18447	175.1761	80	80	start; ctd 148; end; ctd 148
116 SL14	MCORE	1		7/26/08 18:06	7/26/08 18:10	62.1928	175.1763	62.1927	175.177	80	80	time/pos'n of core on the bottom. 8 tubes rigged. 8 suitable samples. SOG=0.2kts with a vertical 1432 lb pullout and 2 dm (20cm) penetration into silty clay / mud. Time/pos'n end =
116 SL14	MCORE	2		7/26/08 18:24	7/26/08 18:28	62.1918	175.1792	62.1916	175.1797	80	80	time/pos'n of core on the bottom.
117 SL13	MOORING	149	Standard	7/26/08 20:39	7/26/08 20:39	62.2009	174.749	62.2009	174.749	70	70	08-BSV-8A
117 SL13	BIO-OPTICS			7/26/08 20:55	7/26/08 21:05	62.2058	174.7468	62.2062	174.7465	70	71	
117 SL13	BIO-OPTICS			7/26/08 21:10	7/26/08 21:21	66.1968	173.8998	66.1972	173.8992	63	65	
117 SL13	CALVET			7/26/08 21:25	7/26/08 21:32	62.21323	174.7502	62.21486	174.7512	71	71	
117 SL13	CTD	149	Standard	7/26/08 21:37	7/26/08 21:47	62.21583	174.7517	62.21814	174.7526	74	74	start; end
118 SL12	CTD	150	Standard	7/26/08 23:09	7/26/08 23:18	62.20105	174.343	62.20195	174.3391	69	69	start; end
118 SL12	CALVET			7/26/08 23:27	7/26/08 23:33	62.20284	174.3359	62.20335	174.3334	68	68	
119 SL11	CTD	151	Standard	7/27/08 1:09	7/27/08 1:18	62.19695	173.918	62.19715	173.9109	64	64	start; end
119 SL11	CALVET			7/27/08 1:24	7/27/08 1:31	62.19713	173.9059	62.19701	173.9008	62	62	
119 SL11	BIO-OPTICS			7/27/08 1:35	7/27/08 1:45	62.1969	173.8998	62.1969	173.8992	64	64	
119 SL11	BIO-OPTICS			7/27/08 1:50	7/27/08 2:00	62.1962	173.8865	62.1963	173.8912	64	64	
120 SL10	CTD	152	Standard	7/27/08 3:02	7/27/08 3:11	62.19728	173.5332	62.19587	173.5271	63	63	start; end
120 SL10	CALVET			7/27/08 3:15	7/27/08 3:20	62.19522	173.5242	62.19451	173.5206	61	61	
121 SL9	CTD	153	Standard	7/27/08 4:32	7/27/08 4:40	62.19741	173.1182	62.19642	173.116	60	60	start; end
121 SL9	CALVET			7/27/08 4:45	7/27/08 4:49	62.19582	173.1153	62.19562	173.113	64	64	
122 SL8	CTD	154	Standard	7/27/08 6:04	7/27/08 6:13	62.19905	172.7058	62.19783	172.7044	60	60	start; end
122 SL8	CALVET			7/27/08 6:18	7/27/08 6:22	62.19667	172.7046	62.19605	172.7047	55	55	
123 70M56	CTD	155	Standard	7/27/08 11:00	7/27/08 11:12	61.94021	174.3643	61.94088	174.3644	74	74	start; ctd 155; end; ctd 155
123 70M56	CALVET			7/27/08 11:18	7/27/08 11:22	61.94129	174.3645	61.9416	174.3638	69	69	
123 70M56	BONGO	25		7/27/08 11:40	7/27/08 11:50	61.9402	174.3643	61.9402	174.3643	74	74	Cast time is approximate
124 70M55	CTD	156	Standard	7/27/08 12:50	7/27/08 12:55	61.85906	174.102	61.85889	174.0978	75	75	start; ctd 156; end; ctd 156
125 70M54	CTD	157	Standard	7/27/08 14:13	7/27/08 14:22	61.72559	173.8498	61.72404	173.8432	72	72	start; ctd 157; end; ctd 157
126 70M53	CTD	158	Standard	7/27/08 15:34	7/27/08 15:44	61.56097	173.7108	61.55877	173.7053	73	73	start; ctd 158; end; ctd 158
126 70M53	CALVET			7/27/08 15:51	7/27/08 15:56	61.55682	173.7013	61.55564	173.6989	69	69	
126 70M53	CTD	159	Prod	7/27/08 16:07	7/27/08 16:24	61.55305	173.6927	61.54904	173.6833	73	73	start; ctd 159; end; ctd 159
126 70M53	BIO-OPTICS			7/27/08 16:32	7/27/08 16:48	61.5468	173.679	61.547	173.6792	73	73	from ship log
127 70M52	CTD	160	Standard	7/27/08 18:07	7/27/08 18:10	61.41036	173.7372	61.41018	173.7371	73	73	start; ctd 160; end; ctd 160
128 70M51	CTD	161	Standard	7/27/08 19:37	7/27/08 19:45	61.24712	173.726	61.24595	173.7231	73	73	start; ctd 161; end; ctd 161
129 70M50	CTD	162	Standard	7/27/08 20:59	7/27/08 21:12	61.07976	173.7745	61.07772	173.7703	77	77	start; end
129 70M50	CALVET			7/27/08 21:19	7/27/08 21:25	61.07663	173.7683	61.07548	173.7657	78	78	
129 70M50	BIO-OPTICS			7/27/08 21:34	7/27/08 21:54	61.0742	173.7627	61.0733	173.7623	70	70	
130 70M49	CTD	163	Standard	7/27/08 23:20	7/27/08 23:32	60.91139	173.8208	60.91329	173.8144	80	80	start; end
131 70M48	CTD	164	Standard	7/28/08 1:04	7/28/08 1:14	60.75782	173.6543	60.75747	173.6462	72	72	start; end
132 70M47	CTD	165	Standard	7/28/08 2:47	7/28/08 2:57	60.58242	173.6637	60.58261	173.6591	70	70	start; end
132 70M47	CALVET			7/28/08 3:03	7/28/08 3:15	60.58236	173.6563	60.58158	173.65	69	69	
133 70M46	CTD	166	Standard	7/28/08 4:17	7/28/08 4:26	60.41767	173.5925	60.41653	173.5863	64	64	start; end
134 70M45	CTD	167	Standard	7/28/08 5:26	7/28/08 5:36	60.25488	173.523	60.25357	173.5175	69	69	start; end
135 70M44	CTD	168	Standard	7/28/08 6:38	7/28/08 6:48	60.11542	173.3251	60.11288	173.323	70	70	start; end
135 70M44	CALVET			7/28/08 6:54	7/28/08 6:58	60.11214	173.3207	60.11181	173.3197	69	69	
136 70M43	CTD	169	Standard	7/28/08 8:04	7/28/08 8:11	60.0461	173.026	60.04477	173.0245	67	67	start; ctd 169; end; ctd 169
137 70M42	CTD	170	Standard	7/28/08 9:08	7/28/08 9:15	59.98245	172.7199	59.98161	172.7179	68	68	start; ctd 170; end; ctd 170
138 70M41	CTD	171	Standard	7/28/08 10:26	7/28/08 10:34	59.90173	172.4213	59.90153	172.4197	74	74	start; ctd 171; end; ctd 171
139 70M40	CTD	172	Standard	7/28/08 11:45	7/28/08 11:53	59.84523	172.0934	59.84521	172.0904	75	75	start; ctd 172; end; ctd 172
140 70M39	CTD	173	Standard	7/28/08 12:44	7/28/08 12:52	59.83067	171.7743	59.82996	171.7713	74	74	start; ctd 173; end; ctd 173
141 70M38	CTD	174	Standard	7/28/08 13:45	7/28/08 13:54	59.77457	171.457	59.77396	171.4541	73	73	start; ctd 174; end; ctd 174
142 70M37	CTD	175	Standard	7/28/08 14:46	7/28/08 14:54	59.71879	171.1457	59.71807	171.1441	72	72	start; ctd 175; end; ctd 175
143 70M36	CTD	176	Prod	7/28/08 15:46	7/28/08 16:03	59.59962	170.9162	59.59779	170.9144	70	70	start; ctd 176; end; ctd 176
143 70M36	BIO-OPTICS			7/28/08 16:12	7/28/08 16:32	59.5948	170.913	59.5948	170.913	70	70	
144 70M35	CTD	177	Standard	7/28/08 17:19	7/28/08 17:29	59.43689	170.8788	59.43597	170.8798	72	72	start; ctd 177; end; ctd 177
145 70M34	CTD	178	Standard	7/28/08 18:18	7/28/08 18:27	59.33101	170.666	59.33177	170.667	70	70	start; ctd 178; end; ctd 178
146 70M33	CTD	179	Standard	7/28/08 19:21	7/28/08 19:29	59.25999	170.3815	59.26033	170.3812	69	69	start; ctd 179; end; ctd 179
147 70M32	CTD	180	Standard	7/28/08 20:27	7/28/08 20:35	59.11232	170.237	59.11232	170.237	66	66	start; end
148 70M31	CTD	181	Standard	7/28/08 21:40	7/28/08 21:50	58.9517	170.3234	58.95328	170.3214	70	70	start; end
149 70M30	CTD	182	Standard	7/28/08 22:56	7/28/08 23:05	58.7849	170.289	58.78682	170.2891	70	70	start; end
149 70M30	BIO-OPTICS			7/28/08 23:08	7/28/08 23:18	58.7928	170.287	58.7928	170.2873	71	71	
149 70M30	BIO-OPTICS			7/28/08 23:20	7/28/08 23:32	58.7927	170.2872	58.7935	170.2873	71	71	
150 70M29	CTD	183	Standard	7/29/08 0:51	7/29/08 1:00	58.61178	170.2					

Station Stn.# Name	Event Type	Cast# CTD Type	Time in Water (UTC)	Time on Deck (UTC)	Start Lat.	Start Long.	End Lat.	End Long.	Start Depth (m)	End Depth (m)	Notes
153 70M26	CTD	186 Radon	7/29/08 5:10	7/29/08 5:25	58.146	169.9082	58.146	169.9082	72	72	Radon; standard cast; Plus triple O2 isotopes
154 70M25	CTD	187 Standard	7/29/08 6:39	7/29/08 6:46	58.04802	169.6468	58.04792	169.648	70	70	start; end
155 70M24	CTD	188 Standard	7/29/08 7:57	7/29/08 8:05	57.97522	169.3615	57.97466	169.3627	70	70	start; ctd 188; end; ctd 188
156 70M23	CTD	189 Standard	7/29/08 9:17	7/29/08 9:24	57.9102	169.0641	57.91053	169.0667	68	68	start; ctd 189; end; ctd 189
156 70M23	MOC		7/29/08 9:30	7/29/08 10:02	57.9083	169.0708	57.8996	169.0443	55	55	
157 70M22	CTD	190 Standard	7/29/08 10:48	7/29/08 10:56	57.84367	168.893	57.84394	168.893	71	71	start; ctd 190; end; ctd 190
157 70M22	MOC		7/29/08 11:01	7/29/08 11:28	57.84508	168.8898	57.83142	168.8834	55	55	
158 70M20	CTD	191 Standard	7/29/08 12:49	7/29/08 12:58	57.63033	168.8139	57.63144	168.8129	70	70	start; ctd 191; end; ctd 191
158 70M20	MOC		7/29/08 13:03	7/29/08 13:34	57.63339	168.8104	57.63374	168.7745	55	55	
159 70M19	CTD	192 Standard	7/29/08 14:38	7/29/08 14:47	57.52684	168.6108	57.52511	168.6078	71	71	start; ctd 192; end; ctd 192
160 70M18	CTD	193 Standard	7/29/08 16:02	7/29/08 16:12	57.49959	168.3034	57.49854	168.3043	70	70	start; ctd 193; end; ctd 193
161 70M17	CTD	194 Standard	7/29/08 17:59	7/29/08 18:11	57.49982	167.988	57.49904	167.9941	71	71	start; ctd 194; end; ctd 194
162 70M16	CTD	195 Standard	7/29/08 19:37	7/29/08 19:50	57.50119	167.6708	57.50057	167.6765	71	71	start; end
163 70M15	CTD	196 Standard	7/29/08 21:11	7/29/08 21:22	57.50159	167.3508	57.50148	167.3549	70	70	start; end
164 70M14	CTD	197 Standard	7/29/08 22:40	7/29/08 22:53	57.52459	167.0425	57.5258	167.045	70	70	start; end
164 70M14	BIO-OPTICS		7/29/08 23:00	7/29/08 23:25	57.5263	167.0462	57.5265	167.0453	69	69	
165 70M13	CTD	198 Radon	7/30/08 0:44	7/30/08 0:55	57.4279	166.8021	57.4279	166.8021	70	70	Radon and triple O2 isotopes collected
165 70M13	CTD	198 Standard	7/30/08 0:44	7/30/08 0:59	57.42788	166.8021	57.42966	166.8037	70	70	from standard cast bottles
166 70M12	CTD	199 Standard	7/30/08 2:03	7/30/08 2:14	57.44466	166.5239	57.4444	166.5234	69	69	start; end
167 70M11	CTD	200 Standard	7/30/08 3:25	7/30/08 3:34	57.31634	166.3303	57.31593	166.3306	71	71	start; end
168 70M10	CTD	201 Standard	7/30/08 4:47	7/30/08 4:57	57.32066	166.0056	57.31907	166.004	71	71	start; end
169 70M9	CTD	202 Standard	7/30/08 5:59	7/30/08 6:09	57.26294	165.7405	57.26116	165.7394	70	70	start; end
170 70M8	CTD	203 Standard	7/30/08 7:12	7/30/08 7:21	57.11079	165.6044	57.11013	165.6041	70	70	start; end
171 70M7	CTD	204 Standard	7/30/08 8:34	7/30/08 8:42	56.99666	165.3775	56.9958	165.3773	72	72	start; ctd 204; end; ctd 204
172 70M6	CTD	205 Standard	7/30/08 9:59	7/30/08 10:09	56.89339	165.1366	56.89239	165.1362	72	72	start; ctd 205; end; ctd 205
172 70M6	MOC		7/30/08 10:44	7/30/08 10:44			56.89852	165.1084	55	55	
173 70M5	CTD	206 Standard	7/30/08 11:43	7/30/08 11:52	56.91044	164.8363	56.91124	164.8363	73	73	start; ctd 206; end; ctd 206
173 70M5	MOC		7/30/08 11:56	7/30/08 12:15	56.9118	164.8361	56.90996	164.8185	55	55	
174 70M4	CTD	207 Standard	7/30/08 13:17	7/30/08 13:25	56.85241	164.5709	56.85302	164.5712	73	73	start; ctd 207; end; ctd 207
174 70M4	MOC		7/30/08 13:29	7/30/08 13:53	56.85418	164.5716	56.85155	164.5482	55	55	
175 70M3	CTD	208 Standard	7/30/08 14:47	7/30/08 14:56	56.83136	164.3087	56.83221	164.3108	73	73	start; ctd 208; end; ctd 208
175 70M3	CALVET		7/30/08 15:08	7/30/08 15:12	56.8321	164.3117	56.83198	164.3118	73	73	
176 70M2	CTD	209 Standard	7/30/08 16:10	7/30/08 16:20	56.85189	164.0647	56.85079	164.0653	74	74	start; ctd 209; end; ctd 209
176 70M2	CALVET		7/30/08 16:28	7/30/08 16:35	56.84925	164.0662	56.84925	164.0662	69	69	
176 70M2	BIO-OPTICS		7/30/08 16:40	7/30/08 16:55	57.42	164.1	57.4183	164.0833	69	69	
177 70M1	CTD	210 Standard	7/30/08 17:54	7/30/08 18:00	56.76693	163.9595	56.76677	163.9596	74	74	start; ctd 210; end; ctd 210
177 70M1	CALVET		7/30/08 18:08	7/30/08 18:18	56.76489	163.9609	56.76489	163.9609	69	69	
177 70M1	BIO-OPTICS		7/30/08 18:20	7/30/08 18:40	56.7568	163.9723	56.7572	163.9733	73	73	

Appendix 2 – Details for sampling and deployment procedures
CTD Rosette water budget.

Priority/Order for CTD Rosette Water Sampling – HLY0803. Circulate clockwise around the rosette, beginning at the deepest Niskin bottle (No. 1).

Priority/ Order	Sampling	Sampler(s)	Sample Vol. /Niskin	Depths	How Often?	Notes
1	Radon	Shull/ Brusseau	20 L	A few		Separate casts
1	Triple oxygen Isotopes	Moran	0.5 L	A few		
2	O ₂ (Winkler)	Calvin Mordy (NOAA/PMEL)	0.7 L (1.4L total for dup.)		Every station	Dup. At 2 depths
2	DIC & Alkalinity	PMEL	1.5 L	6 depths	~1/3 of stations	
2.5	$\delta^{18}\text{O}$	Weingartner, and Others	50ml	3	Selected-all	
3	N Production	Sambrotto/ Burdloff	5-8 L	7	daily	Prod. casts
	C production	Casey/ Tiahlo	1-2 L	7	daily	Prod. casts
	Micro-zooplankton	Stoecker	5 L	10	Half of stns.?	Prod. casts et al.
3	Particulates/ DON/	Sambrotto/ Burdloff	4 L	10	~1/3 of stations	
3	zooplankton	Taylor/ Pleuthner	30L	Chl Max	Every station	Separate casts
3	Phyto. fluorescent properties/ ALFS	Goes and others	0.5			
3	Total Chl	PMEL	1.5L	6	Every station	
3	Fractionated Chl	PMEL	1.5L	6	~1/3 station	
4	$\delta^{15}\text{N}$ -NO ₃	Sambrotto/ Burdloff	0.125 L	All	~12 stns	
4	$\delta^{15}\text{N}$ -DON	Sambrotto/ Burdloff	0.125 L	All	~12 stns	
4	Nutrients	Calvin Mordy (NOAA/PMEL)	0.1 L	All	All	
4	Salinity	SIO	0.6 L	selected	All	

Niskin bottle positions in various water depths

Niskin Bottle no.	Niskin depth	Niskin depth	Niskin depth
1	0	0	0
2	10	10	10
3	20	20	20
4	30	30	30
5	40	40	40
6	50 or 60	50 or 60	50 or 60
7	bottom	75	75
8	chl max	100	100
9		125	150
10		150	200
11		bottom	bottom
12		chl max	chl max

Invitation for educational outreach at Marine Science Camp on St. George Island



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"Serving Coastal Alaska Since 1988"

Ray Sambrotto, Chief Scientist
BEST/BSIERP July 2008 Cruise
By e-mail: sambrott@ldeo.columbia.edu

13 June 2008

Dear Ray:

It's great to hear that R/V Healy will be working in the vicinity of the Pribilof Islands this summer. As we discussed, I will be conducting an intensive marine science camp on St. George Island in July for students from both islands. This is an outgrowth of a short Ocean Science Days program I led with Tribal government and school leaders in April 2008. Students were very inspired and so enthusiastic to pursue marine science studies that their communities, school district and tribal entities pooled resources to hold a science camp this summer. A brief description of the program is attached at the end of this memo.

The students have exhibited deep interest in exploring marine species and ecosystems in the Bering Sea broadly and near their islands. They are also intrigued by research technology being used to examine the marine ecosystem, which they know from an island perspective.

If there is an opportunity for these students to join researchers on the R/V Healy during even a brief working cruise in July, it would be tremendously valuable for them in many ways: exposure to technology, engagement in discussion of ongoing research with scientists from outside, collection of data and or specimens from deeper waters further offshore from the near shore realm in which we will be working during camp.

Please contact me at any time if you wish to discuss a student mini research expedition aboard the R/V Healy. The students will be thrilled to have such an experience, so we will remain open to any means to make this happen for them.

Michelle Ridgway, Director
Pribilof Marine Science Camp

Pribilof Island Marine Science Camp ~

Discovering Wonders of the Bering Sea Ecosystem

From July 8-18, 2008, twenty Aleut students from the Pribilof School District will work together in an intensive marine science camp program. Half of the team will travel from St. Paul Island to join the St. George Island students for field and laboratory research focused entirely on the sea surrounding their island homes. Research projects will build upon king crab ecology, plankton and oceanographic studies initiated in the spring Ocean Science Days program.

Highlights of the summer program include these research topics: marine fish foodwebs; king crab ecology II, oceanographic processes around the Pribilof Islands and nearby shelfbreak, plankton communities, effects of shore ice on intertidal communities, nearshore benthic habitat suitability for king crab.

Students will use GPS to map marine habitats, an ROV to investigate nearshore subtidal habitats, CTD for nearshore pelagic habitat characterization, crab larvae catchment units, plankton nets, van veen grabs, pots and a trawl sampling net to examine species and habitats of interest.